



Pharmacognostic Study of Some Phytochemically Unexploited Floristic Trees of Hathwa, Gopalganj (Bihar)

Sarika Kumari¹ and ^{*Dr. Md.Sarfraz Ahmad²}

¹Department of Botany, Jai Prakash University, Bihar, India.

²Associate Professor, Department of Botany, Gopeshwar College, Hathwa, Gopalganj (Bihar)

Abstract: Pharmacognosy of Indian plants has been a perk for studies and medicinal researches. Hathwa has been blessed with Flora with extraordinary nutritional and medicinal benefits. Plants of *Moringa oleifera* (Lam.), *Holarhena antidyserterica* (Roth)Wall, *Erythrina indica* (Lam.) were found to be present at outskirts of Hathwa region of Gopalganj Bihar, They have been traditionally used in several disorders faced by human body but the stringent record and detailing is missing and this research was accomplished to relate their antimicrobial activities with their phytochemical entities. The plant extracts of bark was chosen because of its abundance presence and easy availability. The extracts were found positive for phenol, quinine, steroids, protein and amino acids The time-kill kinetics profile of extract of bark of *H. antidyserterica* (Roth) against the test organisms *K. pneumoniae*, *P. aeruginosa* and *V. cholerae* at the test concentrations of *Moringa oleifera* showed reduction in number of viable cells over the first 4, 8, and 12 hours *P. aeruginosa*, and *V. cholerae*, respectively, followed by a gradual rise up to the 10thh and 14thh for *K. pneumoniae* when compared to the control (organisms without antimicrobial agent).The time-kill kinetics profile of extract against the test organisms; at test concentrations studied showed reduction in number of viable cells over the first 2, 6, and 24 hours *K. pneumoniae*, *P. aeruginosa*, and *V. cholerae* respectively, followed by a gradual rise up to the 24thh for *P. aeruginosa* and *V. cholerae* and 12thh for *K. pneumoniae* when compared to the control. Therefore from our research it was concluded that *Moringa oleifera* (Lam.), *Holarhena antidyserterica* (Roth)Wall, *Erythrina indica* (Lam.) plants present at the outskirts of Hathwa region possess antimicrobial properties with important nutritional values. The aim of our research was to target some crucially important plants of Hathwa Gopalganj Bihar to study their antimicrobial potential, and this research targeted to explore the Phytoconstituents of those used plants which made them potent for antimicrobial activity.

Keywords: Phytochemical constituents, Time Kill Assay, Physicochemical analysis, Pharmacological activity.

*Corresponding Author

Dr. Md. Sarfaraz Ahmad , AssociateProfessor, Department of Botany, Gopeshwar College, Hathwa, Gopalganj (Bihar)



Received On 10 March 2020

Revised On 14 August 2020

Accepted On 27 August 2020

Published On 05 April 2021

Funding This research did not receive any specific grant from any funding agencies in the public, commercial or not for profit sectors.

Citation Sarika Kumari and Dr. Md. Sarfaraz Ahmad , Pharmacognostic Study of Some Phytochemically Unexploited Floristic Trees of Hathwa, Gopalganj (Bihar).(2021).Int. J. Life Sci. Pharma Res. 11(2), P79-84 <http://dx.doi.org/10.22376/ijpbs/lpr.2021.11.2.P79-84>

This article is under the CC BY- NC-ND Licence (<https://creativecommons.org/licenses/by-nc-nd/4.0/>)



Copyright @ International Journal of Life Science and Pharma Research, available at www.ijlpr.com

