



Platelet Rich Plasma Therapy: A Quick Note for Every Health Care Professional

Yedire Bharath Kumar¹, Hindustan Abdul Ahad^{*2}, Chinthaginjala Haranath², Gopavaram Sumanth²,
Durga Sumanth Pasupuleti² and Srilekha Surapa Reddy¹

¹Department of Pharmaceutics, Raghavendra Institute of Pharmaceutical Education and Research (RIPER), Ananthapuramu – 515721, Andhra Pradesh, India

²Department of Industrial Pharmacy, Raghavendra Institute of Pharmaceutical Education and Research (RIPER), Ananthapuramu – 515721, Andhra Pradesh, India.

Abstract: Blood is a fluid connective tissue consisting of cells and plasma. The plasma occupies 55% of blood, which is rich in immunoglobulins and proteins that have a wide range of applications in the medical field. Utilizing this plasma to tackle various diseases/disorders is called plasma therapy. Health care professionals require basic and quick knowledge of plasma therapy. It was attempted to bring a quick guide about plasma and its effective utilization in tackling deadly diseases. The plasma is being used in several issues like tissue regeneration, wound healing, scar revision, skin rejuvenating effects, alopecia, and now for the coronavirus disease (COVID-19). The Platelet Rich Plasma (PRP) has been used to heal wounds and illnesses. The theory behind PRP therapy is that it will induce the body to develop new, healthy cells that facilitate healing. Plasma contains important components like enzymes, antibodies, coagulation factors, albumin proteins, and fibrinogen. As PRP is rich in the proteins and antibodies, it is used for rare chronic therapies and many severe health problems. PRP therapy is gaining attraction by many health professionals as it is a safe, effective, efficient, and easy approach in procuring, preserving, and therapy. This review summarises and highlights the principle, techniques, method of preparation of PRP, convalescent plasma therapy, and plasma-derived therapies for patients with immunodeficiency or infected with infections to fight pathogens and get cured effectively in a short time.

Keywords: plasma, platelets, health, immunity, treatment

*Corresponding Author

Hindustan Abdul Ahad, Department of Industrial Pharmacy,
Raghavendra Institute of Pharmaceutical Education and
Research (RIPER), Ananthapuramu Andhra Pradesh, India



Received On 30 May 2020

Revised On 01 September 2020

Accepted On 17 September 2020

Published On 03 December 2020

Funding This research did not receive any specific grant from any funding agencies in the public, commercial or not for profit sectors.

Citation Yedire Bharath Kumar, Hindustan Abdul Ahad, Chinthaginjala Haranath, Gopavaram Sumanth, Durga Sumanth Pasupuleti, Srilekha Surapa Reddy, Platelet Rich Plasma Therapy: A quick note for every health care professional.(2020).Int. J. Life Sci. Pharma Res.10(5), P84-89 <http://dx.doi.org/10.22376/ijpbs/lpr.2020.10.5.P84-89>

This article is under the CC BY- NC-ND Licence (<https://creativecommons.org/licenses/by-nc-nd/4.0>)



Copyright © International Journal of Life Science and Pharma Research, available at www.ijlpr.com

