



Deceiving The Myths of Nanotechnology in Relation to Nanotoxicity

Jeenatara Begum^{1*} , Tamalika Chakraborty¹, Pramit Sahoo¹, Pritam Roy¹ and Seababrata Bhakta¹

¹ Guru Nanak Institute of Pharmaceutical Science and Technology (An autonomous institution under MAKAUT), 157/F Nilgunj Road, Panihati, Kolkata-700114, India.

Abstract: The toxicity of nanoparticles (NPs) is a critical research topic in nanotechnology, as it is essential to understand the hazards posed by the wide spectrum of NPs that vary in shape, size, and composition. Previous reviews have yet to thoroughly explore the Biological Effective Doses of NPs, which drive toxicity and are influenced by factors such as solubility, charge, shape, contaminants, and the ability of NPs to translocate from the deposition site in the lungs. This review aims to fill the gap in the literature by providing an overview of the possible toxicity of nanoparticles in zebrafish during growth stages, with a focus on oxidative stress, and exploring the available modes of toxicity that are relevant to conventional pathogenic particles. This review also discusses the effects of nanomaterials on the reproductive system in animal models, providing insight into the potential toxicity of nanoparticles in humans. This review aims to provide a comprehensive overview of the toxicity of nanoparticles and to critically explore the challenges associated with implementing nanotechnology, particularly in the pharmaceutical development of novel therapeutic products and regulatory issues. The review also considers recent uses and projected nanotechnology advancements, providing a basis for future research in this field. In conclusion, this review rectifies the lacunae in previously published reviews by providing a comprehensive overview of the toxicity of nanoparticles and exploring the challenges associated with implementing nanotechnology. The aim and objective of this review are to provide a comprehensive understanding of the toxicity of nanoparticles and to guide future research in this field.

Keywords: Nanotechnology, Nanotoxicity, Biological Effective Dose, Zebra Fish, Nanomedicine, Regulatory Challenges

*Corresponding Author

Jeenatara Begum, Guru Nanak Institute of Pharmaceutical Science and Technology (An autonomous institution under MAKAUT), 157/F Nilgunj Road, Panihati, Kolkata-700114, India.

Received On 19 December, 2022

Revised On 1 May, 2023

Accepted On 9 May, 2023

Published On 1 September, 2023

Funding This research did not receive any specific grant from any funding agencies in the public, commercial or not for profit sectors.

Citation Jeenatara Begum, Tamalika Chakraborty, Pramit Sahoo, Pritam Roy and Seababrata Bhakta, Deceiving The Myths of Nanotechnology in Relation to Nanotoxicity.(2023).Int. J. Life Sci. Pharma Res.13(5), P162-P190 <http://dx.doi.org/10.22376/ijlpr.2023.13.5.P162-P190>

This article is under the CC BY- NC-ND Licence (<https://creativecommons.org/licenses/by-nc-nd/4.0>)

Copyright © International Journal of Life Science and Pharma Research, available at www.ijlpr.com

Int J Life Sci Pharma Res., Volume13., No 5 (September) 2023, pp P162-P190



I. INTRODUCTION

Nanotechnology is the future in terms of enabling technological advances across a wide range of industries by offering potential¹. The acceptance of nanotechnology and nano-enabled products are very reliable and have effective use on public cum consumer confidence in both human and environmental safety of this new technology. At the same time, we must ensure that it has been done effectively with the safety and regulation of new technology. The proactiveness of government regulators and international organizations such as the OECD, ISO, and BSI helps or tries to understand nanotechnology and how best to facilitate its safe development and use². Abundant examples of other reports and opinions which have been more specific in their remit, like addressing nanoparticle terminology and definitions³, inhalation toxicity testing for nanomaterials⁴, management of the risk of carbon nanotubes⁵ and specific regulatory frameworks such as REACH for nanoparticles with the effectiveness^{6,7}, and this all for improvement of the safe handling of nanotechnology. But despite the obvious hard work, funding, and good intention that is being focused on the safe development of nanotechnology, there still needs to be more certainty and Besides all the good side effects and therapeutic effects of nanoparticles, there is concern that nanomaterials may harbor an unknown mode of toxicity or 'nano-specific effects.' Much effort has been directed toward understanding these 'nano-specific effects' that lead to the various modes of action in identifying nanoparticle toxicity. All nano-sized particles have novel size-dependent properties, and indeed, it has been argued by Auffan and colleagues (2009) that the evidence for novel size-dependent properties, besides the particle size, should be the primary criterion in any definition of nanoparticles that have relation to health and safety⁸. Recently, a review by Fubini *et al.* and this argument was considered further. It was acknowledged that material at the nano-level should be 'new' by stating when and why it can be considered nano-material⁹. From the availability of the definitions of nanoparticles, an important consideration for a nanoparticle is based on a threshold dimension(s) of 100 nm^{3,10}. That cannot be derived from toxicological evidence of a step-change in toxicity at 100nm nano-sized substances. Much of the evidence is far for 'nano' effects is acknowledged by Fubini *et al.*, who noted that where the biological response is related to the surface area, which forms the interface of the particle, which is insoluble, or the biological interactions, as we know nanoparticles will, of course, show effects orders of magnitude greater than that of bulk particles at the same mass dose due to vast greater surface area⁹. By taking this, it becomes apparent that in the case of toxicity, at least, passing below this threshold into the nano-realm doesn't need to infer any new and specific properties; therefore, the arbitrary assumption of different and 'nano-specific' toxicity appears to be unfounded. Instead of all this, there is likely to be a more gradual magnification of the intrinsic hazard. This statement is echoed by Norppa *et al.*, that it cannot be generally assumed that nanoscale size would be increased the genotoxicity of nanomaterials, or we can say that nanoparticles are generically genotoxic¹¹. Indeed, the view of 'nano-specific' toxicity could be intrinsically not helpful because it labels all the nanomaterials as hazardous or potentially like this, thereby prejudicing against their use. However, in most cases, as for conventional particles, in common use, nanoparticles show a range of inherent toxicities; the majority is low toxicity. In addition to that, the focus on the search for 'nano-specific' may lead to the effect of 're-inventing the wheel' of what is already

known for the conventional particles and thereby delay the important issue of ensuring that the field of nanoparticle hazard can be adequately tested for, qualified and regulations put in place that can be facilitated this is an efficient and proportional manner. The main purpose of this article is to discuss this general basis of toxicity for nanoparticles because, as shown in recent research and studies, is to demonstrate that the mode of action is, in most, if it is not all, cases the same as that shown for the conventional bulk particles. Generally, we can say to understand the basis of toxicity is to understand the driving component, and this can be a variable entity between the materials of the same as well as differing the physicochemical characteristic, and this is described below about the Biologically Effective Dose (BED)¹².

2. NANOMATERIALS

2.1. Definition

According to the EC recommendation¹³, nanomaterial refers to a natural, incidental¹³, or manufactured material comprising particles¹³, either in an unbound state or as an aggregate wherein one or more external dimensions are in size range of 1– 100nm for $\geq 50\%$ of the particles¹³, according to the number size distribution. In environmental, health, safety, or competitiveness concerns, the number size distribution threshold of 50% may be substituted by a threshold between 1 and 50%¹³. Structures with one or more external dimensions below 1 nm, such as fullerenes, graphene flakes, and single-wall carbon nanotubes, should be considered nanomaterials¹³. Materials with surface area by volume over 60 m²/cm³ are also included¹⁴. It defines a nanomaterial in terms of legislation and policy in the European Union¹³. Based on this definition, the regulatory bodies have released guidance to support drug product development¹³. For example, the EMA working group introduces nanomedicines as purposely designed systems for clinical applications¹³, with at least one component at the nanoscale¹³, resulting in reproducible properties and characteristics¹³ related to the specific nanotechnology application and characteristics for the intended use (route of administration, dose)¹³, associated with the expected clinical advantages of nano-engineering (e.g., preferential organ/tissue distribution¹⁵)¹³. The Food and Drug Administration (FDA) has not established its definition for "nanotechnology," "nanomaterial," "nanoscale," or other related terms, instead of adopting the meanings commonly employed about the engineering of materials that have at least one dimension in size range of approximately 1 nanometer (nm) to 100nm¹³. Based on the current scientific and technical understanding of nanomaterials and their characteristics¹³, FDA advises that evaluations of safety, effectiveness, public health impact, or regulatory status of nanotechnology products should consider any unique properties and behaviors that the application of nanotechnology may impart¹².

2.2. Size

The most important feature to consider is size. The conventional size ranges from 1 to 100 nm. However, the maximum size that a material can have to be considered nanomaterial is an arbitrary value because the physicochemical and biological characteristics of the material do not change abruptly at 100 nm¹⁶.

2.3. Particle Size Distribution

The PSD is widely used in nanomaterial identification, reflecting the range of variation in sizes¹⁷. It is important to set the PSD because a nanomaterial is usually polydisperse, which means it is commonly composed of particles of different sizes¹⁷.

2.4. Surface Area

Surface area determination by volume is a relational parameter. Therefore, the material is under the definition if the surface area by volume is larger than 60 m²/cm³¹⁰.

2.5. Composition

2.5.1. Metal Based

Metal-based NPs are an important class of NPs synthesized due to their functions as semiconductors, electroluminescent and thermoelectric materials¹⁸. Recently, interest and development in nanotechnology have been increasing, so many studies have been conducted to evaluate whether the original features of these NPs, such as the large surface area to volume ratio, negatively affect the environment¹⁹.

2.5.2. Carbon Based

A typical carbon-based nanomaterial is carbon nanotubes. Carbon nanotubes were first discovered by Iijima and Ichihashi²⁰ and Bethune et al.²¹ in 1993²⁰. Carbon nanotubes can show significant electrical conductivity²². Also, their tensile strength²³ and thermal conductivity²⁴ are outstanding due to their nanostructure and the strength of the bonds between carbon atoms; because of these properties of CNs can be utilized in many areas of technology, from biomedicine to nanoelectronics²⁵.

2.5.3. Metal Oxide

Metal-oxide NPs are used as industrial catalysts. TiO₂ nanoparticles may disrupt insulin response in Fao cells and cause pregnancy complications in some animal model studies^{26,27}. Studies have shown that other metal-oxide nanoparticles adversely affect reproduction and neonatal development^{28,29}.

2.5.4. Quantum dots

Quantum dots are engineered nanoscale crystals that can transport electrons and convert a spectrum of light into different colors²⁵. Quantum dots make it possible to study

cellular processes and may notably improve the diagnosis and treatment of diseases such as cancers^{30,31}.

3. BIOLOGICAL EFFECTIVE DOSE

In conventional particle toxicology, the dose is defined by the mass or concentration of particles per unit tissue, number of cells, or surface area of cells in cell culture³². Particles are measured through Mass for risk management purposes; the exception is fibers are counted by number. Toxic effects are complicated and rely on the molecular effects at the cellular level and depend on various properties of the particle, basically the physicochemical properties. If the mass increases, it further increases the dose delivery that drives the toxic effect. Povey et al. define the BED as 'the active dose of the agent of interest' and that 'the nearer the dose specified can be to the active dose of the real agent of interest, the more likely it is that an association may exist between an agent and a disease'³³. It has now defined the BED in particle toxicology as 'the entity within any mass dose of particles that drives a critical pathophysiologically relevant form of toxicity in tissue, such as inflammation, genotoxicity or cellular proliferation'³⁴. The Biological Effective Dose of some pathogenic particles has been recognized; in the case of quartz, it is the unpassivated (active) surface area, and in the case of asbestos, it is the long, bio-persistent fibres³⁴. BEDS are still measured by mass. However, no doubt in the future, the development of measuring instrumentation that directly measures the BED will allow the BED to be metric, improving epidemiological dose: responses and thereby improving risk management³².

4. POTENTIAL HUMAN HEALTH EFFECTS OF NANOMATERIALS

4.1. Major Modes of Exposure

The population exposed, the amount of exposure, and the length of exposure, and these situations have very different types of material that people are exposed to³⁵. During a new material's development, it is feasible to be produced under extremely controlled circumstances, usually in very small quantities. Exposures may happen during synthesis or downstream processes such as packaging, shipping, recovery, and storage once the substance enters commercial production³⁶. Nanomaterials can be released intentionally in processes like contaminated land remediation or as waste or industrial pollutants into the air, soil, or water systems. As a result, nanomaterial contamination of the air, water, food supply, or commercial products containing nanomaterials could expose people to them³⁷.

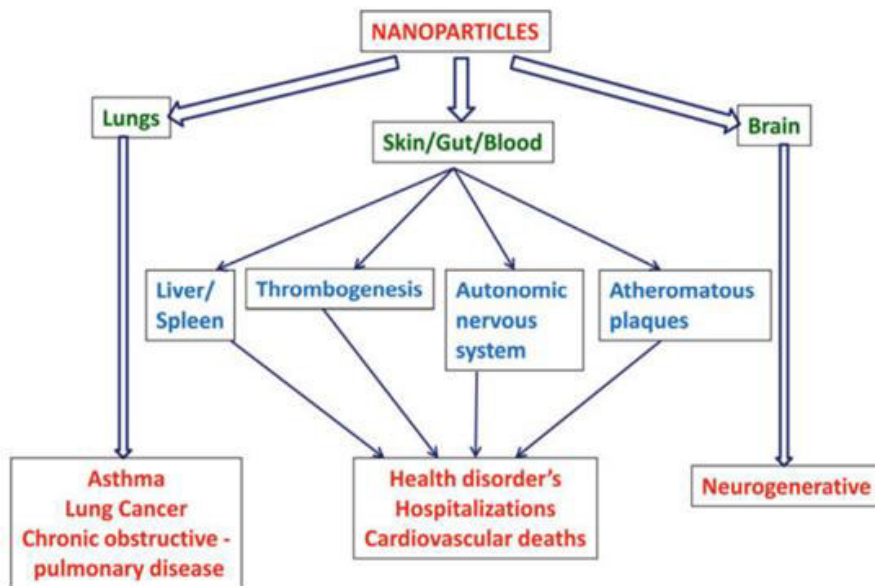


Fig 1: Systematic health effects of nanoparticles on the human body

Image adapted from Viswanath, B. & Kim, S. (2016). Influence of Nano toxicity on Human Health and Environment: The Alternative Strategies [Image]. doi: 10.1007/398_2016_12

4.2. Common Exposure Routes of Humans to Engineered Nanoparticles Present in Consumer Products

| Table 1: Common Exposure Routes of Humans to Engineered Nanoparticles Present in Consumer Products | |
|--|------------------------------|
| Route | Types of consumer products |
| Skin ³⁷ (Dermal) | Sunscreen (lotion) |
| | Skincare (lotion) |
| | Paints and coatings |
| | Sealants |
| | Air fresheners (spray) |
| Lungs ³⁷ (inhalation) | Paints and coatings |
| | Skincare (spray) |
| | Sunscreen (spray) |
| | Food additives and colorings |
| Gastrointestinal tract ³⁷ | Food supplement |
| | Health supplements |
| | Food packaging |

4.3. Effects of Inorganic Nanoparticles on Human Health

Among the most crucial nanomaterials employed in modern technologies are inorganic nanoparticles. Additionally, they are simpler to incorporate into applications³⁸. Insoluble inorganic nanoparticles can be produced using pure metals or various inorganic materials or alloys. They can be distinguished from comparable products found on a wider scale by their nanometric size³⁹. These inorganic nanoparticles lose their electrical, mechanical, and other properties as they become larger⁴⁰. The study of nanomedicine has shown that drug sensitization employing various inorganic nanoparticles (NPs) is a workable and developing method⁴⁰. For instance, when exposed to green light, the well-known photosensitizer Rose Bengal (RB) triggers the production of ROS, which results in cytotoxicity and cell death⁴¹. In addition, the substance releases ions and silver radicals that have an antibacterial effect when it comes into contact with moisture. Lam et al. (2004) identified the cytotoxicity of silver nanoparticles generated by ActicoatTM after finding a significant decline in cell viability in an in vitro investigation of cultured human keratinocytes⁴². Additionally, they showed that 100% anatase nanoparticles,

regardless of size, cause membrane leakage and cell necrosis but do not produce ROS. On the other hand, rutile nanoparticles induce apoptosis by producing ROS. Therefore, the crystal structure and size interaction may be important in mediating nanoparticle toxicity. According to in vitro research by Lucarelli et al. (2004), cobalt (Co) and silica (SiO₂) nanoparticles significantly increased the pro-inflammatory activity of human bone marrow monocytes. Gold nanoparticle (AuNP) particle size and concentration were examined by Yao et al. (2015) for their effects on uptake, accumulation, and cytotoxicity in model intestinal epithelial cells⁴³. As the mean particle size of the AuNPs fell (from 100 to 50 to 15 nm), the rate of absorption by intestinal epithelial cells rose. Still, their cellular accumulation in the epithelial cells shrank. Additionally, mitochondrial membrane depolarization demonstrated that AuNP accumulation resulted in cytotoxicity in intestinal epithelial cells. The results offer crucial insight into the relationship between the dimensions of AuNPs and their absorption through the digestive tract and potential cytotoxicity⁴³. Platinum medicines are given special consideration as anti-cancer treatments. However, no matter how effective they are, platinum medications have downsides. Examples include their dose-limited toxicities, ineffectiveness

against several common malignancies, and the patients' resistance to Pt-based therapy regimens⁴⁴. The cell vacuoles contained PVC, TiO₂, SiO₂, and Co nanoparticles, according to Peters et al. (2004), who investigated the survival and behavior of human endothelial cells in vivo⁴⁵. The synthesis, stability, and toxicity of engineered metal nanoparticles (ENPs) have been thoroughly studied over the past two decades because inorganic elements are an inescapable component of living beings. However, the study of naturally occurring

nanoparticles (NNPs) and their creation, destiny, and ecological implications have recently attracted interest⁴⁰. Solid organic nanoparticles, typically lipids or polymeric substances, make up organic nanoparticles (Lambert et al. 2014). This nanoparticle form has undergone extensive development and research over the past few decades due to its high potential in various industrial fields, including electronic and photonic, conducting materials and sensors, medicine and biotechnology, and others⁴⁶⁻⁴⁸.

4.1. In vivo observed effects supported by in vitro evidence.

| Table 2: In vivo observed effects induced by engineered nanoparticles supported by in vitro evidence. | |
|---|--|
| In vitro evidence | In vivo observed evidence |
| Enhanced cytotoxicity in exposed cell culture samples | Chronic obstructive pulmonary disease (COPD) |
| Proliferative responses brought on by DEP component extracts | Hyperplasia |
| Gap Junction Intercellular Communication (GJIC) changes caused by macrophage-dendritic transepithelial cells | Particle translocation |
| Pneumocytes, macrophages, and other exposed cells in co-cultures that directly activate endothelial cells or indirectly trigger them. Tight junction-related changes to the TEER values | Systemic and endothelial dysfunction |
| When exposed to PM, lung epithelial cells' NADPH-oxidase produces more ROS. | Oxidative stress |
| IL-1b, IL-6, IL-8, TNFa, MCP-1, and other molecules are secreted by lung cells, macrophages, and cocultures. | Local and systemic inflammation |

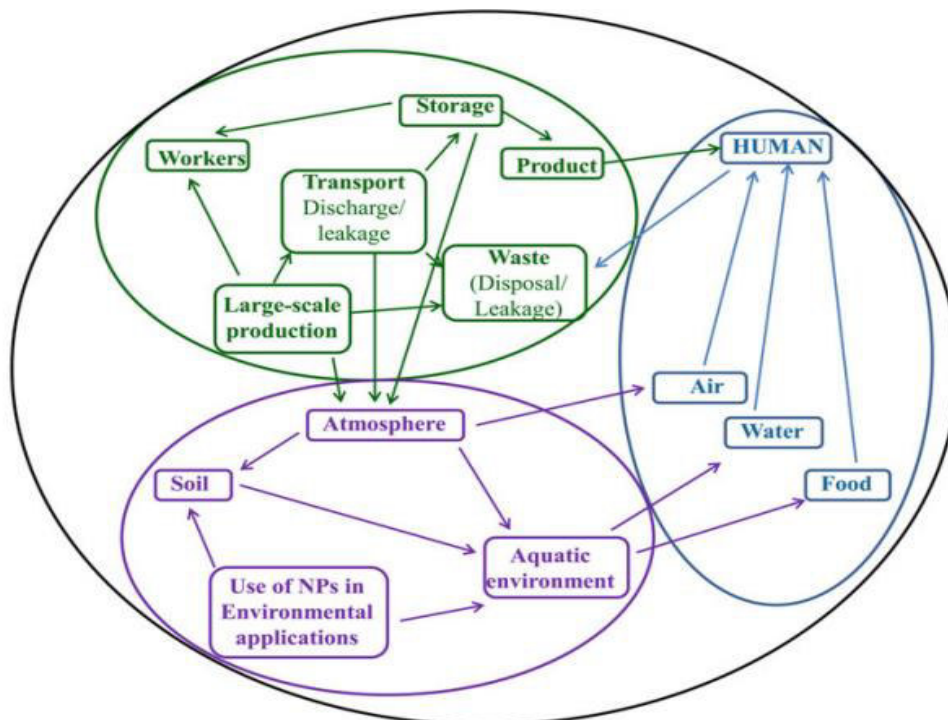


Fig 2: Multiple scenarios through which nanoparticles enter into the environment and humans

Image adapted from Viswanath, B. & Kim, S. (2016). Influence of Nanotoxicity on Human Health and Environment: The Alternative Strategies [Image]. doi: 10.1007/398_2016_12.