I. INTRODUCTION

The products derived from several herbs and plants, being a source of multifunctional curing agents and bioactive compounds, are relatively considered safe for consumption. According to the Food and Agriculture Organization's (FAO) report, about 70-80 % of the world's population, especially in developing countries, relies on herbal medicine to prevent and cure diseases¹ and about 25% of the synthesized drugs are manufactured from medicinal plants². Increased demand for food to tackle hunger and malnutrition problems has been pertinent in developing countries over the last few decades. In Asian and African countries, the vast majority of the population suffers from malnutrition because of the deficiency of essential nutrients in foods. *Moringa oleifera* Lam. (syn. *M. pterygosperma*) belongs to the family *Moringaceae*, commonly known as the 'drumstick' or 'horseradish' tree. It is an affordable and readily available source of major essential nutrients and nutraceuticals, and it has the potential to eradicate malnutrition³. The *Moringa* is often considered as important famine food because of its high resistance to drought and arid conditions owing to their tuberous roots.⁴ Almost each and every part of *Moringa* tree is useful for medicinal, functional food preparations, nutraceuticals, water purification, and biodiesel production; including roots, leaves, flowers, green pods, and seeds⁵. The other chosen Tree in this research is *Holarrhena pubescens* Wall. ex G. Don (Apocynaceae), commonly known as "Kutaja", is an important plant used in indigenous systems of medicine as remedy for bronchitis, hematuria, spermatorrhoea, epilepsy, asthma, piles, leprosy, eczema, diarrhea, fevers and jaundice^{6,7}. Various parts of *H. antidyserterica* (Roth.) Wall have been reported to possess antibacterial activity. The bark has been reported to possess astringent and antidiarrheal properties¹⁰. Leaves of the plant are used to cure scabies¹¹. *H. antidyserterica* (Roth.) Wall. is currently being used in the treatment of various disease conditions without standardization. However, a major constraint which has hindered the acceptance of alternative medicine is the lack of documentation and stringent quality control with this backdrop; it is of prime importance to make an effort towards standardization of the plant material. The process of standardization is achieved by stepwise pharmacognostic studies¹² which in turn help to identify and authenticate plant material. The standardization of a crude drug is an integral part of establishing its correct identity. Before any crude drug can be included in an herbal pharmacopoeia, pharmacognostic parameters and standards must be established. *Erythrina indica* (Lam.) is also one amongst the unstudied plants found in the regions of forest plains as well as some outskirts of Hathwa; Gopalganj. *Erythrina indica* (Lam.) belongs to the family of Fabaceae (Legume family). It possesses many useful entitlements including one said to destroy pathogenic parasites and relieve joint pain; the juice from the leaves is mixed with honey and ingested to treat tapeworm, roundworm and threadworm in India; women used to take this juice to stimulate lactation and menstruation; it is commonly mixed with castor oil to treat dysentery; a warm poultice of the leaves is applied externally to relieve rheumatic joints; and the bark is used as a laxative, diuretic, and expectorant. In a very few researches reported as antibacterial^{13,14} active against Cytotoxicity¹⁵. Analgesic and anti-inflammatory¹⁶. The aim of our research was to target some of those crucially important plants of Hathwa Gopalganj Bihar to study their antimicrobial potential, and this research targeted to explore the Phytoconstituents of

those used plants which made them potent for antimicrobial activity.

2. MATERIALS AND METHODS

2.1 Physicochemical and proximate analysis of tree bark

Moisture content was determined gravimetrically by drying the samples in an oven at 100°C to a constant weight. The dried barks from the above tree sample were subjected to other chemical analyses. Crude protein was determined in accordance with the Kjeldahl method (Method No. 978.04). Crude fat was determined in accordance with the Soxhlet extract method using petroleum ether as the extract agent (60-80°C) (Method No. 930.09)¹⁸. Ash content was assayed by incinerating the samples in a muffle furnace at 550°C (Method No. 930.05)¹⁹.

2.2 Extraction of Phytochemical Constituents from tree bark

About 500g air dried powder of the plant materials of *Erythrina indica* (Lam.), *Moringa oleifera* Lam., and *Holarrhena antidyserterica* (Roth.) Wall. was extracted successively with the following solvent in Soxhlet extractor, and identified as fractions 1-3 as shown below; n-hexane-Fraction-1, Ethyl acetate-Fraction-2, and Methanol fraction-3. The extracts were made in the concentrations of 70% (150 ml) of methanol used for 25 gm of plant powder, 100 percent 150 ml of n-hexane, 100% Ethyl acetate (150ml) for other plant powders. Every time before extracting with the next solvent, the plant material was dried in a hot air oven below 500°C. N-hexane, methanolic, and ethyl acetate extracts were concentrated by distilling off the solvent and then evaporating to dryness. The yield after evaporation was 5 gm after all the phenomena was worked out the extract obtained contained minimal amount of moisture, the extracts were subjected to qualitative tests for the identification of various phytoconstituents.