1. INTRODUCTION

Plasma is the major fluid in the body, that takes up more than half (55%) of the total amount. Plasma is a bright yellow fluid and its key function is to carry foods, hormones, and proteins to certain areas of the body that require them.¹ Along with essential ingredients, plasma contains oxygen, salts, enzymes, anticorps, coagulation factors, albumin, and fibrinogen proteins.² Owing to these contents, plasma therapy will benefit patients recovering from fire burns, anesthesia, injuries, and other medical conditions and save their lives. For rare chronic therapies like Hemophilia and Autoimmune disorders, the antibodies and proteins in plasma are much helpful. People with AB blood groups have the highest demand for collecting plasma, as only 2 in 50 persons have this blood group.³ Plasma serves to disperse heat across the body and preserve homeostasis.⁴ Immune plasma refers to plasma obtained from organisms, following infection resolution and antibody production. Passive antibody therapy by transfusion of recovering plasma may avert clinical contagion or dull clinical harm in persons with fresh pathogen acquaintance.⁵ Antibody therapy may also be adopted for the treatment of patients with signs of varying severity. Passive antibody therapy, though, is more successful, it is performed prophylactically or utilized early after symptoms started. Passive antibody treatment was in operation for more than ten decades. The active mediators are antibodies in contradiction of interest-bearing target pathogen. Passive antibody therapy today trusts primarily on joint immunoglobulin provisions which contain high antibody concentrations. In comparison, plasma has been used in epidemics when there is less time or economic to generate preparations for immunoglobulins.⁶ There are many examples, both historical and modern, of which convalescent plasma was effectively used as a prophylaxis post-exposure (E.g., hepatitis, mumps, polio, measles, rabies) and/or diagnosis of a multitude of infectious diseases (E.g., influenza, Argentine hemorrhagic fever, SARS-CoV, MERS, Ebola, and even COVID-19). Platelet-rich plasma (PRP) is extracted from the blood by obtaining a drop of venous blood, inserting it in a special tube, and centrifuging the blood about 3000 rpm for 3 min in a centrifuge.⁷ This separates the whole blood into its elements, including Cells (Red blood cells [RBC]) and the non-cellular fluid (plasma). The intermediate layer is PRP, which comprises extremely enriched specialist chemicals named growth factors. These growth factors derived from platelets, transforming the β -growth factor, and growth factor for the vascular endothelial cells. Such factors communicate with the local cells and give signals triggering many events including cell division and migration. The fundamental concept behind PRP injection is to administer large doses of growth factors to an injured region, aiming to induce a curative reaction and reduce tissue inflammation. Whole blood injection can induce the same reaction to some degree but a smaller extent. PRP has been used in orthopaedic, and cosmetic surgeries to facilitate recovery.⁸ In the last 5 years, PRP has been recognized as having the ability to heal evaluating tendons, ligaments, and tissues in both recurrent and severe musculoskeletal injuries. The technique is attracting broad public coverage because it has been implemented with retired athletes as early as possible with efforts to restore them to action.

2. PLASMA THERAPY

The plasma therapy has further divided into the following varieties

2.1. PLATELET-RICH PLASMA THERAPY

PRP injections are used for treating broken tendons, tendinitis, muscle injuries, pain associated with arthritis, and joint injuries.⁸ Also, they are more common for cosmetic procedures. For example, dermatologists and hair-replacement specialists use PRP injections to treat a form of androgenic alopecia, which affects both men and women and some dermatologists treat skin-related issues on the face with PRP.

2.2. PREPARATION OF PRP

PRP is found from a sample of the blood of patients collected during diagnosis. A 30 cc venous blood can produce 3-5 cc of PRP based on an individual's baseline platelet count, the system used, and the procedure used.⁹ Blood drawing happens when anticoagulant (citrate dextrose A) is applied to avoid platelet activation preceding to its use. So, a specific tool called the table-top cold centrifuge is adopted.

2.3. PRINCIPLES AND PROCEDURES OF PRP PREPARATION

PRP is made by centrifugation, the acceleration force is modified based on different basic gravity to sediment those cellular constituents. There are plenty of ways to train for PRP. It can either be prepared using the PRP method or Buffy-coat process. Initial centrifugation to discrete red blood cells (RBC) is followed in the PRP cycle. Second centrifugation of the platelets is to concentrate, suspended in the smallest final. Initially, whole blood (WB) is stored in tubes containing anticoagulants.¹⁰ The first step is to isolate RBCs of the remaining WB volume are carried out at steady acceleration. Later the first rotation steps, the WB splits into three layers (the upper layer the intermediate thin layer [which contains mainly platelets and WBC], and the lower layer (consists of RBCs). The upper and intermediate layers are transferred to a vacuum sterile tube for pure production PRP (The P-PRP). The whole layer of intermediate coat and lower layer are transferred for the development of leukocyte rich PRP (L-PRP). The second step of the spin is then executed.¹¹ The second spin 'g' will be adequate to assist in the creation of soft pellets at the foot of the tube. It eliminates the upper side of the volume mainly composed of PPP (platelet-poor plasma). Pellets are homogenized to form the lower 1/3rd (5 ml) of PRP. A Buffy-coat contains a high leukocyte concentration. A very strong one Buffy coat thin layer may be formed from a small capacity of WB (10 ml). The challenge is to separate this thin buffy coat layer, which mainly contains white blood cells (WBCs) and platelets, from the underlying RBC layered by differential centrifuge.