4.2. Environmental Issues

Energy, security, information technology, agriculture, environmental protection, and healthcare are just a few industries where nanotechnology is revolutionizing the landscape⁸². The development of nanomaterials has generated impressive scientific activity, with an exponential rise in the number of peer-reviewed papers on the subject during the past ten years. Currently, national nanotechnology projects exist in more than 60 nations. However, the success or failure of nanotechnology may depend on its capacity to address

environmental challenges. Although there is limited advice for researchers on how to put such practices into practice, Responsible Research and Innovation provide a framework for assessing the ethical dimensions of innovation processes. Any research proposal should be anticipatory, looking ahead to potential technological effects; reflective, looking at the goals and purposes of technologies as well as the uncertainties in risk assessment; deliberative, looking at the idea that public and diverse stakeholders' perspectives are actively taken into account during design processes, and responsive, looking at

the actual alteration and shaping of technological trajectories in response to deliberation⁸³. Many scientists are putting in a lot of effort to address several important environmental problems, such as the following: • To what extent might the manufacturing and usage of nanoproducts be expected to result in the release of hazardous elements into the environment? • What possible environmental problems can this nanotechnology cause? • Because nanoproducts bioaccumulate in living tissue, may nanotechnology contribute to environmental degradation? • What impact will laws have on this nanotechnology? Researchers looking at how nanoparticles affect ecology have found that some nanomaterials are hazardous to the environment. The precautionary principle should be used to reduce preventive risk, notwithstanding ongoing scientific uncertainties. Environmental inputs should be avoided as much as possible. The environmental relevance of materials and the complexity of natural systems should be increasingly the focus of ecotoxicological study. Due to their tiny size and increased specific surface areas, these products are expected to intensify chemical reactivities sensitive to exposed surface sites⁸⁴.

4.3. Environmental Fate of Nanomaterials in Air

The processes through which ultrafine particles in the air are lost have been clearly defined by numerous studies^{75,85}. However, several pressing difficulties must be resolved to reveal the mechanisms that control their behavior, movement, and destiny⁸⁶. In aerosol systems, nanoparticles will be very mobile and mix quickly. Unlike the other environmental compartments, engineered nanoparticles suspended in air will probably be exposed to sunlight, especially UV wavelengths, to a considerably larger amount⁸⁷. It widens the range of photochemical changes. Additionally, the gravitational settling velocity, which is inversely proportional to particle diameter, influences the deposition of nanoparticles in the air. Smaller nanoparticles in the air deposit much more gradually than larger ones. Agglomeration, as a result, will greatly enhance the deposition of engineered nanomaterials. Other processes are considered significantly less significant or even inappropriate for nanomaterials in the air compared to photochemical reactions, aggregation, and deposition^{84,88}. Understanding possible nanomaterial sources and their degradation, transformation, and existence is necessary to comprehend the fate and behavior of nanomaterials in the environment. Different outcomes for nanomaterials in the environment are anticipated depending on their physical and chemical characteristics, the medium in which they are contained, and interactions with other environmental pollutants. The three main sources of atmospheric nanomaterials are as follows: Specifically, there are three types of emissions: (1) primary emissions, which are defined as those that are outwardly released from industrial combustion and road traffic exhaust; (2) secondary emissions, which are defined as those that are produced in the atmosphere by the compression of low-volatility vapors from atmospheric gas oxidation; and (3) formation during diesel exhaust dilution⁸⁹. Due to a lack of techniques that can separate manufactured nanomaterials from background concentrations from other sources, comparable to the situation in aquatic and terrestrial settings, there needs to be data on engineered nanomaterials in the atmosphere⁸⁹. According to the literature, there are many processes that fine, ultrafine, and nanomaterials can go through in the atmosphere^{90,91}. Some nanomaterials can be created by condensing low-volatility chemicals. They can be shrunk by evaporating adsorbed water or other volatiles,

causing a departure in the particle size distribution but not the overall numerical concentration. Nanomaterials in the atmosphere can mix to produce larger particles while having a lower numerical concentration⁹². Dry and wet deposition, which may remove incredibly small particles of natural origin and presumably create nanomaterials, are other methods for removing nanoparticles from the atmosphere. As a result, particle number concentration falls, and the particle size distribution changes to bigger sizes⁹³.

4.4. Environmental Fate of Nanomaterials in Water

Aggregation and disaggregation, diffusion, the interaction of nanoparticles with natural water components, transformation, biotic and abiotic degradation, and photoreaction can all impact how nanomaterials behave in aquatic environments⁶⁴. The destiny and behavior of manufactured nanomaterials released into the aquatic environment can be understood by referring to the existing literature on the fate and behavior of naturally occurring colloidal particles. Nanomaterials are currently highly suggested for wastewater treatment due to their outstanding features. Although certain studies have documented the numerous advantages of nanotechnology in wastewater cleanup, more research needs to be done on the fate and potential effects of the solid residues that these technologies produce⁹⁴. The impact of particle size and coating material on these behaviors were examined in studies on the aggregation and sedimentation kinetics of citrate- and polyvinylpyrrolidone-coated silver nanoparticles (Cit-AgNPs) in calcium chloride (CaCl₂) solutions. Cit-AgNPs aggregated quickly and settled as the ionic strength increased⁹⁵, whereas PVP-AgNPs did not⁹⁵, due to the PVP coating's steric hindrance effects⁹⁵, even at an ionic strength of 10 mM CaCl₂⁹⁵. It is interesting to note that PVP-AgNPs did not aggregate during the first week of sedimentation, and this propensity is influenced by particle size. These results suggest that the coating material type and particle size significantly impact how nanoparticles behave in water⁹⁵. In addition, nanoparticles may interact with aquatic life and have detrimental consequences at different levels of biological organization. Despite a recent study of the ecotoxicological concerns that ENMs may pose to aquatic creatures⁹⁶⁻⁹⁹, Their biological danger and mode of action are still unknown. Due to interactions with natural organic matter, natural colloids, and suspended particulate matter, nanoparticles in aquatic settings may aggregate and perhaps silt from the solution. Sedimentation and aggregation may aid in the movement of nanoparticles from the water column to benthic sediments. In addition, depositing and filter-feeding species in aquatic habitats bioaccumulate nanoparticles. Since there are no reliable and sensitive analytical techniques for identifying and characterizing nanoparticles in complex environmental matrices such as natural fluids and soils¹⁰⁰, although such interactions have not yet been well researched, they may have a considerable impact on the destiny and toxicity of nanoparticles.

4.5. Environmental Fate of Nanomaterials in Soil

A layered food web structure and a complex interface between gases, solids, water, organic and inorganic substances, and living things are matriculated by soil. Because they are so small, nanomaterials can pierce soil pores¹⁰¹. They can become immobilized because dirt particles adhere to their enormous surface area¹⁰². Sedimentation, filtration, or straining can be used to immobilize large aggregates of nanomaterials in smaller pores¹⁰¹. In the natural porous environment, there are

currently limited reports on the movement and destiny of nanomaterials. Reports state that the transfer speed depends on the kind of nanomaterials employed^{103,104}. While most nanoparticle toxicity mechanisms are unknown, some probable causes include membrane rupture or membrane potential, protein oxidation, genotoxicity, interruption of energy transmission, creation of reactive oxygen species, and release of hazardous components¹⁰⁵. High surface area to volume ratios, surface charges, hydrophobic and lipophilic groups enabling them to interact with proteins and membranes, complementary effects of nanostructures that inhibit enzyme activity, bioaccumulation, and increasing

chemical composition their reactivity could all contribute to these toxicity mechanisms¹⁰⁶. Polymers and surfactants improve the transport of nanoparticles. Numerous researchers are examining the part that natural organic matter plays in transport assisted by nanoparticles. The soil matrix's characteristics may influence the diffusion and mobility of nanoparticles. The physical-chemical features of the nanoparticles, the characteristics of the soil and environment, and the interaction of the nanoparticles with naturally occurring colloidal material all affect how mobile they are in soils. Table 3 lists some of the current ENPs along with their impacts on human health and the environment.

Table 3: List of some existing ENPs and their health and environmental effects

| Nanoparticle | Environmental effects | Health effects |
|---------------------------------|---|--|
| Carbon nanotubes | cause indirect impacts when in contact with environmental organisms' surfaces; harm the environment | Apoptosis, lowered cell viability, lung toxicity, oxidative stress, slowed cell growth, skin irritation, etc. |
| Fullerenes | Effects on aquatic ecosystems, soil organisms, enzymes, and chemical binding to fullerenes may impact the toxicity of other environmental pollutants. | Some examples are reduced cell viability, oxidative stress, apoptosis, and delayed cell growth. |
| Heterogeneous nanostructures | Numerous physical, chemical, and environmental factors, including ecosystem harm, affect toxicity. | Cellular growth arrest, and occasionally even cell death, chromatin condensation, and free radical production |
| Nanosilver | being released into the environment, it passes through various changes and manifests negative effects. | Non-specific immune system changes, altered cell signaling, apoptosis, cell necrosis, oxidative stress, etc. |
| Nanostructured flame retardants | persistent and have a propensity to build up in the environment, harmful to wildlife, flora, etc. | Cardiovascular effects, fibrosis, oxidative stress, cytotoxicity, carcinogenic, etc. |
| Polymeric nanoparticles | Environmental exposure risk factor potential | Oxidative stress, inflammation, changes in the shape and operation of cells, etc. |
| Silicon-based nanoparticles | Potentially dangerous environmental exposure factors, detrimental ecosystem impact, etc. | Heart problems, cytotoxicity, a rise in oxidative stress, etc. |
| TiO ₂ nanoparticles | Stress photosynthetic organisms and the carbon and nitrogen cycles in an aquatic habitat. | In humans, excessive exposure may lead to increased oxidative stress, slowed cell growth, minor lung abnormalities, etc. |

5. NANO-BIO INTERFACE AND NANOTOXICOLOGY

5.1. Nano—bio interface

Research in numerous fields of nanotechnology has mostly centered on proteins and nucleic acids^{107–111}. Compared to a 10 nm nanoparticle, a single cell, which is generally tens of microns, is immense (Fig 3). To research biological processes, including medication transport and cellular-level bioimaging, scientists worldwide have been using a variety of inorganic, organic, and composite nanoparticles^{112–116}. Many publications

have recently examined the relationship between a protein and a nanoparticle^{117–119}. Compared to a 10 nm nanoparticle, the APP and a tiny therapeutic molecule (such as DHED) are incredibly small, making it challenging to probe biologically significant nanoparticle molecules. In truth, a nanoparticle put into a live system will interact with the environment endless times, regardless of size. Studies on the interface between biological systems and nanostructured materials, starting with proteins and moving up to the cell, will be a significant step forward in understanding bio-systems important to pharmacology, pharmacology, and medicine.

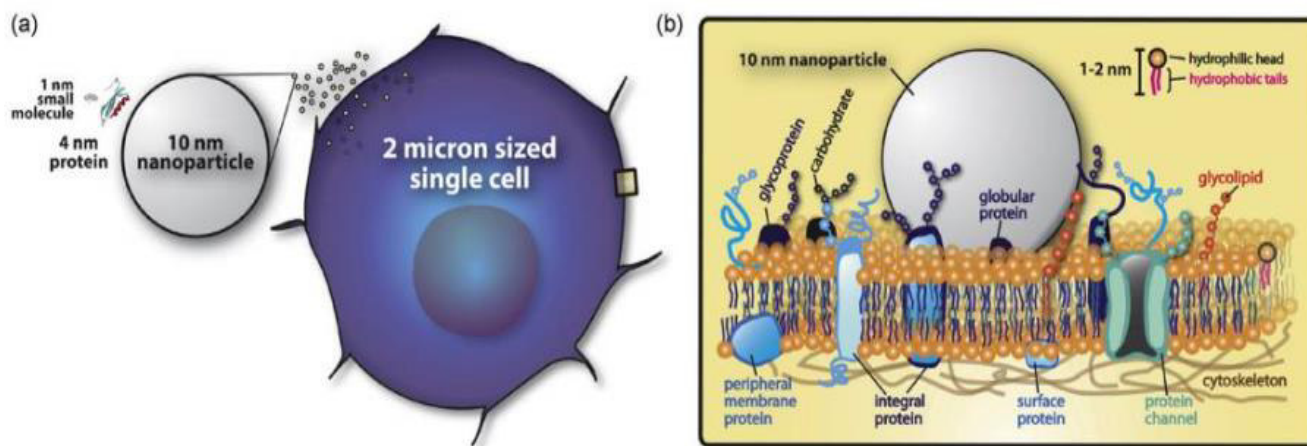


Fig 3: Compared to a 10 nm nanoparticle, proteins (e.g., APP; X-ray crystal structure obtained from www.pdb.org (Berman et al., 2000), protein ID 2FKL; visualization done by Accelrys Discovery Studio Visualization 1.7 software) and small molecules (e.g., DHED) are small in size and volume. A mammalian cell of proteins, nucleic acids, and other small to large molecules is a thousand times larger in volume and size compared to a 10 nm nanoparticle. (b) Cell membrane incorporating various proteins and a single 10 nm nanoparticle.

Image adapted from Viswanath, B. & Kim, S. (2016). Influence of Nanotoxicity on Human Health and Environment: The Alternative Strategies [Image]. doi: 10.1007/398_2016_12

Studying the bio-nano interface is a completely different undertaking because there are no straightforward tools for probing the interaction in real-time or in situ. On the other hand, nanotoxicology, which is the study of the bioeffects of nanomaterials, is a rapidly expanding discipline with some immediate use. Recent years have seen a significant increase in studies into the toxicity of nanomaterials on the environment and living systems. For instance, the University of California has a robust nanotoxicology program led by UCLA and UCSB as part of its UC Toxic Substances Research and Teaching Program

(<http://www.cnsi.ucla.edu/staticpages/education/nanotox-program>;

<http://www.bren.ucsb.edu/news/press/nanotoxicology.htm>).

For the first time in US history, Berkeley (CA) has chosen to control nanotechnology through the law, with UC Berkeley and LBNL (Lawrence Berkeley National Laboratory) involved in various nanotech¹²⁰. The International Council on Nanotechnology (ICON) and the Center for Biological and Environmental Nanotechnology (CBEN) at Rice University are both aiming to compile a database of materials based on nanotechnology (<http://cben.rice.edu/>; <http://icon.rice.edu/>). NCL (Nanotechnology Characterization Laboratory), run by a chemist specializing in nanomaterials with dimensions less than 100 nm, was recently established as a separate organization by the National Cancer Institute (NCI). Internationally, Singapore's IBN (Institute of Bioengineering and Nanotechnology), run by A*STAR, is a multidisciplinary research park that merges the study of biological systems at the nanoscale scale. The fact that a materials scientist serves as the organization's head suggests that IBN focuses more on the materials it creates, which will help the transition from nanotechnology to biotechnology. From the perspective of both the material and the biological system, a basic understanding of nanomaterial toxicity (nanotoxicology) is highly desired. Toxicology assessments of nanoscale materials should attract greater attention than ever from the general public, the government, or those involved in nanomaterial development, with the rising commercialization of goods

ranging from tennis balls to cosmetics^{121–124}. The knowledge gained from these studies on nanotoxicology should assist scientists in making better decisions on the kind of nanomaterial that can be utilized to investigate, for instance, the synaptic plasticity of a neuron. To do this, we will examine the literature on the development of nanotoxicology and offer a few tables to help with material selection. With the available data, however, it is usually challenging to determine the toxicity of particular nanomaterials since, like any tiny molecule (such as a medicine), toxicity is dose, exposure, and route dependent. Furthermore, it is impossible to predict the effects of nanotoxicology on humans just from investigations on cultured cells or animals.

5.2. Nanotoxicology

Different forms of artificial nanomaterials currently exist due to businesses' and academics' unprecedented and intensely focused efforts in recent years. Over 3200 papers were published exclusively on producing nanostructured materials between 2006 and 2007, an exponential rise (Figure 4). This enormous rise in publications has led to the release of hundreds of in vitro toxicology research^{124–130}, as well as countless evaluations and viewpoints^{121–123,131–138}. Contrarily, in vivo, toxicology needs the test subject to internalize the test sample, whether a little mouse or a large creature like a dog or a monkey. This method examines toxicity (i.e., LD50, pathophysiology) through inhalation, injection, and oral digesting. However, given the extensive use of synthetic engineering, testing the toxicity of nanomaterials on whole animals is challenging^{139–146} is carried out extremely specifically by various research groups, and access to proprietary information on synthesis—especially from the industry—can be challenging. Additionally, setting up, carrying out, and controlling an in vivo test is a difficult ethical and administrative task. Individual research initiatives must work with institutional approval organization(s) like IACUC (Institutional Animal Care and Use Committees).