2.3 Phytochemical Screening of the extract

The extracts obtained from the above process were then carried out for the phytochemical analysis. The phytochemical analysis was carried out for different bioactive compound determination using different standard procedures. This analysis was carried out in the similar way for Ethyl acetate, n-hexane, methanol extracts. The Alkaloid test was done by Mayer's test. For the test 1 ml of plant extract was mixed with 3ml of ammonia solution and the mixture was allowed to stand for a few minutes. Then 10 ml of chloroform was added to the same tube. The tube was shaken and the content was filtered. The chloroform was evaporated by keeping the tubes in a water bath. Then 3ml of Mayer's reagent was added to the tube. The result interpretation is by the formation of cream colour precipitate indicating presence of alkaloids.²⁰ For the test of Amino acids by method mentioned by Humphrey et al.²¹ 1ml of plant extract was treated with 0.5ml of the Ninhydrin reagent (0.25%). The content was boiled for a few minutes. The result interpretation is by the formation of blue colour indicating presence of amino acid in the extracts. The flavonoid was determined as an Alkaline Reagent test. For this, 1ml of plant extract was treated with a dilute sodium hydroxide solution that leads to the appearance of intense

yellow colour which fades away after addition of a few drops of dilute acid. The result is interpreted as positive when the colour disappears indicating presence of flavonoids. This test was followed by a test of Glycosides Keller-Killiani method was used for determining the test result. In the test 1 ml of plant extract was treated with a few drops of glacial acetic acid and ferric chloride solution and this content was mixed thoroughly. Next concentrated sulphuric acid was added to the tube and the tube was observed for the formation of two layers that is lower reddish brown layer and upper acetic acid layer. The result interpretation is done after the upper layer turns bluish green indicating the presence of glycosides in the sample.²² The presence of phenols in the extract was assessed through Ferric chloride test. 1ml of plant extract was treated with 3-4 drops of ferric chloride solution. The positive result is indicated by the formation of a bluish black colour indicating the presence of phenols. Test for the presence of protein was carried out by Xanthoprotic test. In this test 1 ml plant extract was treated with few drops of concentrated nitric acid. Formation of yellow colour is interpreted as the positive result for the test. Further Steroid test was carried out Liebermann Burchard test. For this 1ml of plant extract was mixed with 10 ml of chloroform. Then equal volume of concentrated sulphuric acid was added to

the tubes. This leads to formation of two layers, the upper layer turns red and the sulphuric acid layer shows green fluorescence and this indicates the presence of steroids. Quinine was estimated by Virk et al.²³.

2.4 Time-Kill Kinetics Assay

Time-kill kinetics of methanol extracts of *Erythrina indica* (Lam.), *Moringa oleifera* Lam., and *Holarrhena antidysenterica* (Roth.) Wall. *volveacea* was carried out following the procedure described by Virk et al.¹⁰ From the methanolic extract three different concentrations 500 μ l 1000 μ l 1500 μ l was taken and an inoculum size of CFU/ml was added and incubated at 37°C. Aliquots of 1.0 ml of the medium were taken at time intervals of 0, 1, 2, 3, 4, 5, 6, 12, and 24 h, for bacteria, and inoculated aseptically into 20 ml nutrient agar and incubated at 37°C for 24 h. A control test was performed for the organisms without the extracts or reference antibiotic. The colony forming unit (CFU) of the organisms was determined. The procedure was performed in triplicate (three independent experiments) and a graph of the log CFU/ml was plotted against time.

3. RESULTS AND DISCUSSION

3.1 Preliminary phytochemical analysis of crude plant extracts of *Erythrina indica* (Lam.), *Moringa oleifera* (Lam.) and *Holarrhena antidysenterica* (Roth.) Wall.

Table I. Phytochemical study of plant extracts *H. antidysentrica*, *Erythrina indica* and *Moringa oleifera*

Plant Constituents	<i>H. antidysentrica</i> (Roth.) Wall			<i>Erythrina indica</i> (Lam.)			<i>Moringa oleifera</i> (Lam.)		
	Ethanol Extract	Petroleum Ether Extract	Aqueous Extract	Ethanol Extract	Petroleum Ether Extract	Aqueous Extract	Ethanol Extract	Petroleum Ether Extract	Aqueous Extract
Steroid	+	-	+	+	-	+	-	+	+
Phenol	-	-	+	-	+	-	+	-	+
Amino acid	+	-	-	+	+	+	-	-	-
Protein	-	+	+	+	-	+	-	+	-
Glycoside	+	-	+	-	+	-	+	-	+
Quinine	-	+	-	+	-	+	-	+	-
Triterpenoid	+	-	+	-	+	-	+	-	+

In the above table preliminary phytochemical evaluation performed for plant extracts of *H. antidysentrica* (Roth.) Wall, *Erythrina indica* (Lam.) and *Moringa oleifera* (Lam.) presence of Phytoconstituents were obtained in their different solvent extracts.