2.4. PRP PREPARATION METHOD

2.4.1. ROUTINE CENTRIFUGATION METHOD

The important steps involved in the preparation of PRP preparation.¹²

- WB of acid citrate dextrose (ACD) in tubes are obtained by venipuncture
- Do not cool the blood before or after the separation of the platelets at any time.
- Blood centrifuge uses a 'soft' spin.
- Moves supernatant plasma with platelets (without anticoagulant) into another sterile tube.

- Centrifuge tube to receive a platelet concentrate at a higher rate (a quick spin).
- The downside 1/3rd is PRP, and the upside 2/3rd is platelet-poor plasma (PPP).
- Remove PPP and hang up the platelet pellets in a minimum plasma quantity (2-4 ml) by shaking the tube gently.

2.5. BUFFY COAT METHOD

This method needs the following steps.¹³

- WB should be processed, before centrifugation, at 20°C to 24°C.
- WB centrifuge at 'high' velocity.
- Due to its density, three layers are formed: the bottom layer composed of RBCs, the middle layer composed of platelets and WBCs, and the top layer of PPP.
- Supernatant plasma is removed from container tops.
- The Buffy-coat coating is moved to another sterile pipeline.
- To separate WBCs or use low-velocity centrifuge leukocyte filtration.

There is no such thing as Consensus as to whether or not to cause platelets before application, and which one agonist with. Some researcher's trigger thrombin or calcium platelets while others use platelets beforehand with no activation, suggesting better results received.

2.6. CONVALESCENT PLASMA THERAPY

Convalescent blood products (CBPs), taken from a patient who has survived an earlier infection and earned humoral immunity. Pathogen accountable for the disease by extracting whole blood or plasma is a potential source of specific human antibodies. CBP transfusion can counteract the pathogen and in time contributes to blood circulation eradication.¹⁴

- Convalescent whole blood (CWB), convalescent plasma (CP) or convalescent serum (CS)
- Collective human immunoglobulin (Ig) for IM or IV administration
- High-title patient Ig
- Monoclonal or polyclonal antibodies

CP has been the focus of growing concern, especially in the midst of epidemics of great scale. The apheresis plasma is currently the preferred therapeutic tool as a greater amount is obtained each session, the likelihood of more regular donations, and the lack of an effect on the donor's haemoglobin due to the reinfusion of their RBCs. Recruitment of donors living in areas where an epidemic has broken out may offer the added value of giving specific passive immunity opposite to the local infectious agent that has been artificially acquired, whereas CBP supplied from other regions may be less active due to (possible) strain differentiation of the pathogen concerned. Nonetheless, it can be difficult to classify, pick, and retain potential donors, as convalescent subjects must also follow donor selection requirements, in compliance with national policies and standard procedures. However, as indicated by the World Health Organization (WHO), due to the possible value of the treatment, certain donor selection criteria, while intended to protect the donor's health, maybe relaxed. Notably, the use of pathogen inactivation may guarantee additional protection

and therefore endorse less stringent requirements for selection.¹⁵

2.7. SCIENTIFIC UNCERTAINTIES AND LIMITATIONS REGARDING THE CONVALESCENT PLASMA USE

While its effectiveness and safety have not yet been thoroughly established, treatment with CP may be a viable choice in the treatment/prophylaxis of many infectious diseases, both in combination with other drugs/preventive measures and as the only therapy when there is no possible medication.¹⁶