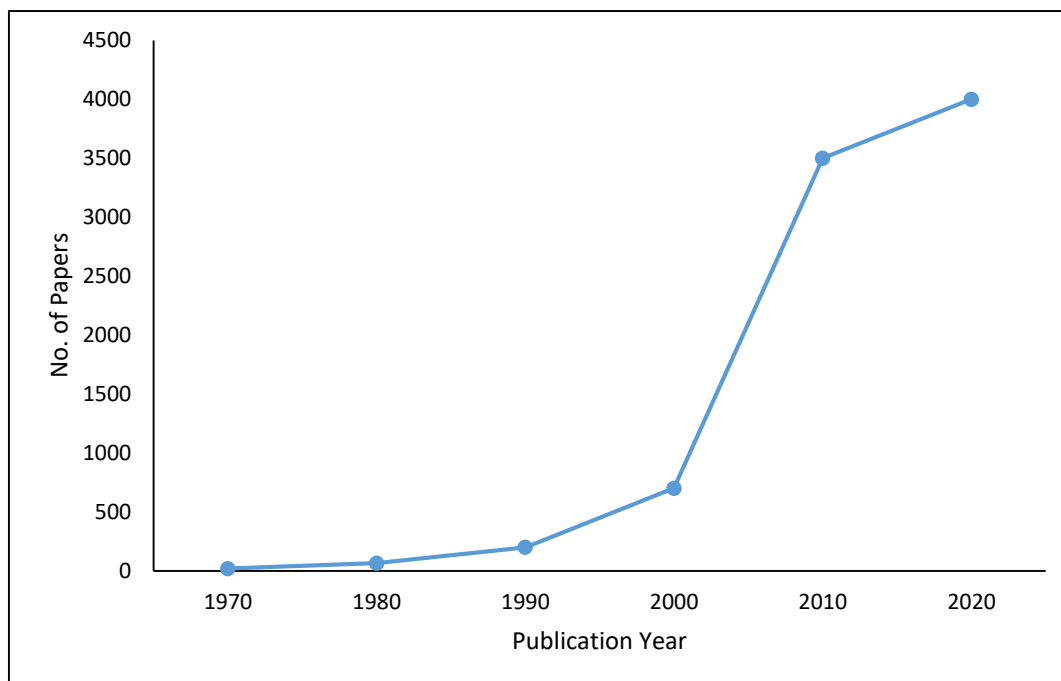


Fig 4: The number of papers published solely on the synthesis of nanostructured materials (According to Web of Science Search Results)

6. DYNAMIC BEHAVIOR OF NANOMATERIAL AND APPLICATION IN NANOMEDICINE

The application of nanomaterials in medical field purposes in the form of nanomedicine: which has three different areas in it is a diagnosis that is mainly known as nano-diagnosis, the second one is controlled drug delivery, also known as monotherapy, and the last one is regenerative medicine. A new area of the medical world that mainly combines diagnostics and therapy, termed theragnostic, is emerging and is a promising approach that holds in both systems, which are the same that are diagnosis/imaging agents and the other is medicine. The promise held by nanomedicine is the changes in clinical practice through the introduction of novel medicines for both diagnosis and treatment, which has enabled to address the of unmet medical needs (a) by integrating effective molecules that otherwise could not be used due to having the high toxicity (e.g., Mepact), (b) by exploiting multiple mechanisms of actions (e.g., Nanomag, multifunctional gels) (c) by maximizing efficacy (e.g., by increasing bioavailability) and also by reducing the dose and the toxicity, (d) by providing drug targeting, controlled and site-specific release, and by favoring a preferential distribution within the body (e.g., in areas with cancer lesions) and that is improved the transportation across biological barriers¹⁴⁷. The result of the intrinsic properties of nanomaterials has brought so many advantages to the development of the pharmaceutical world. Because of the small size of the nanomaterials or nanoparticles, it has a high specific surface area about the volume. Therefore, the surface energy of the particle is increased by making the nanomaterials much more reactive. The absorbance characteristics of the nanomaterials towards the biomolecules, e.g., protein and lipids, have a large tendency when it is in contact with the biological fluid. Important interactions with living matter mainly rely on the plasma/serum biomolecule adsorption layer, known as "corona," which mainly forms on the surface of the colloidal nanoparticles¹⁴⁸. Its composition mainly depends on the portal of entry into the body and on the particular fluid from which the nanoparticle comes, which may be blood, lung fluid, gastrointestinal fluid,

etc. Changes in "corona" can be influenced by additional dynamic changes that constitute the nanoparticle crosses from one biological compartment to another one¹⁴⁹. Besides that, the optical, electrical, and magnetic properties also can be changed and harmonic by the electron confinement in the nanomaterials. In addition, nanomaterials can also be engineered to have different sizes, shapes, chemical compositions, and surfaces, and they can interact with specific biological targets¹³⁸. By restoring careful particle design, we will get a successful biological outcome. For these reasons, comprehensive knowledge of the interactions between nanomaterials and biological systems is required. Among of two, the first one is related to the physiopathological nature of the diseases. The main biological processes behind the diseases occur at the nanoscale and can rely on, e.g., mutated genes, misfolded proteins, viral infection, or bacterial infection. Understanding of the molecular processes will be provided with the rational design of engineered nanomaterials to target the specific action site that is mainly desired site of action in the body¹⁵⁰. Another concern is the interaction between the environment of the biological fluids and the nanomaterial or nanoparticle surface. In the context of characterization of the biomolecules, the corona is of the uttermost importance for understanding the mutual interaction between nanoparticle and cell called nanoparticle-cell affects the biological responses. This intersection mainly comprises dynamic mechanisms involving the exchange between biological components' surfaces, e.g., proteins, membranes, phospholipids, vesicles, organelles, and the nanomaterial or nanoparticle surfaces. The interaction stems from the composition of the suspending media and the nanomaterial. The size, shape, surface area, surface charge, chemistry, energy, roughness, porosity, valence, conductance states, the presence of ligands, or the hydrophobic/hydrophilic character are some characteristics of the nanomaterials that influence the respective surface properties. In addition, the presence of water molecules, acids and bases, salts, and multivalent ions will influence the interaction. All these aspects will govern the characteristics of the interface between the biological components and nanomaterial and promote different cellular

fates¹⁵⁰. A piece of deeper knowledge of how the physicochemical properties of the bio interface influence the cellular signaling pathway and kinetics and transport will thus provide critical rules that design the nanomaterials¹⁵¹.

7. CONSIDERATION OF DOSE IN PERSPECTIVE OF NANOMEDICINE

The COVID-19 pandemic reminds us that we need high-value flexible solutions to urgent clinical needs, including simplified diagnostic technologies suitable for use in the field and for delivering targeted therapeutics¹⁵². Nanotechnology is an important resource for this, as a generic platform of technical solutions to tackle complex medical challenges¹⁵³. Even though there are more than 50 formulations currently on the market, and the recent approval of 3 key nanomedicine products (e.g., Onpatro, Hensify, and Vyxeos), has revealed that the nanomedicine field is concretely able to model products that overcome critical barriers in conventional medicine in a special unique manner¹⁵⁴, and also to deliver within the cells new drug-free therapeutic effects by using pure physical modes of action and therefore make a difference in patients' lives¹⁵⁴. One major advantage of nanomedicines as designed objects over other medicinal products is their high level of uncoupling between their functional requirements and their design parameters (nanoparticle & drug, for instance), described by the general theory of axiomatic design by P Suh in the 1990s¹⁵⁴. However, it is often claimed that nanomedicine failed to meet the initial expectations in drug delivery since less than 2% of the active pharmacological ingredient (API) is locally released, e.g., in cancer treatment in the tumoral tissues¹⁵². On the other hand, Abraxane demonstrates a significantly higher response rate, longer time to tumor progression, and absence of hypersensitivity reactions¹⁵⁵. Nanotechnology also expurgates transdermal delivery, a safe, noninvasive method of administering drugs¹⁵⁶. Applied directly onto the skin, transporting large-molecular weight proteins like vaccines across the skin is relatively inefficient. Recent evidence has shown that this barrier can be covered by properly structured nanosized particles¹⁵⁶. Nanoparticles can also provide an efficient delivery tool for drugs bypassing the blood-brain barrier, such as chemotherapeutic agents for brain malignancies, antiepileptics, and anesthetics (e.g., Dalargin)¹⁵⁷. For example, Polysorbate 80-coated nanoparticles loaded with doxorubicin (5 mg/kg) achieved high brain levels of 6 µg/g brain tissue. In contrast, all the controls¹⁵⁷, including uncoated nanoparticles and doxorubicin solutions mixed with polysorbate, did not reach the analytical detection¹⁵⁷.

8. NANOTOXICOLOGICAL CLASSIFICATION SYSTEM

Hitherto, different risk assessment approaches have been reported. The DF4nanoGrouping framework concerns a functionality-driven scheme for grouping nanomaterials based on their intrinsic properties, system-dependent properties, and toxicological effects¹⁵⁸. Accordingly, nanomaterials are categorized into four groups, including possible subgroups¹³. The four main groups encompass (1) soluble, (2) persistent high aspect ratio, (3) passive, that is, nanomaterials without obvious biological effects, and (4) active nanomaterials¹³, that

is, those demonstrating surface-related specific toxic properties. The DF4nanoGrouping foresees a stepwise evaluation of nanomaterial properties and effects with increasing biological complexity¹³. In case studies that include carbonaceous nanomaterials, metal oxide, metal sulfate nanomaterials, amorphous silica, and organic pigments (all nanomaterials with primary particle sizes smaller than 100nm), the usefulness of the DF4nanoGrouping for nanomaterial hazard assessment has already been established¹³. It facilitates the grouping and targeted testing of nanomaterials. It also ensures that enough data for the risk assessment of a nanomaterial are available and fosters the use of non-animal methods¹⁵⁹. More recently, DF4nanoGrouping developed three structure-activity relationship classification decision tree models by identifying structural features of nanomaterials mainly responsible for the surface activity based on a reduced number of descriptors: one for intrinsic oxidative potential, two for protein carbonylation, and three for no observed adverse effect concentration¹⁶⁰. Keck and Müller also proposed a nanotoxicological classification system (NCS) (Figure 5) that ranks the nanomaterials into four classes according to the respective size and biodegradability¹⁶¹. Due to the size effects, this parameter is assumed as truly necessary because when nanomaterials are getting smaller and smaller, there is an increase in solubility¹³, which is more evident in poorly soluble nanomaterials than in soluble ones¹³. The adherence to the surface of membranes increases with the decrease in size, and another important aspect related to the size that must be considered is the phagocytosis by macrophages¹³. Above 100 nm, nanomaterials can only be internalized by macrophages, a specific cell population, while nanomaterials below 100nm can be internalized by any cell due to endocytosis¹³. Thus, nanomaterials below 100nm are associated with higher toxicity risks than nanomaterials above 100 nm¹⁶¹. Biodegradability was considered a required parameter in almost all pharmaceutical formulations¹³. The term biodegradability applies to the biodegradable nature of the nanomaterial in the human body¹³. Biodegradable nanomaterials will be eliminated from the human body¹³. Even if they cause inflammation or irritation, the immune system will return to its regular function after elimination¹³. Conversely, non-biodegradable nanomaterials will stay forever in the body and change the normal function of the immune system¹⁶¹. Two more factors must be considered besides the NCS: the route of administration and the biocompatibility surface¹³. When the NCS13 classifies a particle, toxicity depends on the route of administration. For example, the same nanomaterials applied dermally or intravenously can pose different risks to the immune system¹³. In turn, a non-biocompatibility surface (NB) can activate the immune system by adsorption to proteins like opsonins¹³, even if the particle belongs to class I of the NCS (Figure 5)¹³. The biocompatibility (B) is dictated by the physicochemical surface properties, irrespective of the size and biodegradability¹³. It can lead to a further subdivision into eight classes I-B, I-NB, IV-B, and IV-NB¹⁶¹. NCS is a simple guide to evaluating the risk of nanoparticles, but many other parameters play a relevant role in nanotoxicity determination¹⁶¹. Other suggestions encompass more general approaches, combining elements of toxicology, risk assessment modeling, and tools developed in multicriteria decision analysis¹⁶².

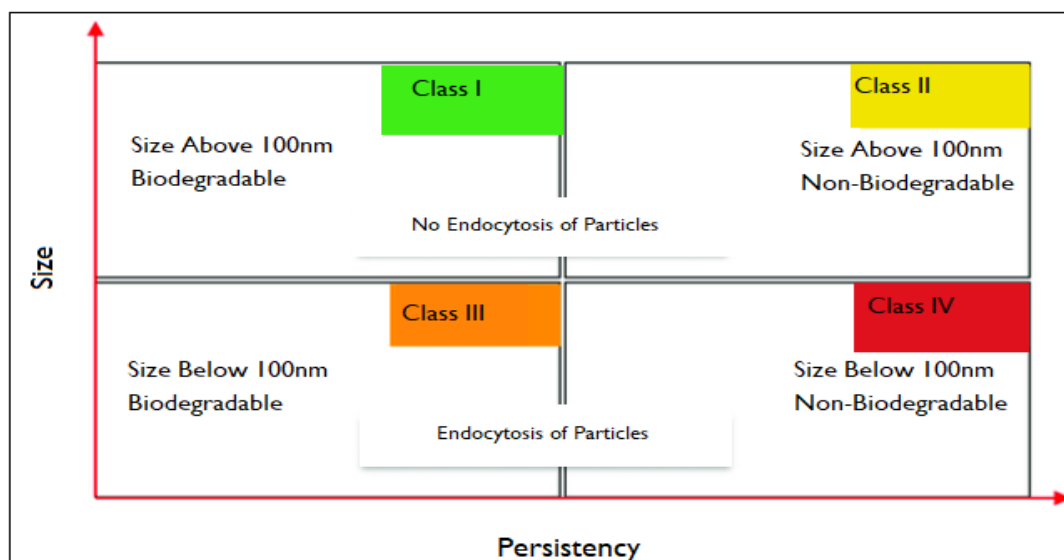


Fig 5. Nanotoxicological Classification

9. TOXIC EFFECTS OF NANOPARTICLES ON SYSTEMS

Experimental studies have demonstrated that nanoparticles harm numerous systems by entering the body in three ways. This section largely uses animal experiments to describe the harmful effects of nanomaterials on systems.

9.1. Circulatory system:

Nemmar et al. found that intravenous administering iron oxide nanoparticles to mice caused DNA damage and myocardial oxidative stress¹⁶³. Magaye et al. discovered cardiac toxicity-arrhythmia and toxic effects in organs such as the liver, spleen, and lung in a study of rats receiving intravenous Ni nanoparticles¹⁶⁴.

9.2. Digestive system

Zirconia oxide nanoparticles at 100 ppm induce liver injury in rats, claim Arefian et al.¹⁶⁵. Mice's liver is likewise harmful to iron oxide nanoparticles.¹⁶⁴

9.3. Endocrine system

Oral iron oxide nanoparticles have been linked to abnormal thyroid hormone levels in rats, according to Yousefi et al.¹⁶⁴.

9.4. Immune system

According to Xu et al., TiO₂ nanoparticles significantly increased the number of white blood cells in mice¹⁶⁶. Additionally, white blood cell production is increased by iron oxide nanoparticles, with the liver and spleen being the most immunologically impacted organs¹⁶⁷.

9.5. Respiratory system

According to Cai et al., the lungs become hazardous when metal nanoparticles (cobalt oxide, nickel oxide, and titanium oxide) are delivered via oropharyngeal aspiration.¹⁶⁸ Iron oxide nanoparticles have also been linked to pulmonary damage in rats, according to Sadeghi et al.¹⁶⁹.

9.6. Urinary system

According to Saranya et al., kidney cells in monkeys, pigs, and cattle are toxic to zinc oxide, iron oxide, and copper nanoparticles¹⁷⁰. Furthermore, TiO₂ nanoparticles administered intraperitoneally to rats result in kidney deterioration, according to Fartkhoni et al.¹⁷¹.

9.7. Nervous system

When vision and hearing toxicity in animal ears and eyes were investigated, very little or no harm was discovered overall^{172,173}.

9.8. Reproductive system

Zinc oxide nanoparticles were administered intraperitoneally to mice, and Mozaffari et al. found that this resulted in a loss and reduction of seminiferous tubule cells¹⁷⁴. According to Kong et al., nickel nanoparticles affect rat sperm motility and FSH and LH hormone levels¹⁷⁵.

10. TOXICITY MECHANISMS OF NANOPARTICLES

The mechanical impacts brought on by the physicochemical characteristics of nanoparticles are what induce toxicity. Reactive oxygen species (ROS) are produced directly or indirectly, which is the fundamental process of creating hazardous effects. In vitro, ROS production is harmful via a variety of cell pathways^{176,177}. In mitochondria, ATP is produced due to the conversion of molecular oxygen to water. During this process, superoxide anions and radicals with various oxygens are generated. Hydroxyl radicals, single oxygen radicals, hydrogen peroxide radicals, and superoxide anion radicals are some ROS generated¹⁷⁷. Overproduction of free radicals, which interfere with cellular signaling and the mitogenic response in cells, causes damage to their physiological activities^{178,179}, resulting in cell disruption. Nanomaterials affect cells in cytotoxic and genotoxic ways (Figure 6). Nanomaterials have modest dimensions, but because of their high surface reactivity and specific surface area, they emit more ROS¹⁸⁰.

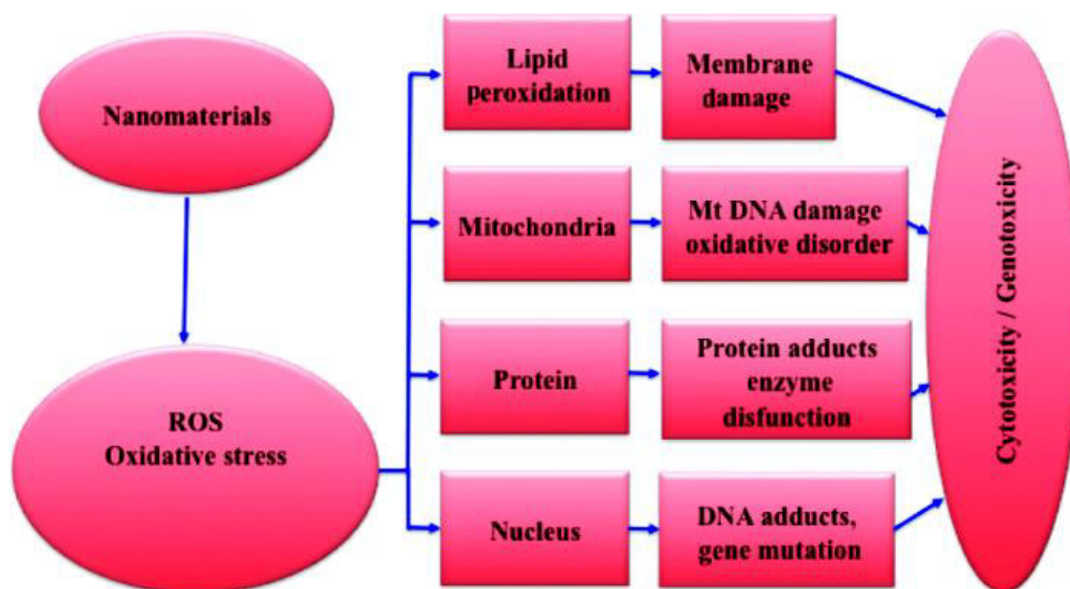


Fig 6: ROS and nanomaterial toxicity

Image adapted from Viswanath, B. & Kim, S. (2016). Influence of Nanotoxicity on Human Health and Environment: The Alternative Strategies [Image]. doi: 10.1007/398_2016_12

Studies in living tissues, including human erythrocytes and skin fibroblasts, have shown that different nanomaterials can be hazardous by activating ROS¹²⁴. Kim et al. claim that nano-Ag produces genotoxicity and oxidative stress in cultured live tissue. Nano-Ag causes mutations in mice by boosting the generation of ROS, according to Mei et al.^{181,182}. Hsin et al. claim that nano-Ag activates ROS in the mitochondrial pathway to induce cytotoxicity¹⁸³. According to Akhtar et al., nano-CuO lipid peroxidation and ROS generation from silica nanoparticles cause cytotoxicity in cell membranes and mouse embryonic fibroblasts^{184,185}. According to Girgis et al., nano-Au toxicity in mice was brought about by increased oxidative stress¹⁸⁶. Shvedova et al. claims that keratinocytes and bronchial epithelial cells are cytotoxic to single-walled carbon nanotubes, forming ROS and mitochondrial dysfunction¹⁸⁷. According to Winnik and Maysinger, quantum dots cause cytotoxicity by boosting ROS production¹⁸⁸. According to reports, nano-ZnO damages human bronchial epithelial cells by causing them to produce more ROS¹⁸⁹. When the cytotoxic effects of nano-TiO₂, Co₃O₄, ZnO, and CuO in hepatocyte cells were evaluated, it was observed that nano-CuO had the highest cytotoxic effect. Nano-FeO was shown to have a cytotoxic effect via enhancing ROS production and apoptosis^{190,191}. Nanomaterial toxicity is affected by various parameters, including surface area, surface coating, molecular size and shape, oxidation status, solubility, and the degree of aggregation and agglomeration¹⁹². It has been found that the size of the nanoparticles directly affects how dangerous they are. Amorphous nano-silica is hazardous to human cells, according to Yoshida et al., since it increases the production of ROS and damages DNA^{38,193}. Additionally, based only on size, nanoparticles are more harmful to organs the smaller they are¹⁹⁴. According to studies, the formation of ROS by wire-shaped nanoparticles damages DNA and has harmful effects¹⁹⁵. Studies on the relationship between nanomaterial shape and toxicity have found that the shape does not significantly affect the toxicity of nano Au in human skin keratinocyte cells¹⁹⁶. In contrast, hexagonal crystals are more hazardous than rod-shaped crystals, according to a study on

nano-ZnO crystals¹⁹⁷. Biocompatibility and nanoparticle contact area are closely proportional. In a study of zebrafish embryos, Ispas et al. found that dendritic zebrafish embryos were more hazardous than spherical ones¹⁹⁸. A typical nanomaterial utilized in medication delivery systems is silica. Nano-silica has various harmful effects at different pore volumes¹⁹⁸. The cationic-charged nano silica-titanium particles are extremely poisonous, according to Oh et al.^{188,199}—studies on the dimensions, form, and association of the surface components of quantum dots with nanotoxicity^{188,199}. In investigations on the toxicity of fullerene, the groups attached to the surfaces of these nanoparticles are crucial. Given that fullerenes are thought to produce free oxygen radicals, which are thought to cause cytotoxicity, there are fullerenes with antioxidant activity by adding malonyl groups to their surface¹⁹². The impact of a nanomaterial's solubility on toxicity has been studied. ZnO nanoparticles are less hazardous than soluble copper metal, claim Studer et al.¹⁹⁹. Shen et al. found that dissolving nano-ZnO cells is useful for bringing about the cytotoxic impact²⁰⁰. According to Mahto et al.²⁰¹, When quantum dots are dissolved in water, more ROS are produced, which results in cytotoxicity. Nano-TiO₂ and nano-ZnO materials are negatively impacted by UV and visible light. It is how toxicity is caused by photoexcitation using electrons²⁰². Studies on the toxicity of graphene and aggregation have been carried out in various biological sectors, including drug delivery systems, biosensors, and labelling²⁰³. In addition, Kim et al.¹⁸¹ highlighted the significance of aggregation and accumulation in the toxicity caused by nano-Ag. It is still being researched in toxicity tests on various organisms, including plants, rodents, and people. In engineering, metallic and carbon nanomaterials are frequently used in various applications. Additionally, metal nanoparticles are frequently applied in food, medicine, and cosmetics²⁰⁴. Depending on how often they are used, sun creams and lotions containing nano titanium and nano zinc can harm the skin and the environment²⁰⁵. Researchers have shown that carbon nanotubes harm cells and that nano copper oxide is effective in cytotoxicity and DNA damage^{206,207}.