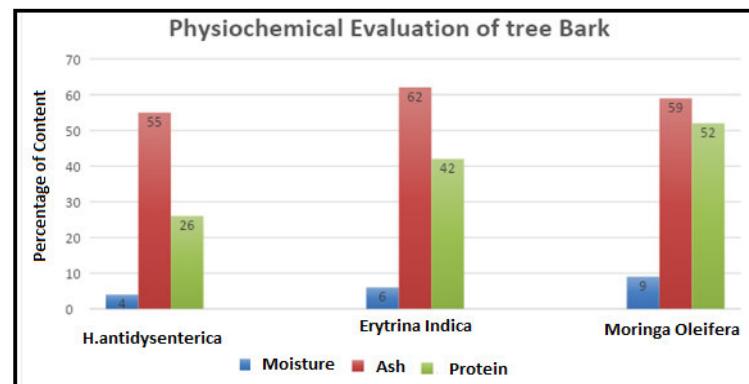


Figure 1: The above plot represents the Physicochemical evaluation of the Tree Bark, Ash content was found most in the bark of *Erythrina indica* Lam.; While Protein content was found to be highest in *Moringa oleifera* (Lam.) *H. antidysenterica* (Roth.) Wall. was found to be average in physicochemical Evaluation.

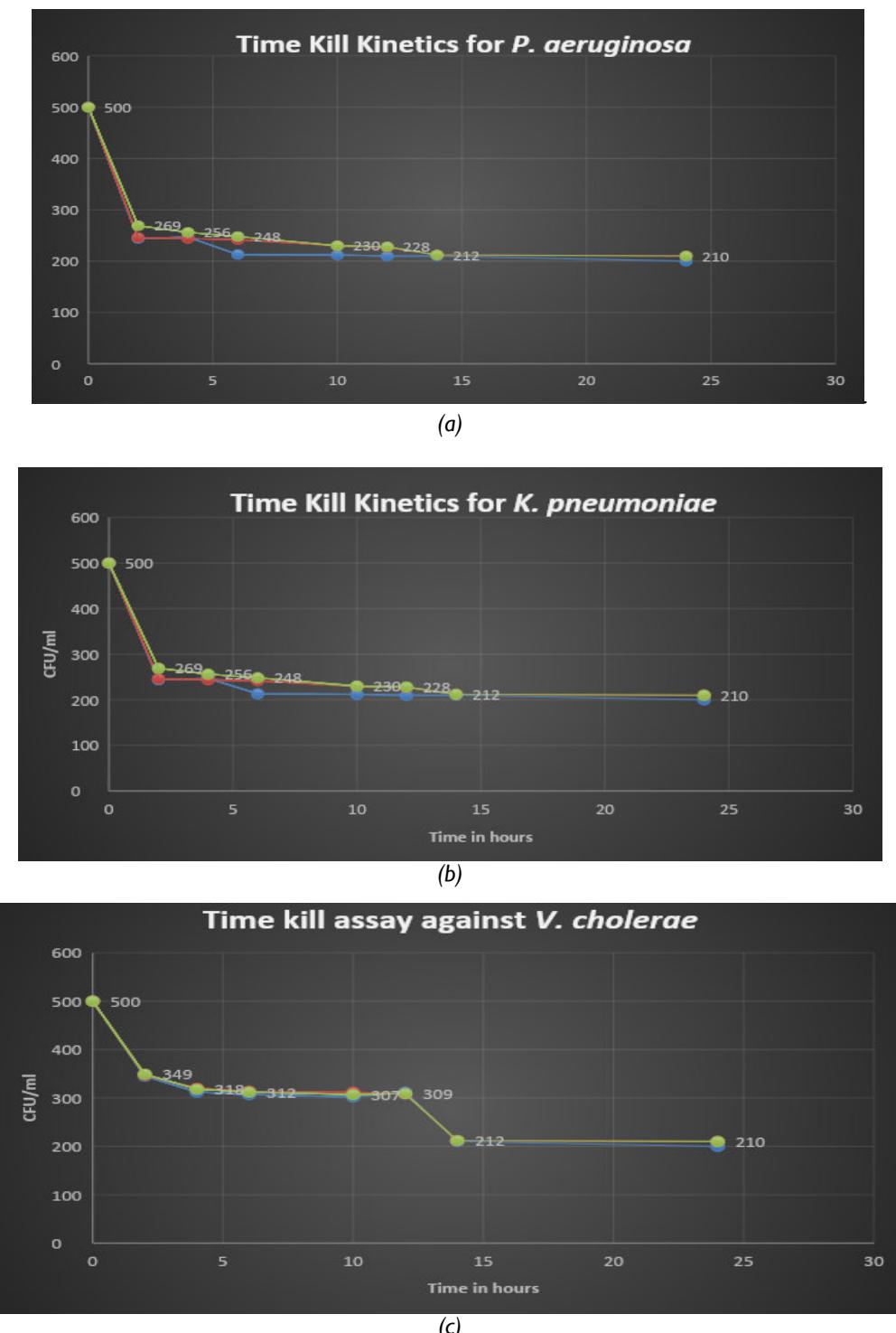


Figure 2 (a) , (b) and (c) Time kill Kinetics of Bark extract was represented slight fall of microbial colony when observed after the time gap 2, 4, 6, 8, 10, 12, 14, 24 hours against *V. cholerae*, *K. pneumoniae* and *P. aeruginosa*.

H. antidysentrica (Roth) Wall. is currently being used in the treatment of various disease conditions with out standardization. The standardization of a crude drug is an integral part of establishing its correct identity. Before any crude drug can be included in an herbal pharmacopoeia, pharmacognostic parameters and standards must be established. The extractive values are useful to evaluate the chemical constituents present in the crude drug and also help in estimation of specific constituents soluble in a particular solvent²⁵. The preliminary phytochemical evaluation (Table 1) revealed the presence of several secondary metabolites which are known to possess various pharmacological effects. In last four decades the scientists are keen to evaluate many

plant drugs used in medicinal folklore, due to their specific healing properties, health action and non-toxic effects.²⁶ In this dimension pharmacognostic study of *H. antidysentrica* is a substantial step and it further requires a long term study to evaluate pharmacological action as well as the therapeutic efficacy and toxicity of plant parts to establish as the drug. The pharmacognostic study of *H. antidy senterica* (Roth) Wall. has been carried out for the first