2.8. DRAWBACKS OF CP TRANSFUSION

However, some problems still need to be discussed in evaluating the advisability of introducing a large-scale CP transfusion program.¹ The lack of high-quality research (i.e., randomized clinical trials) The risk of transmitting pathogens to transfusion service workers (E.g., handling laboratory specimens from contaminated pre-transfusion testing recipients) The need for advertising: Case-fatality levels in CP studies would be affected not only by the risk factors of patients but also by the particular supportive care offered by clinical centres. Immunotherapy with monoclonal antibodies may be highly effective. Many health care workers moved to Europe or the United States received CP and survived, but also benefited from experimental therapies and adequate supporting treatment. The risk of other transfusion-borne infections (E.g., human immunodeficiency virus, hepatitis B virus, Hepatitis C virus, and Syphilis) cannot be eliminated in endemic areas and pathogen control technologies can play a key role in ensuring safe transfusion of CPs.

2.9. PLASMA-DERIVED THERAPIES FOR IMMUNODEFICIENCY

Plasma-derived therapies replace missing or deficient proteins that allow people to lead healthier and more successful lives. Patients who rely on these therapies typically need daily infusions or injections during their entire lives. Plasma protein therapy treating diseases and disorders are called rare diseases because they affect a fairly small percentage of the population; most are chronic, inherited disorders.¹⁸

2.10. IG REPLACEMENT THERAPY

This therapy is a lifelong, life-saving procedure that needs to be performed periodically. It is important not to skip or neglect the treatment as each treatment only offers temporary protection against infection. Another kind of plasma-derived therapy is specifically used to treat hereditary angioedema (HAE), patients. For people living with this disease, there is a missing or malfunctioning part of the immune system called the C₁ esterase inhibitor (C₁-INH), and C₁-INH concentrate derived from plasma can be administered to avoid and cure the symptoms of HAE-related inflammation.¹⁹

2.11. ROUTES OF IG REPLACEMENT THERAPY

IG replacement therapy is either delivered subcutaneously (sc) or intravenously (iv) as an injection.²⁰

2.12. IV ROUTE

Infusion of IG straight through a vein into the bloodstream. The main benefit of using this route is that it is possible to prescribe a more dose of IG compared with the SC route and that care only has to be given every 3 to 4 weeks. This can be given as iv infusion which takes 2–4 h to administer, and needs hospitalization for its administration. This can also be provided at home by a nurse or a qualified carer. Slight side effects can happen during or after IV infusions.

2.13. SC ROUTE

Use either a handheld infusion pump (syringe driver) or rapid push technique, injection of IG is just below the skin of the upper arm, belly, thighs or buttocks. The rapid push technique is an easy way to move the IG under the skin using a syringe at a convenient pace.

2.14. SIDE EFFECTS OF IG THERAPY

Most patients with IG replacement therapy will not

experience significant side effects. Many patients may however report the following.²¹

- Cranks
- Lightheaded, tired or sick
- Fever
- Chills
- Feeling sick
- Itching
- Skin tingling
- Pains at the joint
- Quick heartbeats

These side effects are less frequent with the administration route of SC than the route of IV. Nonetheless, SC infusions at the site of injection may lead to some swelling and pain. Some side effects lead to slowing the rate of infusion and maintaining congenial hydration before and during treatment (intake of alcohol must be limited to prevent dehydration).

2.15. APPLICATIONS OF PRP

Plasma therapy is being successfully used for many years, few are highlighted in table I.