10.1. Effect of Metal Oxide Nanoparticles in Zebrafish

Table 4: Properties and applications of mostly used metal oxide nanoparticles

| Metal oxide nanoparticles | Physical and chemical properties | Potential applications in medicine (tested <i>in vitro/in vivo</i>) | Biomedical and applications in life science (in use and commercial products) |
|--|--|--|---|
| Aluminium oxide ²⁰⁸ (Al ₂ O ₃) | high melting point, strong corrosion resistance, high melting stability, and high thermal and mechanical stability. | Drug delivery. | — |
| Copper oxide ^{209,210} (CuO) | Catalyst and high-temperature superconductors Click or tap here to enter text. | Anticancer treatment. | Antimicrobial coating agents. |
| Iron oxide ^{209,211} (α -Fe ₂ O ₃ , γ -Fe ₂ O ₃ , and Fe ₃ O ₄) | Superparamagnetic and magnetic hyperthermia properties, catalyst. | Antibacterial agent, drug delivery, anticancer treatment (photothermal therapy, chemotherapy, and magnetic hyperthermia therapy), theragnostic (near-infrared imaging and positron emission tomography, single-photon emission computed tomography, and ultrasound imaging). | Iron-deficient anemia treatment (Venofer®, Feraheme®, and Rienso®). Solid tumor treatment (NanoTherm®). Magnetic resonance imaging (in liver: Feridex I.V.®, Endorem®, and Resovist®; in gastrointestinal: Gastromark™ and Lumirem®; and in blood pooling: Supravist®). |
| Magnesium oxide ^{208,209} (MgO) | High ionic character, catalyst, and semiconductor. | Antibacterial agent and anticancer treatment (hyperthermia therapy) and tissue engineering. | Antimicrobial agents (in the food industry). |
| Nickel oxide ²¹⁰ (NiO) | Catalyst, magnetic properties, and high electrochemical stability. | Anticancer treatment (cytotoxic properties). | — |
| Silica dioxide ²¹² (SiO ₂) | Low density. | Antibacterial agent, drug and gene delivery, anticancer treatment, and biosensor. | Additive in drugs and cosmetics. |
| Titanium oxide ²¹³ (TiO ₂) | Semiconductor, photocatalyst, and high chemical stability. | Anticancer treatment (photodynamic, photothermal, so no dynamic therapy, chemodynamic therapy, and radiotherapy), theragnostic (bioimaging), drug delivery, and tissue engineering. | UV-A and UV-B radiation filters (in sunscreens, cosmetics). Antimicrobial agents (in food packaging and biomedical devices and dentistry & orthopedic implants). |
| Zinc oxide ²¹⁴ (ZnO) | In semiconductor, photocatalyst has high chemical stability, large exciton binding energy, and high isoelectric point. | Anticancer treatment (photodynamic, photothermal, and sonodynamic therapy), theragnostic (bioimaging), drug delivery, and tissue engineering. | UV-A and UV-B radiation filters (in sunscreens, cosmetics). Antimicrobial agents (in toothpaste, dental implants, food packaging, and as a food additive). |

Table 5: Impact of IO NPs on zebrafish

| Stage | NP diameter | Treatment time | Tested concentrations | General toxicity response | Specific ROS responses |
|------------------------|-------------|----------------|-------------------------------------|---|------------------------|
| Embryos ²¹⁵ | 22 nm | 144 h | 0.3; 0.6; 1.25; 2.5; 5; and 10 mg/L | High mortality rate and cardiotoxicity (reduction of heartbeat rate) and morphological alterations. | — |
| Embryos ²¹⁶ | 6-12 nm | 120 hpf | SP IONs, S PION-DX, SP ION-CS, | SP ION-CS: reduced survival rate, SPI ON- | — |

| | | | | | |
|-------------------------------|------------|------------------------------------|---|--|---|
| | | | SP ION-T, SPI ON-T-PEG, SP ION@SiO ₂ : 0.125 mM, 0.5 mM, 2.0 mM, and 8.0 mM | CS, and SP ION@SiO ₂ delay in hatching rate; SP ION-DX, SP ION-T-PEG, and SP ION-T: slightly premature hatching; SP ION-CS and SPI ON@SiO ₂ : reduction in locomotor activity; and SP ION-CS, SP ION-T-PEG SP ION@SiO ₂ reduction in escape behavior. | |
| Embryos ²¹⁷ | | 168 hpf | 0.1, 0.5, 1 and 5, 10, 50, and 100 mg/L | Mortality concentration and exposure time-dependent; LC50 = 53.35 mg/L; delay in hatching rate, LC50 = 36.06 mg/L; and different malformations (pericardial edema and tissue ulceration and body arcuation). | — |
| Embryos ²¹⁸ | 40 nm | 96 h | Fe ₃ O ₄ NPs: 100-800 µg/mL bare Cr@Fe ₃ O ₄ : 5, 150, 300, and 600 mg/mL | Fe ₃ O ₄ NPs: dose- and time-dependent delay in hatching rate; slight decrease in embryo viability; Cr@Fe ₃ O ₄ : NPs high mortality in 2-week-old larvae; dose-dependent accumulation in the digestive tract. | — |
| Embryos ²¹⁹ | 100-250 nm | 168 hpf | 1, 5, 10, 50, and 100 mg/L | LC50 = 10 mg/L; delay in the hatching rate. | — |
| Embryos ²¹⁵ | 22-45 nm | 96 hpf | 10, 20, 40, 60, 80, 110, 120, 140 ppm | LC50 = 60.17 ppm; delay in hatching rate; reduction in heartbeat rate; and increased teratogenicity. | Dose-dependent decrease of Na ⁺ K ⁺ -ATPase activity; the dose-dependent increase of AChE; increased levels of lipid peroxidation ROS, PC, and NO; an increase of apoptotic bodies; and a decrease of antioxidant enzymes, CAT, SOD, and Gpx. |
| Embryos/adults ²²⁰ | 15 nm | Embryos: 96 hpf Adults: 2 weeks | Embryos: 1, 10, 100, and 1000 ppm Adults: 1, 10 ppm | Embryos: no adverse effect observed Adults: reduced locomotor and exploration activity, increased anxiety, reduced social interaction, tightened shoaling behavior, dysregulation of circadian rhythm locomotor activity and reduction of short-term memory retention, and reduction of serotonin and dopamine. | Increased CAT, cortisol level in the brain; reduction of AChE activity. |
| Adults ²²¹ | 21 nm | 7 days | 100 mg/L | Bare IO NPs accumulate mainly in | Altered expression of genes involved in inflammation, |

| | | | | | |
|-----------------------|--|---------|--------------------------------|---|---|
| | | | | the gills, and coated IO NPs in the liver. | immune response, oxidative stress, antioxidant response, and mitochondria in the gills of Fe ₃ O ₄ -treated fish. Upregulation in the liver of genes involved in immune and inflammation responses and downregulation of genes involved in DNA damage and repair in both exposures and different expression of genes involved in DNA damage/repair and apoptosis (<i>tp53</i>) for starch-coated NPs and upregulation of <i>cyp1a</i> ; and dysregulation of genes involved in the mitochondrial dysfunction pathway. |
| Adults ²¹⁶ | Fe ₂ O ₃ : 80-90 nm Fe ₃ O ₄ : 140-160 nm | 28 days | 4 and 10 mg/L | Shift in coloration, extravasated blood, and chronic toxicity in the gut. | — |
| Adults ²²² | 23 nm | 48 h | 20, 50, 100, 140 and 200 mg/kg | Reduction of AChE activity; impaired swimming. | Increased expression of transcriptional <i>jun</i> , <i>caspase-8</i> , <i>caspase-9</i> , <i>gclc</i> , <i>Gpx1a</i> , <i>CAT</i> , <i>gstp1</i> , and <i>sod2</i> . |

Table 6: Impact of ZnO NPs on zebrafish.

| Stage | NP diameter | Treatment time | Tested concentrations | General toxicity response | Specific ROS responses |
|------------------------|-------------|----------------|--|---|---|
| Embryos ²²³ | 20 nm | 96 h | 0.1, 0.5, 1, 5, 10, and 50 mg/L | Significant decrease of survival rate and delay in hatching rate dose-dependent; 96 h LC50 = 1.793 mg/L; and several abnormalities (body accusation and pericardial edema). | — |
| Embryos ²²⁴ | 20 nm | 96 hpf | 0.1, 0.5, 1, 5, 10 and 50 and 100 mg/L | Decrease of survival rate and delay in hatching rate and incidence of pericardial edema dose-dependent. | Increase in ROS production, low levels of <i>Gstp2</i> and <i>Nqo1</i> expressions, and a downfall in counteracting the ROS by oxidative stress responses. |
| Embryos ²⁰⁸ | <100 nm | 144 hpf | 1, 5, 10, 20, 50, and 100 mg/L | No effect on the survival rate, a significant decrease in the hatching rate, and different malformations (spinal curvature and hyperemia). | Important elevation in the SOD activity and MDA levels in a dose-dependent way; decrease in CAT activity; high levels of ROS; DNA damage only at the highest concentration tested; and important downregulation in <i>Bcl-2</i> , <i>Nqo1</i> , and <i>Gstp2</i> transcriptions and upregulation in <i>Ucp-2</i> level. |
| Embryos ²²⁵ | 30 nm | 96 hpf | 1, 5, 10, 25, 50, and 100 mg/L | Decrease in survival rate and increase in hatching rate dose-dependent; severe decrease in body length. | — |
| Embryos ²¹⁴ | <100 nm | 96 hpf | 1, 5, 10, 20, 50, and 100 mg/L | — | Increase in the lipid peroxidation and SOD activity; upregulation in the expression of |

| | | | | | |
|------------------------|--|---------|-----------------------------|--|--|
| | | | | | the <i>ppax</i> and <i>sod1</i> ; downregulation of <i>cat</i> ; altered expression of antiapoptotic genes (<i>bcl-2</i>) and proapoptotic (<i>Bax</i> , <i>puma</i> , and <i>apaf-1</i>); upregulation of <i>p53</i> gene, with overexpression of its protein; and increase in the activity of caspase-3 and caspase-9. |
| Embryos ²²⁶ | 9.4 nm | 96 hpf | 0.2, 1, and 5 mg/L | Dramatic delay in hatching. | Upregulation of the <i>cat</i> and Cu/Zn-sod transcripts in embryos and downregulation in eleuthero; important upregulation of Mt2<; different expression of mRNA of <i>IL-1β</i> , <i>TNFα</i> , and proinflammatory cytokines in eleuthero-embryos in comparison to embryos; alteration in the <i>jun</i> proto-oncogene (<i>c-jun</i>) embryos treated with high concentration; and perturbation in antiviral and immune-related gene Myxovirus resistance A. |
| Embryos ²²⁷ | 50–70 nm | 144 hpf | 0.1, 0.5, 1, 5, and 10 mg/L | Significant delay in hatching for ZnO NPs and Zn ions; no significant difference in cotreatment with ZnO NPs and NAC; and increased rates of delay in hatching in cotreatment with BSO. | ROS generation; cotreatment with BSO: lower production of GSH. |
| Embryos ²²⁸ | Nanospheres: 27 nm; nano sticks: 32×81 nmM; and SMPs: 202 nm | 120 hpf | 2, 4, 8, 16, and 32 mg Zn/L | LC50 for Zn ²⁺ =7.9 mg Zn/L, LC50 ZnO SMPs =10.0 mg Zn/L LC50 nano sticks =7.1 Zn/L LC50 nanospheres =11.9 mg Zn/L, respectively; higher toxicity of Zn ions compared to the different shaped NPs; and decrease of hatching rate dose-dependent in the embryos treated with all the different kinds of nanoparticles and sulfate, strongest delay in samples exposed to nano sticks. Decrease dose-dependent of swimming activity; nano sticks are more toxic than the other NPs. | — |
| Embryos ²²⁹ | 5, 10, 15, 26, 34, 62, and 70 nm | 120 hpf | 0.016 to 250 mg/L | Significant mortality at 24 hpf for all the coated NPs; no alteration in mortality with bare nanoparticles. | — |

| | | | | | |
|------------------------|----------|--------|---------------------------|---|---|
| Embryos ²²⁵ | 20-30 nm | 96 hpf | 0.01, 0.1, 1, and 10 mg/L | Higher mortality rate by ZnO NPs than ZnSO ₄ ; LC25 for ZnO NPs = 2.64 mg/L; LC25 for ZnSO ₄ = 7.75 mg/L; and significant embryonic malformations after both treatments (tail malformation, pericardial edema, and yolk sac edema). | Downregulation of <i>ogfr12</i> and <i>intl2</i> transcripts; upregulation of <i>cyb5d1</i> . |
|------------------------|----------|--------|---------------------------|---|---|

Table 7: Some of the main physicochemical properties of nanoparticles, as well as the exposure routes and main findings on various animal models

| Animal Model | Administration route and exposure time | Nanoparticle | Surface Chemistry | Size/nm | Major observations |
|-------------------------------|---|--------------------------------------|---|-------------------------------|--|
| Mouse ²³⁰ | i.p. and i.v. injection, 1, 4, 24 h | Gold | Without surface modification | 2,40 | Macrophage uptake in the liver is less in the spleen, small intestine, and lymph nodes. |
| Rat ²³¹ | i.v. injection, 24 h | Gold | Without surface modification | 10-205 | NPs of 10 nm entered the testis and brain. |
| Mouse ²³² | i.v. Injection, 0.5,2, and 24 h. | MWCNTs | Carboxylated and aminated surface | 20-30 × 0.5-2 mm | Accumulation in testis. |
| Mouse ²³³ | i.v. injection, 0.17, 1, and 24 h | SWCNTs | Without or coated by paclitaxel (PTX)-polyethylene glycol (PEG) | 1-3 × 100 (diameter × length) | Accumulation in liver and spleen, less in the heart, lung, kidney, stomach, intestine, muscle. |
| Rat ¹³⁶ | Whole body inhalation 12 days | MnO ₂ | Without surface modification | 30 | Accumulation in CNS via olfactory bulb. |
| Pig ²³⁴ | Intradermal injection <5 min | CdTe (CdSe) core (shell) type II QDs | Oligomeric, Phosphine | 10 (naked); 18.8 (coated) | Accumulation in the sentinel lymph node. |
| Rat ²³⁵ | Gavage | Polystyrene microspheres | Without surface modification | 50, 100, and, 300 | Accumulation in the liver and spleen via lymph. |
| Mouse ²³⁶ | Intranasal instillation, 2, 10, 20, and 30 days | TiO ₂ | Without surface modification | 10, 25, and 60 | Accumulation in brain through the olfactory bulb. |
| Hairless Mouse ²³⁷ | Dorsal skin expos | TiO ₂ | Hydrophobic or hydrophilic surface | 80, 155 | Accumulation in the spleen, lung, kidney, and brain. |

II. TOXICITY TESTING

Dosing concerns are crucial in determining toxicity, and in vitro, tests are more frequent than in vivo research. One of the models utilized in the toxicity test is the in vitro sedimentation diffusion and dosimeter. This model's core concept is the fundamental separation between exposure (concentration in the cell environment), dose accumulated on the cell surface, and cellular dosage. By being aware of how long it takes for a given dose to be released, we may assess the dose rate as a predictor of response⁶². Because in vitro techniques that assess cell viability and proliferation are widely

used, gene expression analysis, genotoxicity detection, and in vitro hemolysis are also used to diagnose toxicity. Additional techniques for assessing the physicochemical structure of the cell include scanning electron microscopy/energy dispersive X-ray spectroscopy (SEM-EDX), transmission electron microscopy (TEM), atomic force microscopy (AFM), video-enhanced differential interference contrast (VEDIC) microscopy, and fluorescence spectroscopy. The combination of these tests makes it simpler to identify nanotoxicity²³⁸. Current toxicity experiments, their intended use, and the tested nanomaterials are all summarised briefly in the table below.

Table 8: A summary of literature-related toxicity tests of nanomaterials.

| Toxicity test | Purpose | Nanomaterials |
|--|---|--|
| Transmission electron microscopy | Determination of intracellular localization | TiO ₂ , silver, fullerene ²³⁹⁻²⁴¹ |
| Light microscopy | Physicochemical properties | Singled walled carbon nanotubes, silver ^{240,242} |
| Hemoglobin estimation | Homolysis | SiO ₂ ²⁴³ |
| Micronucleus test | Genotoxicity | Different types of nanoparticles ²⁴⁴ |
| Commet assay test | DNA damage | Metal, metal oxide nanoparticles ²⁴⁵ |
| Lactate dehydrogenase | Cell viability | Carbon nanoparticles ^{246,247} |
| Tetrazolium salts | | Carbon nanoparticles, fullerenes ^{248,249} |
| Alamar Blue | | Quantum dots ⁷¹ |
| Propidium iodide | | Carbon nanoparticles ^{58,250} |
| Neutral red assay test | | Carbon nanotubes ^{248,251} |
| Caspase-3 activity | Apoptosis | Silver nanoparticles ²⁴⁰ |
| Acridine orange/ethidium bromide | | Silver nanoparticles ²⁵² |
| ROS production | Oxidative stress | TiO ₂ ²³⁹ |
| Levels of glutathione peroxidase, catalase, superoxide dismutase | | Polymeric nanoparticles ²⁵³ |
| Lipid peroxidation, vitamin | | Singled walled carbon nanotubes ¹⁸⁷ |

Lung injury from nanoparticle exposure through the respiratory tract is common. Therefore, organ-on-a-chip research has become more significant in recent years, and many studies have been undertaken to establish the detection of lung toxicity. By more accurately simulating human reactions with the chip in a 3D human lung model that simulated in vivo settings, Zhang et al. explored nanotoxicity. Using accurate models, this study further illustrated the importance of organ-based toxicity²⁵⁴. According to studies, nanoparticles have a harmful effect after passing through the placenta of mice. In the 3D human placenta model, chip and TiO₂ nanoparticle exposure studies may have similar harmful consequences, claim Yin et al.²⁵⁵. Additionally, research on nanotoxicity was conducted using a cell-on-a-chip (CoC) and a microfluidic system²⁵⁶.

12. REGULATORY CHALLENGES

12.1. Importance of Nanomedicines in the Pharmaceutical Market

Over the last two to three decades means the last 20-30 years, the successful introduction of nanomedicine in both clinical practice and the continuous development in pharmaceutical research has created more sophisticated ones which are mainly entering clinical trials. The nanomedicine market in European Union is composed mainly of nanoparticles, liposomes, nanocrystals, nanoemulsions, polymeric-protein conjugates, and nano complexes²⁵⁷. There are currently available nanomedicines made and approved by the EU (European Union)²⁵⁸.