time. This could also serve in the identification and preparation of a monograph on the plant. The plant *Moringaoleifera* family *Moringaceae* possess a broad spectrum of pharmacological activities. Also, most of the parts of plants like seeds, leaves, flowers and roots are used for treatment of various diseases. Literature reports

that aqueous, ethanolic and methanolic extracts are widely used for investigation, identification, and estimation purposes. In future the active constituents can be isolated and formulated into suitable dosage form and delivery system. Also, in future in-vivo studies based on animal models can be done for better effect²⁷. In this research *Moringa oleifera* (Lam.) was found to possess several various phytochemical entities like phenol in petroleum ether extract, Steroid in ethanolic extract, protein in aqueous extract that could denote that the extraction were efficient and targeted phytochemical entities were found present. The time-kill kinetics profile of extract of bark of *H. antidysentrica* (Roth) Wall. against the test organisms *K. pneumoniae*, *P. aeruginosa* and *V. cholerae* at test concentrations of *Moringa Oleifera* showed reduction in number of viable cells over the first 4, 8, and 12 hours *P. aeruginosa*, and *V. cholerae*, respectively, followed by a gradual rise up to the 10th h and 14th h for *K. pneumoniae* for when compared to the control (organisms without antimicrobial agent) (Figure 2b). The time-kill kinetics profile of extract against the test organisms; at test concentrations studied showed reduction in number of viable cells over the first 2, 6, and 24 hours *K. pneumoniae*, *P. aeruginosa*, and *V. cholerae* respectively, followed by a gradual rise up to the 24th h for *P. aeruginosa* and *V. cholerae* and 12th h (Figure 2 a,b,c) for *K. pneumoniae* when compared to the control²⁸.