Table I. Milestones of plasma therapy for various diseases		
Year	Disorder/disease	References
1953	Primary immunodeficiency disease ²²	(22)
1970	Hemophilia A ²³	(23)
1970	Hemophilia B ²⁴	(24)
1980	Alpha-I antitrypsin deficiency ²⁵	(25)
1995	Kawasaki disease ²⁶	(26)
1996	Chronic inflammatory demyelinating neuropathy (CIDP) ²⁷	(27)
1996	Idiopathic Thrombocytopenic purpura (ITP) ²⁸	(28)
2005	Von willebrand disease ²⁹	(29)
2008	Hereditary Angioedema ³⁰	(30)
2009	Antithrombin III Deficiency ³¹	(31)
2009	Influenza virus ³²	(32)
2013	Cancer ³³	(33)
2014	Ebola virus ³⁴	(34)
2015	Pulmonary disease ³⁵	(35)
2019	Parkinson's disease ³⁶	(36)
2019	COVID-19 ³⁷	(37)

2.16. Latest updates on platelet rich plasma therapy for COVID

It was demonstrated clinic features and therapies of COVID-19 patients with PRP therapy and compared with traditional Chinese medicines³⁸. The assortment, manufacturing, pathogen inactivation, and backing, with a focus on COVID-19 was explained by Focosi *et al*³⁹. Skendros *et al* explored how complement relates with the platelet extracellular traps using COVID-19 specimens⁴⁰. Elizabeth *et al.*, explored that Neutrophil extracellular traps initiates from chromatin unconfined to immobilize pathogens and can trigger immunothrombosis in COVID-19 Acute Respiratory Distress Syndrome⁴¹. Xie *et al.*, reviewed 58 cases of COVID-19 pneumonia within 48 h of admission to the ICU can lessen the use of motorized aeration, curtail the hospital stay, indorse the early retrieval, and mend the actual handling of patients to attain noteworthy clinical usefulness⁴².

3. CONCLUSION

The information in this article will give quick information

about platelet-rich plasma (PRP), its composition, preparation, and its utilization in tackling infections. Platelet-rich plasma (PRP) therapy is a simple, economical, and feasible in the treatment of various clinical issues in the field of dermatology, dentistry, orthopedics, surgery, ophthalmology. Although experience and clinical data, dose are important factors to be optimized for such therapies. Additionally, PRP therapy is implemented to tackle influenza, SARS-CoV, MERS, Ebola, and even COVID-19. Even after having many reported positive effects of using PRP, many studies required to prove its safety and standardization of PRP.

4. AUTHORS CONTRIBUTION STATEMENT

All authors Yedire Bharath Kumar, Hindustan Abdul Ahad, Chinthaginjala Haranath, Gopavaram Sumanth, Durga Sumanth Pasupuleti and Srilekha Surapa Reddy, all authors involved in the collection, editing, and approved the final manuscript.

5. CONFLICT OF INTEREST

Conflict of interest declared none.