12.2. Nanomedicines and Nanosimilars

In the approval process, nanomedicines were introduced under the traditional benefit or risk analysis framework. Another challenge related to the framework is developing a framework mainly for evaluating the follow-on nanomedicines at the time of reference medicine patent expiration²⁵⁹. Nanomedicine is comprised of both biological and non-biological medical products. Biological nanomedicines are obtained mainly from biological sources. At the same time, the

non-biological products are mentioned as non-biological complex drugs (NBCD), where we can find that the active principle consists of different structures²⁶⁰. In introducing generic medicines in the pharmaceutical market, we must demonstrate several parameters, as described elsewhere. A more complete analysis is needed for biological and non-biological nanomedicines, which mainly go beyond the plasma concentration measurement. The therapeutic equivalence and, consequently, interchangeability can be requireable by a stepwise comparison of bioequivalence, safety, and efficacy and this relation to the related medicine²⁶¹. The biological nanomedicines are under the regulatory framework set by the European Medicines Agency (EMA)¹. This framework is an approach to the regulatory system for follow-on biological nanomedicines, which includes the recommendations for the comparative quality, clinical and non-clinical studies²⁶². The regulatory approach for the follow-on "Non-Biological Complex Drugs (NCBD)" is still a process. The industry frequently asks for scientific advice, and the EMA analyzes a case-by-case analysis. Sometimes, the biological framework is the basis for the regulation of the "Non-Biological Complex Drugs (NCBDs)" because they have some common features: the structure cannot be fully characterized, and the *in-vivo* activity is dependent on the process of manufacturing, and consequently, the comparability needs to establish throughout the life cycle, as happens to the biological nanomedicines. Besides this, for some "Non-Biological Complex Drugs (NCBDs)" groups like glatiramer, liposomes, and iron carbohydrate complexes, there are draft regulatory approaches, which may help the regulatory authorities or regulatory bodies to create a final framework for the different "Non-Biological Complex Drugs (NCBDs)" families²⁶³. EMA has already released some papers regarding nanomedicines with a surface coating, block copolymer micelle, intravenous liposomal, and iron-based nano colloidal nanomedicines²⁶⁴. These papers released by the EMA are applied to new nanomedicines and nanosimilars, guiding developers in preparing marketing authorization applications. The principles outlined in these documents address general issues that are regarding the complexity of these nanosystems and provide basic information for the development of the pharmaceutical industry, both the non-clinical and early clinical studies of the

block-copolymer micelle, "liposome-like" and the nanoparticle iron (NPI) medicinal products mainly the drug products that have been created to affect the pharmacokinetics, distribution, and stability of incorporated or conjugated active substances *in vivo*. The important factors are mainly related to the exact nature of the characteristics of the particle, and that can influence the kinetic parameters and, consequently, the toxicity, such as the physicochemical nature of the coating, the stability, and respective uniformity (both in terms of susceptibility to degradation), the bio-distribution of the product and its intracellular fate are especially detailed.

12.3. Market Access and Pharmacokinetics

After obtaining nanomedicine by marketing authorization, there is a long way up to the introduction of nanomedicine in clinical practice or clinical trials in all the European Union countries. It occurs because of the reimbursement and pricing decisions for medicines taken at an individual level in each member state of the European Union (EU)²⁶⁴. In case to provide patients access to medicines, the multidisciplinary process provided by Health Technology Assessment (HTA) is being developed. The Health Technology Assessment HDT generates information about effectiveness, medicine safety, and cost-effectiveness to support the health and political decision-maker²⁶⁴. The study of pharmacoeconomics assumes a crucial role before the commercialization of nanomedicines at the current time. They mainly assess the economic and social importance through the added therapeutic value using indicators such as quality-adjusted life expectancy years and hospitalization²⁶⁴. To harmonize and enhance the entry of new medicines into the clinical trial, they have created the EUnetHTA to provide patients with novel medicines. The main goal of EUnetHTA is to develop decisive, appropriate transport information to help the HTAs in European Union countries.

13. ARGUMENTS FOR NANO-SPECIFIC TOXICITY

It is appropriate to mention that in contrast to the view taken in the published literature, nanoparticles do have nano-specific effects²⁶⁵. For example, Krug and Wick³² refer to surface composition, size, and transport as the factors that contribute to the toxicity of any nanoparticle²⁶⁵. They suggest that for any specific nanoparticle, these three factors come together to form a unique combination forming²⁶⁵ '... a basis for the description of specific reactions and interactions between nanomaterials/nanoobjects and biological systems...'³². These authors argue the obvious result of this contention, namely that each nanoparticle 'must be tested individually'³². We reject a 'counsel of despair' above²⁶⁵, arguing that the lack of nano-specific toxicity forms a basis for benchmarking the large amount of available data on conventional particle-mediated pathogenicity²⁶⁵. We note that the final common pathways for pathological effects, oxidative stress²⁶⁵, inflammation, and genotoxicity, are entirely shared by both nanoparticles and conventional particles, and no novel pathogenic pathways are anticipated²⁶⁵. Therefore while the proximate events such as the transport of nanoparticles into cells may be unusual or even novel²⁶⁶, the final common pathways of oxidative stress inflammation and genotoxicity are impacted by all pathogenic

particles²⁶⁵. Therefore, we can see no reason to invoke nano-specificity to the adverse effects, nor should we anticipate novel pathologies²⁶⁵. Kreyling has demonstrated that the translocation of NP from the lungs varies depending on the nanoparticle size²⁶⁷, with a greater fractional translocation of the smaller nanoparticles²⁶⁷. However, the translocation fraction is extremely small and so of questionable significance²⁶⁵. That it is not significant is supported by the striking absence of reports of extra-pulmonary pathology in many chronic²⁶⁸, high exposure, rat inhalation studies carried out with low solubility, low toxicity nanoparticles in the eighties and nineties, for example²⁶⁸. In the case of human epidemiology of ambient combustion-derived nanoparticles (air pollution/PM) exposure, the only clear extra-pulmonary effects — in cardiovascular disease — are now considered most likely to arise from oxidative stress or inflammatory signals from the lungs. However, translocation is not completely ruled out²⁶⁹.

14. CONCLUSION

There is a huge amount of research and regulatory activity in nanoparticle health and safety. Toxicologists need to comprehensively understand this hazard in the context of varying composition, shape, and size for use in risk assessment. It is very important as the sheer degree of adaptability and variability of engineered nanoparticles against detailed testing of every form produced, so other judgments from other sources as to potential toxicity or mechanism of toxicity of nanomaterials are required. Current research shows that exposure to nanoparticles when administered in high concentrations, can cause severe adverse effects, as shown in zebrafish. TiO₂ NPs, IO NPs, and ZnO NPs are considered nontoxic and widely approved but can also show harmful effects. ZnO NPs cause an increase in the reactive O₂ in response to fluorescent light. ZnO NP increases ROS, which stimulates the apoptotic pathways regulated by caspases and mitochondria, which causes extensive cellular dysfunction even at a lower concentration. IO NPs are associated with oxidative stress and induction of redox-signal pathways(AP); NP size and coating seem to cause cellular dysfunction. Further research is needed to unravel the mechanism of nanotoxicity due to nanoparticles.

15. ACKNOWLEDGEMENT

We acknowledge Guru Nanak Institute of Pharmaceutical Science and Technology for providing us with the infrastructure and support system.

16. AUTHORS CONTRIBUTION STATEMENT

The authors of this review article, Pramit Sahoo, Pritam Roy, and Seabrata Bhakta, contributed equally to the article's conception, research, and writing. In addition, Jeenatara Begum and Tamalika Chakraborty provided critical feedback and supervised the project. All authors have reviewed and approved the final version of the manuscript.

17. CONFLICT OF INTEREST

Conflict of interest declared none.

18. REFERENCES

1. Hobson DW. Commercialization of nanotechnology. *Wiley Interdiscip Rev Nanomed Nanobiotechnol.* 2009;1(2):189-202. doi 10.1002/WNAN.28, PMID 20049790.
2. Bowman DM. More than a decade of mapping today's regulatory and policy landscapes following the publication of *Nanoscience and Nanotechnologies: opportunities and Uncertainties.* *NanoEthics.* 2017;11(2):169-86. doi: 10.1007/S11569-017-0281-X.
3. Nanotechnologies—terminology ISO. Definitions for nano-objects—nanoparticle, nanofibre, and nanoplate. International Organization for Standardization; 2008.
4. OECD. O for EC and D. Inhalation toxicity testing: expert meeting on potential revisions to OECD test guidelines and guidance document. Series on the Safety of Manufactured Nanomaterials. Vol. 35; 2012.
5. Executive H and S. Risk Management of Carbon Nanotubes. 2009.
6. Aitken RA, Bassan A, Friedrichs S, et al. Specific advice on fulfilling information requirements for nanomaterials under REACH (RIP-on2); 2011.
7. Aitken RA, Bassan A, Friedrichs S, et al. Specific advice on exposure assessment and hazard/risk characterization for nanomaterials under REACH (RIP-on 3)-final project report; 2011.
8. Auffan M, Rose J, Bottero JY, Lowry GV, Jolivet JP, Wiesner MR. Towards a definition of inorganic nanoparticles from an environmental, health, and safety perspective. *Nat Nanotechnol.* 2009;4(10):634-41. doi 10.1038/nano.2009.242, PMID 19809453.
9. Fubini B, Ghiazza M, Fenoglio I. Physico-chemical features of engineered nanoparticles relevant to their toxicity. <http://dx.doi.org/103109/174353902010509519>. *Nanotoxicology.* 2010;4(4):347-63. doi: 10.3109/17435390.2010.509519, PMID 20858045.
10. Potocnik J. Commission recommendation of 18 October 2011 on the definition of nanomaterial. *Off J Eur Commun Legis.* 2011;275:38-40.
11. Norppa H, Catalań J, Falck G, Hannukainen K, Siivola K, Savolainen K. Nano-specific genotoxic effects. *J Biomed Nanotechnol.* 2011;7(1):19. doi: 10.1166/JBN.2011.1179, PMID 21485781.
12. Donaldson K, Schinwald A, Murphy F, Cho WS, Duffin R, Tran L, et al. The biologically effective dose in inhalation nanotoxicology. *Acc Chem Res.* 2013;46(3):723-32. doi: 10.1021/ar300092y, PMID 23003923.
13. Soares S, Sousa J, Pais A, Vitorino C. Nanomedicine: principles, properties, and regulatory issues. *Front Chem.* 2018;6:360. doi: 10.3389/fchem.2018.00360, PMID 30177965.
14. Anonymous. Commission recommendation of 18 October 2011 on the definition of nanomaterial text with EEA relevance – publications office of the EU; n.d. [cited 9/8/2022] Available from: <https://op.europa.eu/en/publication-detail/-/publication/17af73d9-da70-4a46-a421-c62e3d1df6ce/language-en>.
15. Anonymous. Considering whether an FDA-regulated product involves the application of nanotechnology | FDA; n.d. [cited 9/8/2022] Available from: <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/considering-whether-fda-regulated-product-involves-application-nanotechnology>.
16. Bleeker EAJ, de Jong WH, Geertsma RE, Groenewold M, Heugens EH, Koers-Jacquemijns M, et al. Considerations on the EU definition of a nanomaterial: science to support policy making. *Regul Toxicol Pharmacol.* 2013;65(1):119-25. doi: 10.1016/J.YRTPH.2012.11.007, PMID 23200793.
17. Boverhof DR, Bramante CM, Butala JH, Clancy SF, Lafranconi M, West J, et al. Comparative assessment of nanomaterial definitions and safety evaluation considerations. *Regul Toxicol Pharmacol.* 2015;73(1):137-50. doi: 10.1016/J.YRTPH.2015.06.001, PMID 26111608.
18. Seabra AB, Durán N. Nanotoxicology of metal oxide nanoparticles. *Metals.* 2015;5(2):934-75. doi: 10.3390/MET5020934.
19. Schrand AM, Rahman MF, Hussain SM, Schlager JJ, Smith DA, Syed AF. Metal-based nanoparticles and their toxicity assessment. *Wiley Interdiscip Rev Nanomed Nanobiotechnol.* 2010;2(5):544-68. doi: 10.1002/WNAN.103, PMID 20681021.
20. Iijima S, Ichihashi T. Single-shell carbon nanotubes of 1-nm diameter. *Nature.* 1993;363(6430):603-5. doi: 10.1038/363603a0.
21. Bethune DS, Kiang CH, de Vries MS, Gorman G, Savoy R, Vazquez J, et al. Cobalt-catalyzed growth of carbon nanotubes with single-atomic-layer walls. *Nature.* 1993;363(6430):605-7. doi 10.1038/363605a0.
22. Tans SJ, Devoret MH, Dai H, Thess A, Smalley RE, Geerligs LJ, et al. Individual single-wall carbon nanotubes as quantum wires. *Nature.* 1997;386(6624):474-7. doi 10.1038/386474a0.
23. Yu MF, Lourie O, Dyer MJ, Moloni K, Kelly TF, Ruoff RS. Strength and breaking mechanism of multiwalled carbon nanotubes under tensile load. *Science.* 2000;287(5453):637-40. doi 10.1126/science.287.5453.637. PMID 10649994.
24. Berber S, Kwon YK, Tománek D. Unusually high thermal conductivity of carbon nanotubes. *Phys Rev Lett.* 2000;84(20):4613-6. doi: 10.1103/PhysRevLett.84.4613, PMID 10990753.
25. Akçan R, Aydoğan HC, Yildirim MŞ, Taştekin B, Sağlam N. Nanotoxicity: a challenge for future medicine. *Turk J Med Sci.* 2020;50(4):1180-96. doi: 10.3906/SAG-1912-209, PMID 32283898.
26. Gurevitch D, Shuster-Meiseles T, Nov O, Zick Y, Rudich A, Rudich Y. TiO₂ nanoparticles induce insulin resistance in liver-derived cells directly and via macrophage activation. *Nanotoxicology.* 2012;6(8):804-12. doi: 10.3109/17435390.2011.625128, PMID 22007682.
27. Mohammadipour A, Fazel A, Haghiri H, Motejaded F, Rafatpanah H, Zabihi H, et al. Maternal exposure to titanium dioxide nanoparticles during pregnancy; impaired memory and decreased hippocampal cell proliferation in rat offspring. *Environ Toxicol Pharmacol.* 2014;37(2):617-25. doi: 10.1016/J.ETAP.2014.01.014, PMID 24577229.
28. Willhite CC, Karyakina NA, Yokel RA, Yenugadhati N, Wisniewski TM, Arnold IM, et al. A systematic review of potential health risks posed by pharmaceutical, occupational, and consumer exposures to metallic and nanoscale aluminum, aluminum oxides, aluminum

- hydroxide, and its soluble salts. *Crit Rev Toxicol*. 2014;44:Suppl 4;Suppl 4(Suppl 4):1-80:1-80. doi: 10.3109/10408444.2014.934439, PMID 25233067.
29. Blum JL, Xiong JQ, Hoffman C, Zelikoff JT. Cadmium associated with inhaled cadmium oxide nanoparticles impacts fetal and neonatal development and growth. *Toxicol Sci*. 2012;126(2):478-86. doi: 10.1093/TOXSCI/KFS008, PMID 22240978.
 30. Tripathi SK, Kaur G, Khurana RK, Kapoor S, Singh B. Quantum dots and their potential role in cancer theranostics. *Crit Rev Ther Drug Carrier Syst*. 2015;32(6):461-502. doi: 10.1615/CRITREVTHERDRUGCARRIERSYST.2015012360, PMID 26559550.
 31. Luo G, Long J, Zhang B, Liu C, Ji S, Xu J, et al. Quantum dots in cancer therapy. *Expert Opin Drug Deliv*. 2012;9(1):47-58. doi: 10.1517/17425247.2012.638624, PMID 22171712.
 32. Donaldson K, Poland CA. Nanotoxicity: challenging the myth of nano-specific toxicity. *Curr Opin Biotechnol*. 2013;24(4):724-34. doi: 10.1016/j.copbio.2013.05.003, PMID 23768801.
 33. Povey A. Molecular assessment of exposure, effect, and effect modification. *Epidemiology of work-related diseases*. 2nd ed; 2008. p. 463-84. doi: 10.1002/9780470695005.CH21.
 34. Donaldson K, Schinwald A, Murphy F, Cho WS, Duffin R, Tran L, et al. The biologically effective dose in inhalation nanotoxicology. *Acc Chem Res*. 2013;46(3):723-32. doi: 10.1021/ar300092y, PMID 23003923.
 35. Civeira MS, Pinheiro RN, Gredilla A, de Vallejuelo SF, Oliveira ML, Ramos CG, et al. The properties of the nano-minerals and hazardous elements: potential environmental impacts of Brazilian coal waste fire. *Sci Total Environ*. 2016;544:892-900. doi: 10.1016/j.scitotenv.2015.12.026, PMID 26706762.
 36. Leo BF, Chen S, Kyo Y, Herpoldt KL, Terrill NJ, Dunlop IE, et al. The stability of silver nanoparticles in a model of pulmonary surfactant. *Environ Sci Technol*. 2013;47(19):11232-40. doi: 10.1021/es403377p, PMID 23988335.
 37. Buzea C, Pacheco II, Robbie K. Robbie K. Nanomaterials and nanoparticles: sources and toxicity. *Biointerphases*. 2007;2(4):MR17-71. doi: 10.1116/1.2815690, PMID 20419892.
 38. Auffan M, Rose J, Wiesner MR, Bottero JY. Chemical stability of metallic nanoparticles: a parameter controlling their potential cellular toxicity in vitro. *Environ Pollut*. 2009;157(4):1127-33. doi: 10.1016/j.envpol.2008.10.002, PMID 19013699.
 39. Tee JK, Ong CN, Bay BH, Ho HK, Leong DT. Oxidative stress by inorganic nanoparticles. *Wiley Interdiscip Rev Nanomed Nanobiotechnol*. 2016;8(3):414-38. doi: 10.1002/WNAN.1374, PMID 26359790.
 40. Sharma VK, Filip J, Zboril R, Varma RS. Natural inorganic nanoparticles – formation, fate, and toxicity in the environment. *Chem Soc Rev*. 2015;44(23):8410-23. doi: 10.1039/C5CS00236B, PMID 26435358.
 41. Chaudhuri S, Sardar S, Bagchi D, Dutta S, Debnath S, Saha P, et al. Photoinduced dynamics and toxicity of a cancer drug in the proximity of inorganic nanoparticles under visible light. *ChemPhysChem*. 2016;17(2):270-7. doi: 10.1002/CPHC.201500905, PMID 26563628.
 42. Lam CW, James JT, McCluskey R, Hunter RL. Pulmonary toxicity of single-wall carbon nanotubes in mice 7 and 90 days after intratracheal instillation. *Toxicol Sci*. 2004;77(1):126-34. doi: 10.1093/TOXSCI/KFG243, PMID 14514958.
 43. Yao M, He L, McClements DJ, Xiao H. Uptake of gold nanoparticles by intestinal epithelial cells: impact of particle size on their absorption, accumulation, and toxicity. *J Agric Food Chem*. 2015;63(36):8044-9. doi: 10.1021/ACS.JAFC.5B03242, PMID 26313743.
 44. Parker JP, Ude Z, Marmion CJ. Exploiting developments in nanotechnology for the preferential delivery of platinum-based anti-cancer agents to tumors: targeting some of the hallmarks of cancer. *Metallomics*. 2016;8(1):43-60. doi: 10.1039/C5MT00181A, PMID 26567482.
 45. Peters K, Unger RE, Kirkpatrick CJ, Gatti AM, Monari E. Effects of nano-scaled particles on endothelial cell function in vitro: studies on viability, proliferation, and inflammation. *J Mater Sci Mater Med*. 2004;15(4):321-5. doi: 10.1023/B:JMSM.0000021095.36878.1B, PMID 15332593.
 46. Grimsdale AC, Chan KL, Martin RE, Jokisz PG, Holmes AB. Synthesis of light-emitting conjugated polymers for applications in electroluminescent devices. *Chem Rev*. 2009;109(3):897-1091. doi: 10.1021/cr000013v, PMID 19228015.
 47. Niu X, Zou W, Liu C, Zhang N, Fu C. Modified nanoprecipitation method to fabricate DNA-loaded PLGA nanoparticles. *Drug Dev Ind Pharm*. 2009;35(11):1375-83. doi: 10.3109/03639040902939221, PMID 19832638.
 48. Chen Y, Wang Q, Wang T. Facile large-scale synthesis of brain-like mesoporous silica nanocomposites via selective etching. *Nanoscale*. 2015;7(39):16442-50. doi: 10.1039/C5NR04123F, PMID 26394819.
 49. TS K, S E, M E-M, et al. Improved drug loading and antibacterial activity of minocycline-loaded PLGA nanoparticles prepared by solid/oil/water ion pairing method. *Int J Nanomedicine* 2012;7:221; doi: 10.2147/IJN.S27709.
 50. Liu Y, Deng H, Xiao C, Xie C, Zhou X. Cytotoxicity of calcium rectorite micro/nanoparticles before and after organic modification. *Chem Res Toxicol*. 2014;27(8):1401-10. doi: 10.1021/TX500115P, PMID 25025490.
 51. Lehto M, Karilainen T, Róg T, Cramariuc O, Vanhala E, Tornaes J, et al. Co-exposure with fullerene may strengthen the health effects of organic industrial chemicals. *PLOS ONE*. 2014;9(12):e114490. doi: 10.1371/JOURNAL.PONE.0114490, PMID 25473947.
 52. Kim JS, Yoon TJ, Yu KN, Kim BG, Park SJ, Kim HW, et al. Toxicity and tissue distribution of magnetic nanoparticles in mice. *Toxicol Sci*. 2006;89(1):338-47. doi: 10.1093/TOXSCI/KFJ027, PMID 16237191.
 53. Lockman PR, Koziara JM, Mumper RJ, Allen DD. Nanoparticle surface charges alter blood-brain barrier integrity and permeability. *J Drug Target*. 2004;12(9-10):635-41. doi: 10.1080/10611860400015936, PMID 15621689.
 54. Alvarez-Román R, Naik A, Kalia YN, Guy RH, Fessi H. Skin penetration and distribution of polymeric nanoparticles. *J Control Release*. 2004;99(1):53-62. doi: 10.1016/j.jconrel.2004.06.015, PMID 15342180.
 55. Donaldson K, Aitken R, Tran L, Stone V, Duffin R, Forrest G, et al. Carbon nanotubes: a review of their properties about pulmonary toxicology and workplace