4. CONCLUSION

It is noteworthy that these trees which are in abundant in

8. REFERENCES

1. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Front Pharmacol.* 2014;4:177. doi: 10.3389/fphar.2013.00177, PMID 24454289.
2. Pan SY, Zhou SF, Gao SH, Yu ZL, Zhang SF, Tang MK, Sun JN, Ma DL, Han YF, Fong WF, Ko KM. New perspectives on how to discover drugs from herbal medicines: CAM's outstanding contribution to modern therapeutics. *Evid Based Complement Alternat Med.* 2013;2013:627375. doi: 10.1155/2013/627375, PMID 23634172.
3. Kunyanga CN, Imungi JK, Vellingiri V. Nutritional evaluation of indigenous foods with potential food-based solution to alleviate hunger and malnutrition in Kenya. *J App Bioscience.* 2013;67(8):5277-88. doi: 10.4314/jab.v67i0.95049.
4. Padayachee B, Baijnath H. An overview of the medicinal importance of Moringaceae. *J Med Plants Res.* 2012;6(48):5831-9. doi: 10.5897/JMPR12.1187.
5. Saini SS, Bindslev-Jensen C, Maurer M, Grob JJ, Bülbül Baskan EB, Bradley MS, Canvin J, Rahmaoui A, Georgiou P, Alpan O, Spector S, Rosén K. Efficacy and safety of omalizumab in patients with chronic idiopathic/spontaneous urticaria who remain symptomatic on H1 antihistamines: a randomized, placebo-controlled study. *J Invest Dermatol.* 2015;135(1):67-75. doi: 10.1038/jid.2014.306, PMID 25046337.
6. Bhattacharjee SK. Handbook of medicinal plants. Aavishkar Publishers; 2000. p. 74.
7. Guha Bakshi DN, Sensarma P, Pal DC. Lexicon of medicinal plants in India. Naya Prakash. 199943-49.
8. Jolly CI, Mechery NR. Comparative pharmacognostical, physicochemical and antibacterial studies on seeds of *Holarrhena antidysenterica* Wall and *Wrightia tinctoria* R. Br. *Indian J Pharm Sci.* 1996;58(2):51.
9. Shrestha S, Subaramaihha SR, Subbaiah SG, Eshwarappa RS, Lakkappa DB. Evaluating the antimicrobial activity of methanolic extract of rhus succedanea leaf gall. *Bioimpacts.* 2013;3(4):195-8. doi: 10.5681/bi.2013.035. PMID 24455483.
10. Wankhede V, Waghmare SP, Pajai KS, Siddiqui MF, Hajare SW, et al. Efficacy of polyherbal antidiarrhoeal formulation in diarrhoeic goats. *J Pharmacogn Phytochem.* 2019;8(3):912-4.
11. Prajapati ND, Purohit SS, Sharma AK, Kumar T. A handbook of medicinal plants. Jodhpur; 1995. p. 875.
12. Shah U. *Cissus quadrangularis* L.: phytochemicals, traditional uses and pharmacological activities-a review. *Int J Pharm Pharm Sci.* 2011;3:41-4.
13. Amir F, Yam WS, Koay YC. Phytochemical constituents and biological activities of *Erythrina indica*. *Eur J Chem.* 2011;2(4):561-5. doi: 10.5155/eurjchem.2.4.561-565.455.
14. Kone WM, Solange KN, Dosso M. Assessing sub-Saharan erythrina for efficacy: traditional uses, biological activities and phytochemistry. *Pak J Biol Sci.* 2011 May 15;14(10):560-71. doi: 10.3923/pjbs.2011.560.571, PMID 22097091.
15. Amir F, Yam WS, Koay YC. Phytochemical constituents and biological activities of *Erythrina indica*. *Eur J Chem.* 2011;2(4):561-5. doi: 10.5155/eurjchem.2.4.561-565.455.

outskirts of Hathwa region of Gopalganj (Bihar) of plants used by the practitioners of this area could reach verge of disappearance have been found to have relevant pharmacological activities, which validate their traditional usages. Among the plants used by the local Ayurveda and Unani practitioners, the plant *Erythrina indica* (Lam.), *Moringa oleifera* Lam. and *Holarrhena antidysenterica* (Roth.) Wall. merit extensive scientific researches. The complete evaluation and record completion is mandatory therefore further analytic researches such as GC-MS and HPLC could make this work implicate and effective.