6. REFERENCES

1. Yerokhin AL, Nie X, Leyland A, Matthews A, Dowe S. Plasma electrolysis for surface engineering. *Surface and coatings technology*. 1999 Dec 15;122(2-3):73-93. doi: 10.1016/S0257-8972(99)00441-7
2. Misra NN, Pankaj SK, Segat A, Ishikawa K. Cold plasma interactions with enzymes in foods and model systems. *Trends in Food Science & Technology*. 2016 Sep 1; 55:39-47. doi: 10.1016/j.tifs.2016.07.001
3. Parsonnet J, Friedman GD, Orentreich N, Vogelmann H. Risk for gastric cancer in people with CagA positive or CagA negative *Helicobacter pylori* infection. *Gut*. 1997 Mar 1;40(3):297-301. doi: 10.1136/gut.40.3.297
4. Iwama GK, McGeer JC, Pawluk MP. The effects of five fish anaesthetics on acid-base balance, hematocrit, blood gases, cortisol, and adrenaline in rainbow trout. *Canadian journal of zoology*. 1989 Aug 1;67(8):2065-73. doi: 10.1139/z89-294
5. Previously PD. *Society of General Internal Medicine*. doi: 10.1046/j.1525-1497.17.s1.9.x
6. Isobe T, Osserman EF. Patterns of amyloidosis and their association with plasma-cell dyscrasia, monoclonal immunoglobulins and Bence-Jones proteins. *New England Journal of Medicine*. 1974 Feb 28;290(9):473-7. doi: 10.1056/NEJM197402282900902
7. Mei-Dan O, Lippi G, Sánchez M, Andia I, Maffulli N. Autologous platelet-rich plasma: a revolution in soft tissue sports injury management? *The Physician and sportsmedicine*. 2010 Dec 1;38(4):127-35. doi: 10.3810/psm.2010.12.1835
8. Sampson S, Gerhardt M, Mandelbaum B. Platelet rich plasma injection grafts for musculoskeletal injuries: a review. *Current reviews in musculoskeletal medicine*. 2008 Dec 1;1(3-4):165-74. doi: 10.1007/s12178-008-9032-5
9. Dhurat R, Sukesh MS. Principles and methods of preparation of platelet-rich plasma: a review and author's perspective. *Journal of cutaneous and aesthetic surgery*. 2014 Oct;7(4):189. doi: 10.4103/0974-2077.150734
10. Textor J. Autologous biologic treatment for equine musculoskeletal injuries: platelet-rich plasma and IL-1 receptor antagonist protein. *Veterinary Clinics: Equine Practice*. 2011 Aug 1;27(2):275-98. doi: 10.1016/j.cveq.2011.05.001
11. Shimojo AA, Perez AG, Galdames SE, Brissac IC, Santana MH. Stabilization of porous chitosan improves the performance of its association with platelet-rich plasma as a composite scaffold. *Materials Science and Engineering: C*. 2016 Mar 1; 60:538-46. doi: 10.1016/j.msec.2015.11.080
12. Perez AG, Lana JF, Rodrigues AA, Luzo AC, Belangero WD, Santana MH. Relevant aspects of centrifugation step in the preparation of platelet-rich plasma. *ISRN hematology*. 2014 Mar 25;2014. doi: 10.1155/2014/176060
13. Fijnheer R, Pietersz RN, De Korte D, Gouwerok CW, Dekker WJ, Reesink HW, Roos D. Platelet activation during preparation of platelet concentrates: a comparison of the platelet-rich plasma and the buffy coat methods. *Transfusion*. 1990 Sep;30(7):634-8. doi: 10.1046/j.1537-2995.1990.30790385523.x