- safety. *Toxicol Sci.* 2006;92(1):5-22. doi: 10.1093/TOXSCI/KFJ130, PMID 16484287.
56. Kumar S, Sharma A, Tripathi B, Srivastava S, Agrawal S, Singh M, et al. Enhancement of hydrogen gas permeability in electrically aligned MWCNT-PMMA composite membranes. *Micron.* 2010;41(7):909-14. doi: 10.1016/J.MICRON.2010.05.016, PMID 20579893.
 57. Wang H, Wang J, Deng X, Sun H, Shi Z, Gu Z, et al. Biodistribution of carbon single-wall carbon nanotubes in mice. *J Nanosci Nanotechnol.* 2004;4(8):1019-24. doi: 10.1166/JNN.2004.146, PMID 15656196.
 58. Pantarotto D, Briand JP, Prato M, Bianco A. Translocation of bioactive peptides across cell membranes by carbon nanotubes. *Chem Commun (Camb).* 2004;4(1):16-7. doi: 10.1039/B311254C, PMID 14737310.
 59. Monteiro-Riviere NA, Nemanich RJ, Inman AO, Wang YY, Riviere JE. Multi-walled carbon nanotube interactions with human epidermal keratinocytes. *Toxicol Lett.* 2005;155(3):377-84. doi: 10.1016/J.TOXLET.2004.11.004, PMID 15649621.
 60. Cui D, Tian F, Ozkan CS, Wang M, Gao H. Effect of single wall carbon nanotubes on human HEK293 cells. *Toxicol Lett.* 2005;155(1):73-85. doi: 10.1016/J.TOXLET.2004.08.015, PMID 15585362.
 61. Tian F, Cui D, Schwarz H, Estrada GG, Kobayashi H. Cytotoxicity of single-wall carbon nanotubes on human fibroblasts. *Toxicol In Vitro.* 2006;20(7):1202-12. doi: 10.1016/J.TIV.2006.03.008, PMID 16697548.
 62. Bottini M, Bruckner S, Nika K, Bottini N, Bellucci S, Magrini A, et al. Multi-walled carbon nanotubes induce T lymphocyte apoptosis. *Toxicol Lett.* 2006;160(2):121-6. doi: 10.1016/J.TOXLET.2005.06.020, PMID 16125885.
 63. Jia G, Wang H, Yan L, Wang X, Pei R, Yan T, et al. Cytotoxicity of carbon nanomaterials: single-wall nanotube, multi-wall nanotube, and fullerene. *Environ Sci Technol.* 2005;39(5):1378-83. doi: 10.1021/ES048729L.
 64. Favi PM, Valencia MM, Elliott PR, Restrepo A, Gao M, Huang H, et al. Shape and surface chemistry affect the cytotoxicity and cellular uptake of metallic nanorods and nanospheres. *J Biomed Mater Res A.* 2015;103(12):3940-55. doi: 10.1002/JBM.A.35518, PMID 26053238.
 65. El-Ansary A, Al-Daihan S, ben Bacha AB, Kotb M. Toxicity of novel nanosized formulations used in medicine. *Methods Mol Biol.* 2013;1028:47-74. doi: 10.1007/978-1-62703-475-3_4, PMID 23740113.
 66. Perez JE, Contreras MF, Vilanova E, Felix LP, Margineanu MB, Luongo G, et al. Cytotoxicity and intracellular dissolution of nickel nanowires. <http://dx.doi.org/10.3109/1743539020151132343>. *Nanotoxicology.* 2016;10(7):871-80. doi: 10.3109/17435390.2015.1132343, PMID 26692167.
 67. Jeannet N, Fierz M, Schneider S, Künzi L, Baumlin N, Salathe M, et al. Acute toxicity of silver and carbon nano aerosols to normal and cystic fibrosis human bronchial epithelial cells. *Nanotoxicology.* 2016;10(3):279-91. doi: 10.3109/17435390.2015.1049233, PMID 26011645.
 68. Liu H, Liu T, Wang H, Li L, Tan L, Fu C, et al. Impact of pegylation on the biological effects and light heat conversion efficiency of gold nanoshells on silica nano rattles. *Biomaterials.* 2013;34(28):6967-75. doi: 10.1016/J.BIOMATERIALS.2013.05.059, PMID 23777913.
 69. Aschberger K, Johnston HJ, Stone V, Aitken RJ, Tran CL, Hankin SM, et al. Review of fullerene toxicity and exposure – appraisal of a human health risk assessment, based on open literature. *Regul Toxicol Pharmacol.* 2010;58(3):455-73. doi: 10.1016/J.YRTPH.2010.08.017, PMID 20800639.
 70. Saathoff JG, Inman AO, Xia XR, Riviere JE, Monteiro-Riviere NA. In vitro toxicity assessment of three hydroxylated fullerenes in human skin cells. *Toxicol Vitro.* 2011;25(8):2105-12. doi: 10.1016/J.TIV.2011.09.013.
 71. Sayes CM, Fortner JD, Guo W, Lyon D, Boyd AM, Ausman KD, et al. The differential cytotoxicity of water-soluble fullerenes. *Nano Lett.* 2004;4(10):1881-7. doi: 10.1021/nl0489586.
 72. Injac R, Prijatelj M, Strukelj B. Fullerenol nanoparticles: toxicity and antioxidant activity. *Methods Mol Biol.* 2013;1028:75-100. doi: 10.1007/978-1-62703-475-3_5, PMID 23740114.
 73. Oughton DH, Hertel-Aas T, Pellicer E, Mendoza E, Joner EJ. Neutron activation of engineered nanoparticles as a tool for tracing their environmental fate and uptake in organisms. *Environ Toxicol Chem.* 2008;27(9):1883-7. doi: 10.1897/07-578.1, PMID 19086315.
 74. Kahru A, Ivask A. Mapping the dawn of nano ecotoxicological research. *Acc Chem Res.* 2013;46(3):823-33. doi: 10.1021/ar3000212, PMID 23148404.
 75. Stone V, Nowack B, Baun A, van den Brink N, Kammer Fv, Dusinska M, et al. Nanomaterials for environmental studies: classification, reference material issues, and strategies for physico-chemical characterization. *Sci Total Environ.* 2010;408(7):1745-54. doi: 10.1016/J.SCITOTENV.2009.10.035, PMID 19903569.
 76. Kahru A, Dubourguier HC. From ecotoxicology to nanoecotoxicology. *Toxicology.* 2010;269(2-3):105-19. doi: 10.1016/J.TOX.2009.08.016, PMID 19732804.
 77. Sigg L, Behra R, Groh K, Isaacson C, Odzak N, Piccapietra F, et al. Chemical aspects of nanoparticle ecotoxicology. *Chimia (Aarau).* 2014;68(11):806-11. doi: 10.2533/chimia.2014.806, PMID 26508489.
 78. Hassellöv M, Readman JW, Ranville JF, Tiede K. Nanoparticle analysis and characterization methodologies in environmental risk assessment of engineered nanoparticles. *Ecotoxicology.* 2008;17(5):344-61. doi: 10.1007/S10646-008-0225-X, PMID 18483764.
 79. Handy RD, von der Kammer F, Lead JR, Hassellöv M, Owen R, Crane M. The ecotoxicology and chemistry of manufactured nanoparticles. *Ecotoxicology.* 2008;17(4):287-314. doi: 10.1007/S10646-008-0199-8, PMID 18351458.
 80. Rickerby DG, Morrison M. Nanotechnology and the environment: A European perspective. *Sci Technol Adv Mater.* 2007;8(1-2):19-24. doi: 10.1016/J.STAM.2006.10.002.
 81. Lv M, Huang W, Chen Z, Jiang H, Chen J, Tian Y, et al. Metabolomics techniques for nanotoxicity investigations. *Bioanalysis.* 2015;7(12):1527-44. doi: 10.4155/BIO.15.83, PMID 26168257.
 82. Gao Y, Jin B, Shen W, Sinko PJ, Xie X, Zhang H, et al. China and the United States--Global partners, competitors, and collaborators in nanotechnology

- development. *Nanomedicine*. 2016;12(1):13-9. doi: 10.1016/j.NANO.2015.09.007, PMID 26427355.
83. Spruit SL, Hoople GD, Rolfe DA. Just a cog in the machine? The individual responsibility of researchers in nanotechnology is a duty to collectivize. *Sci Eng Ethics*. 2016;22(3):871-87. doi: 10.1007/s11948-015-9718-1, PMID 26538353.
 84. Loux NT, Su YS, Hassan SM. Issues in assessing environmental exposures to manufactured nanomaterials. *Int J Environ Res Public Health*. 2011;8(9):3562-78. doi: 10.3390/IJERPH8093562, PMID 22016703.
 85. Lowry GV, Gregory KB, Apte SC, Lead JR. Transformations of nanomaterials in the environment. *Environ Sci Technol*. 2012;46(13):6893-9. doi: 10.1021/es300839e. PMID 22582927.
 86. Meesters JA, Veltman K, Hendriks AJ, van de Meent D. Environmental exposure assessment of engineered nanoparticles: why REACH needs adjustment. *Integr Environ Assess Manag*. 2013;9(3):e15-26. doi: 10.1002/IEAM.1446, PMID 23633247.
 87. Mitrano DM, Motellier S, Clavaguera S, Nowack B. Review of nanomaterial aging and transformations through the life cycle of nano-enhanced products. *Environ Int*. 2015;77:132-47. doi: 10.1016/j.ENVINT.2015.01.013, PMID 25705000.
 88. Soni D, Naoghare PK, Saravanadevi S, et al. Release, transport, and toxicity of engineered nanoparticles. *Rev Environ Contam Toxicol*. 2015;234:1-47. doi: 10.1007/978-3-319-10638-0_1.
 89. Baalousha M, Lead JR. Overview of nanoscience in the environment. *Environ Hum Health Impacts Nanotechnol*. 2009;1-29. doi: 10.1002/9781444307504.CHI.
 90. Tiwari AJ, Marr LC. The role of atmospheric transformations in determining environmental impacts of carbonaceous nanoparticles. *J Environ Qual*. 2010;39(6):1883-95. doi: 10.2134/JEQ2010.0050, PMID 21284286.
 91. Gouin T, Roche N, Lohmann R, Hodges G. A thermodynamic approach for assessing the environmental exposure of chemicals absorbed to microplastic. *Environ Sci Technol*. 2011;45(4):1466-72. doi: 10.1021/es1032025, PMID 21268630.
 92. Gidhagen L, Johansson C, Omstedt G, Langner J, Olivares G. Model simulations of NOx and ultrafine particles close to a Swedish highway. *Environ Sci Technol*. 2004;38(24):6730-40. doi: 10.1021/ES0498134, PMID 15669334.
 93. Clarke AG, Robertson LA, Hamilton RS, Gorbunov B. A Lagrangian model of the evolution of the particulate size distribution of vehicular emissions. *Sci Total Environ*. 2004;334-335:197-206. doi: 10.1016/j.SCITOTENV.2004.04.038, PMID 15504506.
 94. Nogueira V, Lopes I, Rocha-Santos T, Gonçalves F, Pereira R. Toxicity of solid residues resulting from wastewater treatment with nanomaterials. *Aquat Toxicol*. 2015;165:172-8. doi: 10.1016/j.AQUATOX.2015.05.021, PMID 26057932.
 95. Jang MH, Bae SJ, Lee SK, Lee YJ, Hwang YS. Effect of material properties on stability of silver nanoparticles in water. *J Nanosci Nanotechnol*. 2014;14(12):9665-9. doi: 10.1166/JNN.2014.10161, PMID 25971117.
 96. Rocha TL, Gomes T, Sousa VS, Mestre NC, Bebianno MJ. Ecotoxicological impact of engineered nanomaterials in bivalve mollusks: an overview. *Mar Environ Res*. 2015;111:74-88. doi: 10.1016/j.MARENRES.2015.06.013, PMID 26152602.
 97. Grillo R, Rosa AH, Fraceto LF. Engineered nanoparticles and organic matter: a review of the state-of-the-art. *Chemosphere*. 2015;119:608-19. doi: 10.1016/j.CHEMOSPHERE.2014.07.049, PMID 25128893.
 98. Ma S, Lin D. The physicochemical interactions at the interfaces between nanoparticles and aquatic organisms: adsorption and internalization. *Environ Sci Process Impacts*. 2013;15(1):145-60. doi: 10.1039/C2EM30637A, PMID 24592433.
 99. Matranga V, Corsi I. Toxic effects of engineered nanoparticles in the marine environment: model organisms and molecular approaches. *Mar Environ Res*. 2012;76:32-40. doi: 10.1016/j.MARENRES.2012.01.006, PMID 22391237.
 100. Tiede K, Hanssen SF, Westerhoff P, Fern GJ, Hankin SM, Aitken RJ, et al. How important is drinking water exposure for the risks of engineered nanoparticles to consumers? *Nanotoxicology*. 2016;10(1):102-10. doi: 10.3109/17435390.2015.1022888, PMID 25962682.
 101. Mukhopadhyay SS. Nanotechnology in agriculture: prospects and constraints. *Nanotechnol Sci Appl*. 2014;7(2):63-71. doi: 10.2147/NSA.S39409, PMID 25187699.
 102. Viswanath B, Kim S. Influence of nanotoxicity on human health and environment: the alternative strategies. *Rev Environ Contam Toxicol*. 2017;242:61-104. doi: 10.1007/398_2016_12/COVER.
 103. Li XQ, Elliott DW, Zhang WX. Zero-valent iron nanoparticles for abatement of environmental pollutants: materials and engineering aspects. *Critical Reviews in Solid State and Materials Sciences*. 2006;31(4):111-22. doi: 10.1080/10408430601057611.
 104. Boxall ABA, Tiede K, Chaudhry Q. Engineered nanomaterials in soils and water: how do they behave and could they pose a risk to human health? *Nanomedicine (Lond)*. 2007;2(6):919-27. doi: 10.2217/17435889.2.6.919, PMID 18095854.
 105. Zharov VP, Mercer KE, Galitovskaya EN, Smeltzer MS. Photothermal nanotherapeutics and nanodiagnostics for selective killing of bacteria targeted with gold nanoparticles. *Biophys J*. 2006;90(2):619-27. doi: 10.1529/BIOPHYSJ.105.061895, PMID 16239330.
 106. Jafar G, Hamzeh G. Ecotoxicity of nanomaterials in soil. *Ann Biol Res*. 2013.
 107. Kim J, Grate JW, Wang P. Nanostructures for enzyme stabilization. *Chem Eng Sci*. 2006;61(3):1017-26. doi: 10.1016/j.CES.2005.05.067.
 108. Samorì B, Zuccheri G. DNA codes for nanoscience. *Angew Chem Int Ed Engl*. 2005;44(8):1166-81. doi: 10.1002/ANIE.200400652, PMID 15532060.
 109. Sarikaya M, Tamerler C, Jen AKY, Schulten K, Baneyx F. Molecular biomimetics: nanotechnology through biology. *Nat Mater*. 2003;2(9):577-85. doi: 10.1038/NMAT964, PMID 12951599.
 110. Seeman NC. DNA in a material world. *Nature*. 2003;421(6921):427-31. doi: 10.1038/nature01406, PMID 12540916.
 111. Zhao X, Zhang S. Molecular designer self-assembling peptides. *Chem Soc Rev*. 2006;35(11):1105-10. doi: 10.1039/B511336A, PMID 17057839.
 112. Åkerman ME, Chan WCW, Laakkonen P, Bhatia SN, Ruoslahti E. Nanocrystal targeting in vivo. *Proc Natl*