5. ACKNOWLEDGEMENT

The first author thanked the Department of Botany, Gopeshwar College Hathwa (Gopalganj) and Jai Prakash University, Chapra (Bihar) India for the identification and Collection of Plant species as well as guiding support and in writing and research.

6. AUTHORS CONTRIBUTION STATEMENT

In this research work all the Laboratory work was performed by Miss Sarika Kumari Under the Supervision Dr. Sarfaraz Ahmad and also he has helped in writing work. Further the data was brought to composition as research articles. All the authors have made discussions on methodology as well as results and compiled the manuscript.

7. CONFLICT OF CONFLICT

Conflict of interest declared none

16. Anonymous. The Wealth of India (A dictionary of Indian raw materials and industrial product). In: Gupta VK, editor. 3rd ed. New Delhi: National Institute of Science Communication and Council of Industrial and Scientific Research; 2007 87-9.
17. Horwitz W, Latimer G. AOAC-Association of official analytical chemists. Official Methods of Analysis of AOAC International. 18th ed, Gaithersburg, Maryland, USA. 2005;45:75-6.
18. Horwitz W, Latimer G. AOAC-Association of official analytical chemists. Official Methods of Analysis of AOAC International. 18th ed, Gaithersburg, Maryland, USA. 2005;45:75-6.
19. Horwitz W, Latimer G. AOAC-Association of official analytical chemists. Official Methods of Analysis of AOAC International. 18th ed, Gaithersburg, Maryland, USA. 2005;45:75-6.
20. Culvenor CCJ, Fitzgerald JS. A field method for alkaloid screening of plants. *J Pharm Sci*. 1963;52(3):303-4. doi: 10.1002/jps.2600520327.
21. Humphrey JM, Chamberlin AR. Chemical synthesis of natural product peptides: coupling methods for the incorporation of non coded amino acids into peptides. *Chem Rev*. 1997;97(6):2243-66. doi: 10.1021/cr950005s, PMID 11848900.
22. Sonibare MA, Soladoye MO, Esan OO, Sonibare OO. Phytochemical and antimicrobial studies of four species of *Cola* Schott & Endl. (Sterculiaceae). *Afr J Trad Complement Altern Med*. 2009;6(4):518-25. doi: 10.4314/ajtcam.v6i4.57182, PMID 20606771.
23. Virk JK, Kumar S, Singh R, Tripathi AC, Saraf SK, Gupta V, Bansal P. Isolation and characterization of quinine from *Polygonatum verticillatum*: A new marker approach to identify substitution and adulteration. *J Adv Pharm Technol Res*. 2016;7(4):153-8. doi: 10.4103/2231-4040.191427, PMID 27833896.
24. Tsuji BT, Yang JC, Forrest A, Kelchlin PA, Smith PF. In vitro pharmacodynamics of novel rifamycin ABI-0043 against *Staphylococcus aureus*. *J Antimicrob Chemother*. 2008 Jul 1;62(1):156-60. doi: 10.1093/jac/dkn133, PMID 18400804.
25. Ozarkar KR. Studies on anti-inflammatory effects of two herbs *Cissus quadrangularis* Linn. and *Valeriana wallichii* DC using mouse model [doctoral dissertation] [Ph.D. thesis]. Mumbai: University of Mumbai. p. 765; 2007.
26. Ganapathy PS, Ramachandra YL, Sudeep HV, Pavan KB, Achar KS, Padmalatha R. Pharmacognostic and phytochemical evaluation of *Holarrhena Antidysenterica* Wall. *Asian Aust J Plant Sci Biotechnol*. 2009;31(7):47-50.
27. Paikra BK, Dhongade HKJ, Gidwani B. Phytochemistry and pharmacology of *Moringa oleifera* Lam. *J Pharmacopuncture*. 2017;20(3):194-200. doi: 10.3831/KPI.2017.20.022, PMID 30087795.
28. Boakye PA, Olechowski C, Rashiq S, Verrier MJ, Kerr B, Witmans M, Baker G, Joyce A, Dick BD. A critical review of neurobiological factors involved in the interactions between chronic pain, depression, and sleep disruption. *Clin J Pain*. 2016;32(4):327-36. doi: 10.1097/AJP.0000000000000260, PMID 26035521.