14. Marano G, Vaglio S, Pupella S, Facco G, Catalano L, Liunbruno GM, Grazzini G. Convalescent plasma: new evidence for an old therapeutic tool? *Blood Transfusion*. 2016 Mar;14(2):152. doi: 10.2450/2015.0131-15
15. Sahr F, Ansumana R, Massaquoi TA, Idriss BR, Sesay FR, Lamin JM, Baker S, Nicol S, Conton B, Johnson W, Abiri OT. Evaluation of convalescent whole blood for treating Ebola Virus Disease in Freetown, Sierra Leone. *Journal of Infection*. 2017 Mar 1;74(3):302-9. doi: 10.1016/j.jinf.2016.11.009
16. Van Griensven J, Edwards T, de Lamballerie X, Semple MG, Gallian P, Baize S, Horby PW, Raoul H, Magassouba NF, Antierens A, Lomas C. Evaluation of convalescent plasma for Ebola virus disease in Guinea. *New England Journal of Medicine*. 2016 Jan 7;374(1):33-42. doi: 10.1056/NEJMoa1511812
17. Mora-Rillo M, Arsuaga M, Ramírez-Olivencia G, de la Calle F, Borobia AM, Sánchez-Seco P, Lago M, Figueira JC, Fernández-Puntero B, Viejo A, Negredo A. Acute respiratory distress syndrome after convalescent plasma use: treatment of a patient with Ebola virus disease contracted in Madrid, Spain. *The Lancet Respiratory Medicine*. 2015 Jul 1;3(7): 554-62. doi: 10.1016/S2213-2600(15)00180-0
18. Kaveri SV, Maddur MS, Hegde P, Lacroix-Desmazes S, Bayry J. Intravenous immunoglobulins in immunodeficiencies: more than mere replacement therapy. *Clinical & Experimental Immunology*. 2011 Jun; 164:2-5. doi: 10.1111/j.1365-2249.2011.04387.x
19. Chapel HM, Spickett GP, Ericson D, Engl W, Eibl MM, Bjorkander J. The comparison of the efficacy and safety of intravenous versus subcutaneous immunoglobulin replacement therapy. *Journal of clinical immunology*. 2000 Mar 1;20(2):94-100. doi: 10.1023/A:1006678312925
20. Berger M. Subcutaneous immunoglobulin replacement in primary immunodeficiencies. *Clinical immunology*. 2004 Jul 1;112(1):1-7. DOI: 10.1016/j.clim.2004.02.002
21. Berger M, Cupps TR, Fauci AS. Immunoglobulin replacement therapy by slow subcutaneous infusion. *Annals of internal medicine*. 1980 Jul 1;93(1_Part_1):55-6. doi: 10.7326/0003-4819-93-1-55
22. Berger M, Rojavin M, Kiessling P, Zenker O. Pharmacokinetics of subcutaneous immunoglobulin and their use in dosing of replacement therapy in patients with primary immunodeficiencies. *Clinical immunology*. 2011 May 1;139(2):133-41. doi: 10.1016/j.clim.2011.01.006
23. Franchini M. Plasma-derived versus recombinant Factor VIII concentrates for the treatment of haemophilia A: recombinant is better. *Blood Transfusion*. 2010 Oct;8(4):292. doi: 10.2450/2010.0067-10
24. Brinkhous KM, Shanbrom E, Roberts HR, Webster WP, Fekete L, Wagner RH. A new high-potency glycine-precipitated antihemophilic factor (AHF) concentrate: treatment of classical hemophilia and hemophilia with inhibitors. *Jama*. 1968 Aug 26;205(9):613-7. doi: 10.1001/jama.1968.03140350023005