- Acad Sci U S A. 2002;99(20):12617-21. doi: 10.1073/pans.152463399, PMID 12235356.
113. Allen TM, Cullis PR. Drug delivery systems: entering the mainstream. *Science*. 2004;303(5665):1818-22. doi: 10.1126/SCIENCE.1095833, PMID 15031496.
 114. Arap W, Pasqualini R, Ruoslahti E. Cancer treatment by targeted drug delivery to tumor vasculature in a mouse model. *Science*. 1998;279(5349):377-80. doi: 10.1126/SCIENCE.279.5349.377, PMID 9430587.
 115. Gref R, Minamitake Y, Peracchia MT, Trubetskoy V, Torchilin V, Langer R. Biodegradable long-circulating polymeric nanospheres. *Science*. 1994;263(5153):1600-3. doi: 10.1126/SCIENCE.8128245, PMID 8128245.
 116. Martin CR, Kohli P. The emerging field of nanotube biotechnology. *Nat Rev Drug Discov*. 2003;2(1):29-37. doi: 10.1038/NRD988, PMID 12509757.
 117. Sui J, Tleugabulova D, Brennan JD. Direct and indirect monitoring of peptide-silica interactions using time-resolved fluorescence anisotropy. *Langmuir*. 2005;21(11):4996-5001. doi: 10.1021/LA0473963, PMID 15896042.
 118. Taylor JR, Fang MM, Nie S. Probing specific sequences on single DNA molecules with bioconjugated fluorescent nanoparticles. *Anal Chem*. 2000;72(9):1979-86. doi: 10.1021/AC991331I, PMID 10815954.
 119. Klein J. Probing the interactions of proteins and nanoparticles. *Proc Natl Acad Sci U S A*. 2007;104(7):2029-30. doi: 10.1073/pans.0611610104, PMID 17284585.
 120. Lerer D. Big things in small packages: evaluating the City of Berkeley's nanotechnology ordinance effectiveness as a model of targeted transparency ARTICLE big things in small packages: evaluating the City of Berkeley's nanotechnology ordinance effectiveness as a model of targeted transparency; 2013.
 121. Colvin VL. The potential environmental impact of engineered nanomaterials. *Nat Biotechnol*. 2003;21(10):1166-70. doi: 10.1038/nbt875, PMID 14520401.
 122. Balbus JM, Maynard AD, Colvin VL, Castranova V, Daston GP, Denison RA, et al. Meeting report: hazard assessment for nanoparticles—report from an interdisciplinary workshop. *Environ Health Perspect*. 2007;115(11):1654-9. doi: 10.1289/ehp.10327, PMID 18007999.
 123. Nel A, Xia T, Mädler L, Li N. Toxic potential of materials at the nanolevel. *Science*. 2006;311(5761):622-7. doi: 10.1126/SCIENCE.1114397, PMID 16456071.
 124. Oberdörster G, Oberdörster E, Oberdörster J. Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect*. 2005;113(7):823-39. doi: 10.1289/EHP.7339, PMID 16002369.
 125. Derfus AM, Chan WCW, Bhatia SN. Probing the cytotoxicity of semiconductor quantum dots. *Nano Lett*. 2004;4(1):11-8. doi: 10.1021/nl0347334, PMID 28890669.
 126. Gurr JR, Wang ASS, Chen CH, Jan KY. Ultrafine titanium dioxide particles without photoactivation can induce oxidative damage to human bronchial epithelial cells. *Toxicology*. 2005;213(1-2):66-73. doi: 10.1016/j.tox.2005.05.007, PMID 15970370.
 127. Ramires PA, Cosentino F, Milella E, Torricelli P, Giavaresi G, Giardino R. In vitro response of primary rat osteoblasts to titania/hydroxyapatite coatings compared with transformed human osteoblast-like cells. *J Mater Sci Mater Med*. 2002;13(8):797-801. doi: 10.1023/A:1016183326864, PMID 15348568.
 128. Soto KF, Carrasco A, Powell TG, et al. Comparative in vitro cytotoxicity assessment of some manufactured nanoparticulate materials characterized by transmission electron microscopy. *J Nanopart Res*. 2005;7(2-3):145-69. doi: 10.1007/S11051-005-3473-1/METRICS.
 129. Suh WH, Jang AR, Suh Y-H, Suslick K. Porous, hollow, and ball-in-ball metal oxide microspheres: preparation, endocytosis, and cytotoxicity. *Adv Mater*. 2006;18(14):1832-7. doi: 10.1002/ADMA.200600222.
 130. Yoshida K, Morita M, Mishina H. Cytotoxicity of metal and ceramic particles in different sizes. *JSME Int J Ser C*. 2003;46(4):1284-9. doi: 10.1299/JSMEC.46.1284.
 131. Borm PJA, Kreyling W. Toxicological hazards of inhaled nanoparticles—potential implications for drug delivery. *J Nanosci Nanotechnol*. 2004;4(5):521-31. doi: 10.1166/JNN.2004.081, PMID 15503438.
 132. Dobrovolskaia MA, McNeil SE. Immunological properties of engineered nanomaterials. *Nat Nanotechnol*. 2007;2(8):469-78. doi: 10.1038/nano.2007.223, PMID 18654343.
 133. Garnett MC, Kallinteri P. Nanomedicines and nanotoxicology: some physiological principles. *Occup Med (Lond)*. 2006;56(5):307-11. doi: 10.1093/OCCMED/KQL052, PMID 16868128.
 134. Handy RD, Shaw BJ. Toxic effects of nanoparticles and nanomaterials: implications for public health, risk assessment and the public perception of nanotechnology. *Health Risk & Society*. 2007;9(2):125-44. doi: 10.1080/13698570701306807.
 135. Hardman R. A toxicologic review of quantum dots: toxicity depends on physicochemical and environmental factors. *Environ Health Perspect*. 2006;114(2):165-72. doi: 10.1289/EHP.8284, PMID 16451849.
 136. Elder A, Gelein R, Silva V, Feikert T, Opanashuk L, Carter J, et al. Translocation of inhaled ultrafine manganese oxide particles to the central nervous system. *Environ Health Perspect*. 2006;114(8):1172-8. doi: 10.1289/EHP.9030, PMID 16882521.
 137. Medina C, Santos-Martinez MJ, Radomski A, Corrigan OI, Radomski MW. Nanoparticles: pharmacological and toxicological significance. *Br J Pharmacol*. 2007;150(5):552-8. doi: 10.1038/SJ.BJP.0707130, PMID 17245366.
 138. Oberdörster G, Oberdörster E, Oberdörster J. Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect*. 2005;113(7):823-39. doi: 10.1289/EHP.7339, PMID 16002369.
 139. Cushing BL, Kolesnichenko VL, O'Connor CJ. Recent advances in the liquid-phase syntheses of inorganic nanoparticles. *Chem Rev*. 2004;104(9):3893-946. doi: 10.1021/cr030027b, PMID 15352782.
 140. Dai H. Carbon nanotubes: synthesis, integration, and properties. *Acc Chem Res*. 2002;35(12):1035-44. doi: 10.1021/AR0101640, PMID 12484791.
 141. Huber DL. Synthesis, properties, and applications of iron nanoparticles. *Small*. 2005;1(5):482-501. doi: 10.1002/SMLL.200500006, PMID 17193474.
 142. Jeong U, Teng X, Wang Y, Yang H, Xia Y. Superparamagnetic colloids: controlled synthesis and

- niche applications. *Adv Mater.* 2007;19(1):33-60. doi: 10.1002/ADMA.200600674.
143. Lee J, Kim J, Hyeon T. Recent progress in synthesizing porous carbon materials. *Adv Mater.* 2006;18(16):2073-94. doi: 10.1002/ADMA.200501576.
 144. Lu AH, Salabas EL, Schüth F. Magnetic nanoparticles: synthesis, protection, functionalization, and application. *Angew Chem Int Ed Engl.* 2007;46(8):1222-44. doi: 10.1002/ANIE.200602866, PMID 17278160.
 145. Medintz IL, Uyeda HT, Goldman ER, Mattoussi H. Quantum dot bioconjugates for imaging, labeling, and sensing. *Nat Mater.* 2005;4(6):435-46. doi: 10.1038/NMAT1390, PMID 15928695.
 146. Michalet X, Pinaud FF, Bentolila LA, Tsay JM, Doose S, Li JJ, et al. Quantum dots for live cells, in vivo imaging, and diagnostics. *Science.* 2005;307(5709):538-44. doi: 10.1126/SCIENCE.1104274, PMID 15681376.
 147. Chan VSW. Nanomedicine: an unresolved regulatory issue. *Regul Toxicol Pharmacol.* 2006;46(3):218-24. doi: 10.1016/j.YRTPH.2006.04.009, PMID 17081666.
 148. del Pino Pd, Pelaz B, Zhang Q, Maffre P, Nienhaus GU, Parak WJ. Protein corona formation around nanoparticles – from the past to the future. *Mater Horiz.* 2014;1(3):301-13. doi: 10.1039/C3MH00106G.
 149. Pearson RM, Juettner VV, Hong S. Biomolecular corona on nanoparticles: a survey of recent literature and its implications in targeted drug delivery. *Front Chem.* 2014;2:108. doi: 10.3389/fchem.2014.00108. PMID 25506050.
 150. Kim BYS, Rutka JT, Chan WCW. Nanomedicine. *N Engl J Med.* 2010;363(25):2434-43. doi: 10.1056/NEJMr0912273, PMID 21158659.
 151. Nel AE, Mädler L, Velegol D, Xia T, Hoek EM, Somasundaran P, et al. It is understanding physicochemical interactions at the nano–bio interface. *Nat Mater.* 2009;8(7):543-57. doi: 10.1038/nmat2442, PMID 19525947.
 152. Germain M, Caputo F, Metcalfe S, Tosi G, Spring K, Åslund AKO, et al. Delivering the power of nanomedicine to patients today. *J Control Release.* 2020;326:164-71. doi: 10.1016/j.JCONREL.2020.07.007, PMID 32681950.
 153. Anonymous, ETPN. Nanomedicine European Technology Platform; n.d. [cited 9/8/2022] Available from: <https://etp-nanomedicine.eu/>.
 154. Suh NP, Suh PN. The principles of design. *Oxf Univ Press Demand.* 1990.
 155. Ray MA, Trammell RA, Verhulst S, Ran S, Toth LA. Development of a mouse model for assessing fatigue during chemotherapy. *Comp Med.* 2011;61(2):119-30. PMID 21535922.
 156. Babaie S, del Bakhshayesh ARD, Ha JW, Hamishehkar H, Kim KH. Invasome: A novel nanocarrier for transdermal drug delivery. *Nanomaterials (Basel).* 2020;10(2). doi: 10.3390/NANO10020341, PMID 32079276.
 157. Zhao B, He YY, Chignell CF, Yin JJ, Andley U, Roberts JE. The difference in phototoxicity of cyclodextrin complexed fullerene [γ -CyD]/C60 and its aggregated derivatives toward human lens epithelial cells. *Chem Res Toxicol.* 2009;22(4):660-7. doi: 10.1021/TX800478U, PMID 19281132.
 158. Arts JHE, Hadi M, Keene AM, Kreiling R, Lyon D, Maier M, et al. A critical appraisal of existing concepts for the grouping of nanomaterials. *Regul Toxicol Pharmacol.* 2014;70(2):492-506. doi: 10.1016/j.YRTPH.2014.07.025, PMID 25108058.
 159. Landsiedel R, Ma-Hock L, Wiench K, Wohlleben W, Sauer UG. Safety assessment of nanomaterials using an advanced decision-making framework, the DF4nanoGrouping. *J Nanopart Res.* 2017;19(5):171. doi: 10.1007/s11051-017-3850-6. PMID 28553159.
 160. Gajewicz A, Puzyn T, Odziomek K, Urbaszek P, Haase A, Riebeling C, et al. Decision tree models to classify nanomaterials according to the DF4nanoGrouping scheme. *Nanotoxicology.* 2018;12(1):1-17. doi: 10.1080/17435390.2017.1415388, PMID 29251527.
 161. Müller RH, Gohla S, Keck CM. State of the art of nanocrystals--special features, production, nanotoxicology aspects, and intracellular delivery. *Eur J Pharm Biopharm.* 2011;78(1):1-9. doi: 10.1016/j.EJPB.2011.01.007, PMID 21266197.
 162. Rycroft T, Trump B, Poinatte-Jones K, Linkov I. Nanotoxicology and nanomedicine: making development decisions in an evolving governance environment. *J Nanopart Res.* 2018;20(2). doi: 10.1007/S11051-018-4160-3.
 163. Magaye RR, Yue X, Zou B, Shi H, Yu H, Liu K, et al. Acute toxicity of nickel nanoparticles in rats after intravenous injection. *Int J Nanomedicine.* 2014;9(1):1393-402. doi: 10.2147/IJN.S56212, PMID 24648736.
 164. Arefian Z, Pishbin F, Negahdary M, et al. Potential toxic effects of zirconia oxide nanoparticles on liver and kidney factors; 2015.
 165. Bellusci M, la Barbera A, Padella F, Mancuso M, Pasquo A, Grollino MG, et al. Biodistribution and acute toxicity of a nanofluid containing manganese iron oxide nanoparticles produced by a mechanochemical process. *Int J Nanomedicine.* 2014;9(1):1919-29. doi: 10.2147/IJN.S56394, PMID 24790434.
 166. Babadi VY, Najafi L, Najafi A, et al. Evaluation of Iron oxide nanoparticles effects on tissue and thyroid enzymes in Rats; 2013.
 167. Xu J, Shi H, Ruth M, Yu H, Lazar L, Zou B, et al. Acute toxicity of intravenously administered titanium dioxide nanoparticles in mice. *PLOS ONE.* 2013;8(8):e70618. doi: 10.1371/JOURNAL.PONE.0070618, PMID 23950972.
 168. Awaad A. Histopathological and immunological changes induced by magnetite nanoparticles in the spleen, liver, and genital tract of mice following intravaginal instillation. *J Basic Appl Zool.* 2015;71:32-47. doi: 10.1016/j.jobaz.2015.03.003.
 169. Cai X, Lee A, Ji Z, Huang C, Chang CH, Wang X, et al. Reduction of pulmonary toxicity of metal oxide nanoparticles by phosphonate-based surface passivation. *Part Fibre Toxicol.* 2017;14(1). doi: 10.1186/S12989-017-0193-5.
 170. Sadeghi L, Yousefi Babadi V, Espanani HR. Toxic effects of the Fe2O3 nanoparticles on the liver and lung tissue. *Bratisl Lek Listy.* 2015;116(6):373-8. doi: 10.4149/BLL_2015_071, PMID 26084739.
 171. S S, K V, S P, N R, K K. In vitro cytotoxicity of zinc oxide, iron oxide, and copper nanopowders prepared by green synthesis. *Toxicol Rep.* 2017;4:427-30. doi: 10.1016/j.TOXREP.2017.07.005, PMID 28959669.
 172. Fartkhooni FM, Noori A, Mohammadi A. Effects of titanium dioxide nanoparticles toxicity on the kidney of male rats. *Int J Life Sci.* 2016;10(1):65-9. doi: 10.3126/IJLS.V10I1.14513.

173. Raju HB, Hu Y, Vedula A, Dubovy SR, Goldberg JL. Evaluation of magnetic micro- and nanoparticle toxicity to ocular tissues. *PLOS ONE*. 2011;6(5):e17452. doi: 10.1371/JOURNAL.PONE.0017452, PMID 21637340.
174. Kim DK. Nanomedicine for inner ear diseases: a review of recent in vivo studies. *BioMed Res Int*. 2017;2017:3098230. doi: 10.1155/2017/3098230, PMID 29130038.
175. Ibrahim A. Ibrahim, M.d. AFAAMS; Manal M. Morsy, M.d. HELMMD; Effect of Zinc Oxide Nanoparticles on the Structure of Testis of Adult Albino Rats and the Possible Protective Role of Naringenin. *The Medical Journal of Cairo University*. 2019;87(September):3469-83. doi: 10.21608/mjcu.2019.65644.
176. Kong L, Tang M, Zhang T, Wang D, Hu K, Lu W, et al. Nickel nanoparticles exposure and reproductive toxicity in healthy adult rats. *Int J Mol Sci*. 2014;15(11):21253-69. doi: 10.3390/IJMS151121253, PMID 25407529.
177. Gonzalez L, Lison D, Kirsch-Volders M. Genotoxicity of engineered nanomaterials: A critical review. *Nanotoxicology*. 2008;2(4):252-73. doi: 10.1080/17435390802464986.
178. Yin JJ, Liu J, Ehrenshaft M, Roberts JE, Fu PP, Mason RP, et al. Phototoxicity of Nano titanium dioxides in HaCaT keratinocytes--generation of reactive oxygen species and cell damage. *Toxicol Appl Pharmacol*. 2012;263(1):81-8. doi: 10.1016/J.TAAP.2012.06.001, PMID 22705594.
179. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol*. 2007;39(1):44-84. doi: 10.1016/J.BIOCEL.2006.07.001, PMID 16978905.
180. Meng H, Xia T, George S, Nel AE. A predictive toxicological paradigm for the safety assessment of nanomaterials. *ACS Nano*. 2009;3(7):1620-7. doi: 10.1021/NN9005973, PMID 21452863.
181. Li Y, Yu S, Wu Q, Tang M, Pu Y, Wang D. Chronic Al₂O₃-nanoparticle exposure causes neurotoxic effects on locomotion behaviors by inducing severe ROS production and disrupting ROS defense mechanisms in nematode *Caenorhabditis elegans*. *J Hazard Mater*. 2012;219-220:221-30. doi: 10.1016/J.HAZMAT.2012.03.083, PMID 22521136.
182. Kim S, Ryu DY. Silver nanoparticle-induced oxidative stress, genotoxicity and apoptosis in cultured cells and animal tissues. *J Appl Toxicol*. 2013;33(2):78-89. doi: 10.1002/JAT.2792, PMID 22936301.
183. de Moura MB, dos Santos LS, van Houten B. Mitochondrial dysfunction in neurodegenerative diseases and cancer. *Environ Mol Mutagen*. 2010;51(5):391-405. doi: 10.1002/EM.20575, PMID 20544881.
184. Hsin YH, Chen CF, Huang S, Shih TS, Lai PS, Chueh PJ. ROS mediates the apoptotic effect of nanosilver- and JNK-dependent mechanisms involving the mitochondrial pathway in NIH3T3 cells. *Toxicol Lett*. 2008;179(3):130-9. doi: 10.1016/J.TOXLET.2008.04.015, PMID 18547751.
185. Akhtar MJ, Ahamed M, Kumar S, Siddiqui H, Patil G, Ashquin M, et al. Nanotoxicity of pure silica mediated through oxidant generation rather than glutathione depletion in human lung epithelial cells. *Toxicology*. 2010;276(2):95-102. doi: 10.1016/J.TOX.2010.07.010, PMID 20654680.
186. Akhtar MJ, Ahamed M, Fareed M, Alrokayan SA, Kumar S. Protective effect of sulforaphane against oxidative stress-mediated toxicity induced by CuO nanoparticles in mouse embryonic fibroblasts BALB 3T3. *J Toxicol Sci*. 2012;37(1):139-48. doi: 10.2131/JTS.37.139, PMID 22293418.
187. Fu PP, Xia Q, Hwang HM, Ray PC, Yu H. Mechanisms of nanotoxicity: generation of reactive oxygen species. *J Food Drug Anal*. 2014;22(1):64-75. doi: 10.1016/J.JFDA.2014.01.005, PMID 24673904.
188. Shvedova AA, Castranova V, Kisin ER, Schwegler-Berry D, Murray AR, Gandelsman VZ, et al. Exposure to carbon nanotube material: assessment of nanotube cytotoxicity using human keratinocyte cells. *J Toxicol Environ Health A*. 2003;66(20):1909-26. doi: 10.1080/713853956, PMID 14514433.
189. Winnik FM, Maysinger D. Quantum dot cytotoxicity and ways to reduce it. *Acc Chem Res*. 2013;46(3):672-80. doi: 10.1021/AR3000585, PMID 22775328.
190. Fan Z, Lu JG. Zinc oxide nanostructures: synthesis and properties. *J Nanosci Nanotechnol*. 2005;5(10):1561-73. doi: 10.1166/JNN.2005.182, PMID 16245516.
191. Liu Y, Li X, Bao S, Lu Z, Li Q, Li CM. Plastic protein microarray to investigate the molecular pathways of magnetic nanoparticle-induced nanotoxicity. *Nanotechnology*. 2013;24(17):175501. doi: 10.1088/0957-4484/24/17/175501, PMID 23558511.
192. Wang Y, Aker WG, Hwang HM, Yedjou CG, Yu H, Tchounwou PB. A study of the mechanism of in vitro cytotoxicity of metal oxide nanoparticles using primary catfish hepatocytes and human HepG2 cells. *Sci Total Environ*. 2011;409(22):4753-62. doi: 10.1016/J.SCITOTENV.2011.07.039, PMID 21851965.
193. Nel A, Xia T, Mädler L, Li N. Toxic potential of materials at the nanolevel. *Science*. 2006;311(5761):622-7. doi: 10.1126/SCIENCE.1114397, PMID 16456071.
194. Yoshida T, Yoshikawa T, Nabeshi H, Tsutsumi Y. [Relation analysis between the intracellular distribution of nanomaterials, ROS generation, and DNA damage]. *Yakugaku Zasshi*. 2012;132(3):295-300. doi: 10.1248/YAKUSHI.132.295, PMID 22382833.
195. Boyoglu C, He Q, Willing G, Boyoglu-Barnum S, Dennis VA, Pillai S, et al. Microscopic studies of various sizes of gold nanoparticles and their cellular localization. *ISRN Nanotechnol*. 2013;2013:1-13. doi: 10.1155/2013/123838.
196. Persson H, Købler C, Mølhav K, Samuelson L, Tegenfeldt JO, Oredsson S, et al. Fibroblasts cultured on nanowires exhibit low motility, impaired cell division, and DNA damage. *Small*. 2013;9(23):4006-16. doi: 10.1002/SMLL.201300644, PMID 23813871.
197. Wang S, Lu W, Tovmachenko O, Rai US, Yu H, Ray PC. The challenge in understanding size and shape-dependent toxicity of gold nanomaterials in human skin keratinocytes. *Chem Phys Lett*. 2008;463(1-3):145-9. doi: 10.1016/J.CPLETT.2008.08.039, PMID 24068836.
198. Ispas C, Andreescu D, Patel A, Goia DV, Andreescu S, Wallace KN. Toxicity and developmental defects of different sizes and shapes nickel nanoparticles in zebrafish. *Environ Sci Technol*. 2009;43(16):6349-56. doi: 10.1021/ES9010543, PMID 19746736.
199. Oh WK, Kim S, Choi M, Kim C, Jeong YS, Cho BR, et al. Cellular uptake, cytotoxicity, and innate immune response of silica-titania hollow nanoparticles based on size and surface functionality. *ACS Nano*.