25. Eriksson S. Pulmonary emphysema and alpha-antitrypsin deficiency. *Acta medica Scandinavica*. 1964 Jan 12;175(2):197-205. doi: 10.1111/j.0954-6820.1964.tb00567.x
26. Furusho K, Nakano H, Shinomiya K, Tamura T, Manabe Y, Kawarano M, Baba K, Kamiya T, Kiyosawa N, Hayashidera T, Hirose O. High-dose intravenous gammaglobulin for Kawasaki disease. *The Lancet*. 1984 Nov 10;324(8411):1055-8. doi: 10.1016/S0140-6736(84)91504-6
27. Meulstee J, Darbas A, Van Doorn PA, Van Breiman L, Van der Meche FG. Decreased electrical excitability of peripheral nerves in demyelinating polyneuropathies. *Journal of Neurology, Neurosurgery & Psychiatry*. 1997 Apr 1;62(4):398-400. doi: 10.1136/jnnp.62.4.398
28. Bussel JB, Saal S, Gordon B. Combined plasma exchange and intravenous gammaglobulin in the treatment of patients with refractory immune thrombocytopenic purpura. *Transfusion*. 1988 Jan 2;28(1):38-41. doi: 10.1046/j.1537-2995.1988.28188127949.x
29. Furlan M, Robles R, Morselli B, Sandoz P, Lämmle B. Recovery and half-life of von Willebrand factor-cleaving protease after plasma therapy in patients with thrombotic thrombocytopenic purpura. *Thrombosis and haemostasis*. 1999;81(01):8-13. doi: 10.1055/s-0037-1614408
30. Pickering RJ, Kelly JR, Good RA, Gewurz H. Replacement therapy in hereditary angioedema: successful treatment of two patients with fresh frozen plasma. *The Lancet*. 1969 Feb 15;293(7590):326-30. Doi: 10.1016/S0140-6736(69)91295-1
31. Nathanson S, Frémeaux-Bacchi V, Deschênes G. Successful plasma therapy in hemolytic uremic syndrome with factor H deficiency. *Pediatric Nephrology*. 2001 Jul 1;16(7):554-6. doi: 10.1007/s004670100609
32. Luke TC, Kilbane EM, Jackson JL, Hoffman SL. Meta-analysis: convalescent blood products for Spanish influenza pneumonia: a future H5N1 treatment? *Annals of internal medicine*. 2006 Oct 17;145(8):599-609. doi: 10.7326/0003-4819-145-8-200610170-00139
33. Kajiyama H, Utsumi F, Nakamura K, Tanaka H, Toyokuni S, Hori M, Kikkawa F. Future perspective of strategic non-thermal plasma therapy for cancer treatment. *Journal of clinical biochemistry and nutrition*. 2017;16-65. doi: 10.3164/jcbtn.16-65
34. Kudoyarova-Zubavichene NM, Sergeyev NN, Chepurnov AA, Netesov SV. Preparation and use of hyperimmune serum for prophylaxis and therapy of Ebola virus infections. *The Journal of infectious diseases*. 1999 Feb 1;179(Supplement_1): S218-23. doi: 10.1086/514294
35. Mammoto T, Jiang A, Jiang E, Mammoto A. Platelet-rich plasma extract prevents pulmonary edema through angiopoietin-Tie2 signaling. *American journal of respiratory cell and molecular biology*. 2015 Jan;52(1):56-64. doi: 10.1165/rcmb.2014-0076OC
36. Lee JY, Tuazon JP, Ehrhart J, Sandberg PR, Borlongan CV. Gutting the brain of inflammation: A key role of gut microbiome in human umbilical cord blood plasma therapy in Parkinson's disease model. *Journal of Cellular and Molecular Medicine*. 2019 Aug;23(8):5466-74. doi: oi.org/10.1111/jcmm.14429
37. Casadevall A, Pirofski LA. The convalescent sera option for containing COVID-19. *The Journal of clinical investigation*. 2020 Apr 1;130(4):1545-8. doi: 10.1172/JCI138003.
38. Wan S, Xiang Y, Fang W, Zheng Y, Li B, Hu Y, Lang C, Huang D, Sun Q, Xiong Y, Huang X. Clinical features and treatment of COVID-19 patients in northeast Chongqing. *Journal of medical virology*. 2020 Mar 21. doi: 10.1002/jmv.25783
39. Focosi D, Tang J, Anderson A, Tuccori M. Convalescent plasma therapy for COVID-19: State of the Art. Preprints. 2020 May 28. doi: 10.20944/preprints202004.0097.v7
40. Skendros P, Mitsios A, Chrysanthopoulou A, Mastellos DC, Metallidis S, Rafailidis P, Ntinopoulou M, Sertaridou E, Tsironidou V, Tsigalou C, Tektonidou MG. Complement and tissue factor-enriched neutrophil extracellular traps are key drivers in COVID-19 immunothrombosis. *The Journal of Clinical Investigation*. 2020 Aug 6. doi: 10.1172/JCI141374.
41. Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP, Mostyka M, Baxter-Stoltzfus A, Borczuk AC, Loda M, Cody MJ. Neutrophil Extracellular Traps (NETs) Contribute to Immunothrombosis in COVID-19 Acute Respiratory Distress Syndrome. *Blood*. 2020 Jun 29. doi: 10.1182/blood.2020007008
42. Xie Y, Cao S, Li Q, Chen E, Dong H, Zhang W, Yang L, Fu S, Wang R. Effect of regular intravenous immunoglobulin therapy on prognosis of severe pneumonia in patients with COVID-19. *The Journal of infection*. 2020 Aug; 81(2): 318–356. doi: 10.1016/j.jinf.2020.03.044