- 2010;4(9):5301-13. doi: 10.1021/NN100561E, PMID 20698555.
200. Studer AM, Limbach LK, van Duc L, Krumeich F, Athanassiou EK, Gerber LC, et al. Nanoparticle cytotoxicity depends on intracellular solubility: comparison of stabilized copper metal and degradable copper oxide nanoparticles. *Toxicol Lett.* 2010;197(3):169-74. doi: 10.1016/j.toxlet.2010.05.012, PMID 20621582.
201. Shen C, James SA, de Jonge MD, Turney TW, Wright PF, Feltis BN. Relating cytotoxicity, zinc ions, and reactive oxygen in ZnO nanoparticle-exposed human immune cells. *Toxicol Sci.* 2013;136(1):120-30. doi: 10.1093/toxsci/kft187, PMID 23997113.
202. Jastrzębska E, Bazylińska U, Bułka M, Tokarska K, Chudy M, Dybko A, et al. Microfluidic platform for photodynamic therapy cytotoxicity analysis of nano encapsulated indocyanine-type photosensitizers. *Biomicrofluidics.* 2016;10(1):014116. doi: 10.1063/1.4941681, PMID 26909122.
203. Daimon T, Nosaka Y. Formation and Behavior of Singlet Molecular Oxygen in TiO₂ Photocatalysis Studied by Detection of Near-Infrared Phosphorescence. *J Phys Chem C.* 2007;111(11):4420-4. doi: 10.1021/jp070028Y.
204. Liao KH, Lin YS, MacOsco CW, Haynes CL. Cytotoxicity of graphene oxide and graphene in human erythrocytes and skin fibroblasts. *ACS Appl Mater Interfaces.* 2011;3(7):2607-15. doi: 10.1021/AM200428V, PMID 21650218.
205. Ray PC, Yu H, Fu PP. Toxicity and environmental risks of nanomaterials: challenges and future needs. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev.* 2009;27(1):1-35. doi: 10.1080/10590500802708267, PMID 19204862.
206. Karlsson HL, Cronholm P, Gustafsson J, Möller L. Copper oxide nanoparticles are highly toxic: a comparison between metal and carbon nanotubes. *Chem Res Toxicol.* 2008;21(9):1726-32. doi: 10.1021/TX800064J, PMID 18710264.
207. Zhu X, Hondroulis E, Liu W, Li CZ. Biosensing approaches for rapid genotoxicity and cytotoxicity assays upon nanomaterial exposure. *Small.* 2013;9(9-10):1821-30. doi: 10.1002/SMLL.201201593, PMID 23417999.
208. Bai W, Zhang Z, Tian W, He X, Ma Y, Zhao Y, et al. Toxicity of zinc oxide nanoparticles to zebrafish embryo: A physicochemical study of toxicity mechanism. *J Nanopart Res.* 2010;12(5):1645-54. doi: 10.1007/S11051-009-9740-9.
209. Stankic S, Suman S, Haque F, Vidic J. Pure and multi metal oxide nanoparticles: synthesis, antibacterial and cytotoxic properties. *J Nanobiotechnology.* 2016;14(1):73. doi: 10.1186/S12951-016-0225-6, PMID 27776555.
210. Papadiamantis AG, Jänes J, Voyiatzis E, Sikk L, Burk J, Burk P, et al. Predicting cytotoxicity of metal oxide nanoparticles using the Isalos analytics platform. *Nanomaterials (Basel).* 2020;10(10):1-19. doi: 10.3390/NANO10102017, PMID 33066094.
211. Kawasaki RK, Romano M, Dietrich N, Araki K. Titanium and iron oxide nanoparticles for cancer therapy: surface chemistry and biological implications. *Front Nanotechnol.* 2021;3:68. doi: 10.3389/fnano.2021.735434.
212. He F, Ru X, Wen T. NRF2, a transcription factor for stress response and beyond. *Int J Mol Sci.* 2020;21(13):4777. doi: 10.3390/IJMS21134777, PMID 32640524.
213. Jafari S, Mahyad B, Hashemzadeh H, Janfaza S, Gholikhani T, Tayebi L. Biomedical applications of TiO₂ nanostructures: recent advances. *Int J Nanomedicine.* 2020;15:3447-70. doi: 10.2147/IJN.S249441, PMID 32523343.
214. Brun NR, Lenz M, Wehrli B, Fent K. Comparative effects of zinc oxide nanoparticles and dissolved zinc on zebrafish embryos and eleuthero-embryos: the importance of zinc ions. *Sci Total Environ.* 2014;476-477:657-66. doi: 10.1016/j.scitotenv.2014.01.053, PMID 24508854.
215. D'Amora M, Schmidt TJN, Konstantinidou S, Raffa V, De Angelis F, Tantussi F. Effects of metal oxide nanoparticles in zebrafish. *Oxid Med Cell Longev.* 2022;2022:3313016. doi: 10.1155/2022/3313016, PMID 35154565.
216. Zhang Y, Zhu L, Zhou Y, Chen J. Accumulation and elimination of iron oxide nanomaterials in zebrafish (*Danio rerio*) upon chronic aqueous exposure. *J Environ Sci (China).* 2015;30:223-30. doi: 10.1016/j.jes.2014.08.024, PMID 25872731.
217. Laurent S, Forge D, Port M, Roch A, Robic C, Vander Elst L, et al. Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. *Chem Rev.* 2008;108(6):2064-110. doi: 10.1021/cr068445e, PMID 18543879.
218. Al-Thani HF, Shurbaji S, Yalcin HC. Zebrafish as a model for anticancer nanomedicine studies. *Pharmaceuticals (Basel).* 2021;14(7). doi: 10.3390/PH14070625, PMID 34203407.
219. Mohiuddin Hafiz S, Sameer Kulkarni S, Kapil Thakur M. In-vivo toxicity assessment of biologically synthesized iron oxide nanoparticles in zebrafish (*Danio rerio*). *Biosci Biotech Res Asia.* 2018;15(2):419-25. doi: 10.13005/BBRA/2645.
220. Malhotra N, Chen JR, Sarasamma S, Audira G, Siregar P, Liang ST, et al. Ecotoxicity assessment of Fe₃O₄ magnetic nanoparticle exposure in adult zebrafish at a pertinent environmental concentration by behavioral and biochemical testing. *Nanomaterials (Basel).* 2019;9(6). doi: 10.3390/NANO9060873, PMID 31181856.
221. Zheng M, Lu J, Zhao D. Effects of starch-coating of magnetite nanoparticles on cellular uptake, toxicity and gene expression profiles in adult zebrafish. *Sci Total Environ.* 2018;622-623:930-41. doi: 10.1016/j.scitotenv.2017.12.018, PMID 29227944.
222. de Oliveira GMT, Kist LW, Pereira TCB, Bortolotto JW, Paquete FL, de Oliveira EM, et al. Transient modulation of acetylcholinesterase activity caused by exposure to dextran-coated iron oxide nanoparticles in the brain of adult zebrafish. *Comp Biochem Physiol C Toxicol Pharmacol.* 2014;162(1):77-84. doi: 10.1016/j.cbpc.2014.03.010, PMID 24704546.
223. Zhu X, Zhu L, Duan Z, Qi R, Li Y, Lang Y. Comparative toxicity of several metal oxide nanoparticle aqueous suspensions to zebrafish (*Danio rerio*) early developmental stage. *J Environ Sci Health A Tox Hazard Subst Environ Eng.* 2008;43(3):278-84. doi: 10.1080/10934520701792779, PMID 18205059.

224. Zhu X, Wang J, Zhang X, Chang Y, Chen Y. The impact of ZnO nanoparticle aggregates on the embryonic development of zebrafish (*Danio rerio*). *Nanotechnology*. 2009;20(19):195103. doi: 10.1088/0957-4484/20/19/195103, PMID 19420631.
225. Zhao X, Ren X, Zhu R, Luo Z, Ren B. Zinc oxide nanoparticles induce oxidative DNA damage and ROS-triggered mitochondria-mediated apoptosis in zebrafish embryos. *Aquat Toxicol*. 2016;180:56-70. doi: 10.1016/j.aquatox.2016.09.013, PMID 27658222.
226. Chen TH, Lin CC, Meng PJ. Zinc oxide nanoparticles alter zebrafish's hatching and larval locomotor activity (*Danio rerio*). *J Hazard Mater*. 2014;277:134-40. doi: 10.1016/j.hazmat.2013.12.030, PMID 24424259.
227. Hua J, Vijver MG, Richardson MK, Ahmad F, Peijnenburg WJ. Particle-specific toxic effects of differently shaped zinc oxide nanoparticles to zebrafish embryos (*Danio rerio*). *Environ Toxicol Chem*. 2014;33(12):2859-68. doi: 10.1002/etc.2758, PMID 25244315.
228. Zhou Z, Son J, Harper B, Zhou Z, Harper S. Influence of surface chemical properties on the toxicity of engineered zinc oxide nanoparticles to embryonic zebrafish. *Beilstein J Nanotechnol*. 2015;6(1):1568-79. doi: 10.3762/bjnano.6.160, PMID 26425408.
229. Rahman HS, Othman HH, Abdullah R, Edin HYAS, Al-Haj NA. Beneficial and toxicological aspects of zinc oxide nanoparticles in animals. *Vet Med Sci*. 2022;8(4):1769-79. doi: 10.1002/vms3.814, PMID 35588498.
230. Sadauskas E, Wallin H, Stoltenberg M, Vogel U, Doering P, Larsen A et al. Kupffer cells are central in the removal of nanoparticles from the organism. *Part Fibre Toxicol*. 2007;4:10. doi: 10.1186/1743-8977-4-10. PMID 17949501.
231. de Jong WH, Hagens WI, Krystek P, Burger MC, Sips AJ, Geertsma RE. Particle size-dependent organ distribution of gold nanoparticles after intravenous administration. *Biomaterials*. 2008;29(12):1912-9. doi: 10.1016/j.biomaterials.2007.12.037, PMID 18242692.
232. Bai Y, Zhang Y, Zhang J, Mu Q, Zhang W, Butch ER, et al. Repeated administrations of carbon nanotubes in male mice cause reversible testis damage without affecting fertility. *Nat Nanotechnol*. 2010;5(9):683-9. doi: 10.1038/nano.2010.153, PMID 20693989.
233. Liu Z, Davis C, Cai W, He L, Chen X, Dai H. Circulation and long-term fate of functionalized, biocompatible single-walled carbon nanotubes in mice probed by Raman spectroscopy. *Proc Natl Acad Sci U S A*. 2008;105(5):1410-5. doi: 10.1073/pnas.0707654105, PMID 18230737.
234. Kim JS, Yoon TJ, Yu KN, Kim BG, Park SJ, Kim HW, et al. Toxicity and tissue distribution of magnetic nanoparticles in mice. *Toxicol Sci*. 2006;89(1):338-47. doi: 10.1093/toxsci/kfj027, PMID 16237191.
235. JANI P, HALBERT GW, LANGRIDGE J, Florence AT. Nanoparticle uptake by the rat gastrointestinal mucosa: quantitation and particle size dependency. *J Pharm Pharmacol*. 1990;42(12):821-6. doi: 10.1111/j.2042-7158.1990.tb07033.x, PMID 1983142.
236. Wang B, He X, Zhang Z, Zhao Y, Feng W. Metabolism of nanomaterials in vivo: blood circulation and organ clearance. *Acc Chem Res*. 2013;46(3):761-9. doi: 10.1021/ar2003336, PMID 23964655.
237. Wu J, Wang C, Sun J, Xue Y. Neurotoxicity of silica nanoparticles: brain localization and dopaminergic neurons damage pathways. *ACS Nano*. 2011;5(6):4476-89. doi: 10.1021/nn103530b, PMID 21526751.
238. Thomas DG, Smith JN, Thrall BD, Baer DR, Jolley H, Munusamy P, et al. ISD3: a pharmacokinetic model for predicting the combined effects of particle sedimentation, diffusion, and dissolution on cellular dosimetry for in vitro systems. *Part Fibre Toxicol*. 2018;15(1):6. doi: 10.1186/s12989-018-0243-7, PMID 29368623.
239. Arora S, Rajwade JM, Paknikar KM. Nanotoxicology and in vitro studies: the need of the hour. *Toxicol Appl Pharmacol*. 2012;258(2):151-65. doi: 10.1016/j.taap.2011.11.010, PMID 22178382.
240. Shukla RK, Sharma V, Pandey AK, Singh S, Sultana S, Dhawan A. ROS-mediated genotoxicity induced by titanium dioxide nanoparticles in human epidermal cells. *Toxicol In Vitro*. 2011;25(1):231-41. doi: 10.1016/j.tiv.2010.11.008, PMID 21092754.
241. Jain J, Arora S, Rajwade JM, Omray P, Khandelwal S, Paknikar KM. Silver nanoparticles in therapeutics: developing an antimicrobial gel formulation for topical use. *Mol Pharm*. 2009;6(5):1388-401. doi: 10.1021/mp900056g, PMID 19473014.
242. Foley S, Crowley C, Smaih M, Bonfils C, Erlanger BF, Seta P, et al. Cellular localization of a water-soluble fullerene derivative. *Biochem Biophys Res Commun*. 2002;294(1):116-9. doi: 10.1016/S0006-291X(02)00445-X, PMID 12054749.
243. Fiorito S, Serafino A, Andreola F, Bernier P. Effects of fullerenes and single-wall carbon nanotubes on murine and human macrophages. *Carbon*. 2006;44(6):1100-5. doi: 10.1016/j.carbon.2005.11.009.
244. Yu T, Malugin A, Ghandehari H. Impact of silica nanoparticle design on cellular toxicity and hemolytic activity. *ACS Nano*. 2011;5(7):5717-28. doi: 10.1021/nn2013904, PMID 21630682.
245. Gonzalez L, Sanderson BJS, Kirsch-Volders M. Adaptations of the in vitro MN assay for the genotoxicity assessment of nanomaterials. *Mutagenesis*. 2011;26(1):185-91. doi: 10.1093/mutage/geq088, PMID 21164201.
246. Karlsson HL. The comet assay in nanotoxicology research. *Anal Bioanal Chem*. 2010;398(2):651-66. doi: 10.1007/s00216-010-3977-0, PMID 20640410.
247. Uo M, Tamura K, Sato Y, Yokoyama A, Watari F, Totsuka Y, et al. The cytotoxicity of metal-encapsulating carbon nanocapsules. *Small*. 2005;1(8-9):816-9. doi: 10.1002/sml.200400143, PMID 17193530.
248. Muller J, Huaux F, Moreau N, Misson P, Heilier JF, Delos M, et al. Respiratory toxicity of multi-wall carbon nanotubes. *Toxicol Appl Pharmacol*. 2005;207(3):221-31. doi: 10.1016/j.taap.2005.01.008, PMID 16129115.
249. Flahaut E, Durrieu MC, Remy-Zolghadri M, Bareille R, Baquey C. Investigation of the cytotoxicity of CCVD carbon nanotubes towards human umbilical vein endothelial cells. *Carbon*. 2006;44(6):1093-9. doi: 10.1016/j.carbon.2005.11.007.
250. Seleverstov O, Zabrinyk O, Zscharnack M, Bulavina L, Nowicki M, Heinrich JM, et al. Quantum dots for human mesenchymal stem cell labeling. A size-dependent autophagy activation. *Nano Lett*. 2006;6(12):2826-32. doi: 10.1021/nl061971i, PMID 17163713.

251. Kostarelos K, Lacerda L, Pastorin G, Wu W, Wieckowski S, Luangvilay J, et al. Cellular uptake of functionalized carbon nanotubes is independent of functional group and cell type. *Nat Nanotechnol.* 2007;2(2):108-13. doi: 10.1038/NNANO.2006.209, PMID 18654229.
252. Monteiro-Riviere NA, Inman AO. Challenges for assessing carbon nanomaterial toxicity to the skin. *Carbon.* 2006;44(6):1070-8. doi: 10.1016/j.carbon.2005.11.004.
253. Gopinath P, Gogoi SK, Chattopadhyay A, Ghosh SS. Implications of silver nanoparticle-induced cell apoptosis for in vitro gene therapy. *Nanotechnology.* 2008;19(7):075104. doi: 10.1088/0957-4484/19/7/075104, PMID 21817629.
254. Fernández-Urrusuno R, Fattal E, Féger J, Couvreur P, Thérond P. Evaluation of hepatic antioxidant systems after intravenous administration of polymeric nanoparticles. *Biomaterials.* 1997;18(6):511-7. doi: 10.1016/S0142-9612(96)00178-0, PMID 9111956.
255. Zhang M, Xu C, Jiang L, Qin J. A 3D human lung-on-a-chip model for nanotoxicity testing. *Toxicol Res (Camb).* 2018;7(6):1048-60. doi: 10.1039/C8TX00156A, PMID 30510678.
256. Yin F, Zhu Y, Zhang M, Yu H, Chen W, Qin J. A 3D human placenta-on-a-chip model to probe nanoparticle exposure at the placental barrier. *Toxicol In Vitro.* 2019;54:105-13. doi: 10.1016/j.tiv.2018.08.014, PMID 30248392.
257. Hafner A, Lovrić J, Lakoš GP, Pepić I. Nanotherapeutics in the EU: an overview on the current state and future directions. *Int J Nanomedicine.* 2014;9(1):1005-23. doi: 10.2147/IJN.S55359, PMID 24600222.
258. Ehmman F, Sakai-Kato K, Duncan R, Hernán Pérez de la Ossa D, Pita R, Vidal JM, et al. Next-generation nanomedicines and nano similars: EU regulators' initiatives on developing and evaluating nanomedicines. *Nanomedicine (Lond).* 2013;8(5):849-56. doi: 10.2217/NNM.13.68, PMID 23656268.
259. Tinkle S, Mcneil SE, Mühlebach S, Bawa R, Borchard G, Barenholz YC, et al. Nanomedicines: addressing the scientific and regulatory gap. *Ann N Y Acad Sci.* 2014;1313(1):35-56. doi: 10.1111/NYAS.12403, PMID 24673240.
260. Astier A, Barton Pai A, Bissig M, Crommelin DJA, Flühmann B, Hecq JD, et al. How to select a nanosimilar. *Ann N Y Acad Sci.* 2017;1407(1):50-62. doi: 10.1111/NYAS.13382, PMID 28715605.
261. Mühlebach S, Borchard G, Yildiz S. Regulatory challenges and approaches to characterize nanomedicines and their follow-on similars. *Nanomedicine (Lond).* 2015;10(4):659-74. doi: 10.2217/NNM.14.189, PMID 25723097.
262. Schellekens H, Stegemann S, Weinstein V, de Vlieger JS, Flühmann B, Mühlebach S, et al. How to regulate nonbiological complex drugs (NBCD) and their follow-on versions: points to consider. *AAPS J.* 2014;16(1):15-21. doi: 10.1208/S12248-013-9533-Z, PMID 24065600.
263. CHMP. Reflection paper on the data requirements for intravenous liposomal products developed concerning an innovator liposomal product Final; 2013.
264. Hua S, de Matos MBC, Metselaar JM, Storm G. Current trends and challenges in the clinical translation of nanoparticulate nanomedicines: pathways for translational development and commercialization. *Front Pharmacol.* 2018;9(JUL):790. doi: 10.3389/par.2018.00790, PMID 30065653.
265. Donaldson K, Duffin R, Langrish JP, Miller MR, Mills NL, Poland CA, et al. Nanoparticles and the cardiovascular system: a critical review. *Nanomedicine (Lond).* 2013;8(3):403-23. doi: 10.2217/NNM.13.16, PMID 23477334.
266. Krug HF, Wick P. Nanotoxicology: an interdisciplinary challenge. *Angew Chem Int Ed Engl.* 2011;50(6):1260-78. doi: 10.1002/ANIE.201001037, PMID 21290492.
267. Zhang H, Ji Z, Xia T, Meng H, Low-Kam C, Liu R, et al. Use metal oxide nanoparticle band gap to develop a predictive paradigm for oxidative stress and acute pulmonary inflammation. *ACS Nano.* 2012;6(5):4349-68. doi: 10.1021/NN3010087, PMID 22502734.
268. Kreyling WG, Semmler-Behnke M, Seitz J, Scymczak W, Wenk A, Mayer P, et al. Size dependence of the translocation of inhaled iridium and carbon nanoparticle aggregates from the lung of rats to the blood and secondary target organs. *Inhal Toxicol.* 2009;21;Suppl 1;Suppl 1(SUPPL. 1):55-60:55-60. doi: 10.1080/08958370902942517, PMID 19558234.
269. Oberdörster G. Lung particle overload: implications for occupational exposures to particles. *Regul Toxicol Pharmacol.* 1995;21(1):123-35. doi: 10.1006/RTPH.1995.1017, PMID 7784625